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Review



# **Biomolecules from Microalgae and Cyanobacteria: Applications and Market Survey**

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**Abstract:** Nowadays, microalgae and cyanobacteria have become a promising and sustainable source of useful products, thanks to their richness in bioactive metabolites of high value (antibiotics, toxins, pharmaceutically active compounds, plant growth regulators, and others). These photoautotroph microorganisms generate biomass using photosynthesis. This review, which distinguishes microalgae and Cyanobacteria, often called blue-green microalgae, aims to present their classification and taxonomic diversity as the ecological niches occupied by them. In addition, the usages of open ponds and photobioreactors to produce various microalgae and Cyanobacteria strains and the high-value bioactive compounds from these microorganisms are summarized. Finally, the numerous commercial applications of these phytoplanktons in different fields, such as food, dietary supplements, feed, cosmetic, and biofuel applications, are reviewed.

Keywords: microalgae; Cyanobacteria; bioactivity; bioconversion

#### 1. Introduction

Worldwide population is rising and resulting in an increased demand for food, biofuel, therapeutic molecules, enzymes, pigments, cosmetic products, and others. Given their broad spectrum of applications, the world encourages the production and cultivation of these organisms. At the academic and industrial levels, many researchers are exploring the engineering tools applied to microalgae processes, including techno-economic analysis and integration and intensification processes [1].

Usually, the term algae does not refer to a proper taxonomic group but to an assembly of aquatic organisms that reduce CO<sub>2</sub> and produce O<sub>2</sub> using photosynthesis, in which chlorophyll a is responsible for 50% of the photosynthic activity on earth. Organisms from this group vary in size, from micrometers to several tens of meters for some algae [2]. The larger ones, macroalgae, are generally visible to the naked eye and are often known as "seaweeds". The smallest ones, microalgae, are unicellular, microscopic algae that can occur separately or in colonies. Within this group, photosynthetic microorganisms can be divided into eukaryotic cells, including the phyla Chlorophyta, Rhodophyta, Glaucophyta, Cryptophyta, Euglenozoa, Cercozoa, Heterokontophyta, Haptophyta, and Miozoa; the prokaryotic ones are also called blue-green microalgae or Cyanobacteria [3,4].

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**Copyright:** © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/licenses/by/4.0/). The classification of this diversity is complex, and the taxonomy is subject to frequent upheavals due to the use of molecular phylogeny techniques. Subsequently, some taxonomists contradict classifying Cyanobacteria as microalgae or bacteria [5,6]. Moreover, their evolution was based on the theory of endosymbiosis, according to the characteristics of their mitochondria and plastids [7]. In addition, microalgae and Cyanobacteria are chlorophyll organisms that thrive in water or in very humid environments. Although most abundant in the waters of seas, lakes, and running waters, it is also found on wet rocks and on land. They can live free or fixed to a support structure [8].

Thanks to their different morphological, physiological, and genetic traits, microalgae and Cyanobacteria have the ability to produce various biomolecules and, notably, biologically active metabolites with great potential in a large range of industrial applications [9].

The present work provides a first part that focuses on the richness of microalgal biomass as a basis for the many biotechnological and industrial applications developed in the second part. It also aims to give readers past, present, and future market data.

#### 2. Microalgae

#### 2.1. What Are Microalgae and Cyanobacteria?

According to phycologists, any organisms containing chlorophyll a (Chl a), devoid of stem, roots, leaves, and flowers, can be considered an alga [10]. These organisms are divided into macro- and microalgae, which are essentially differentiated by their sizes and organelle contents. Macroalgae are benthic, multicellular, and able to reach several meters in length, while microalgae are unicellular microscopic eukaryotic phytoplankton. There is a disagreement among taxonomists in classifying this polyphyletic group of organisms of prokaryotic microalgae, known as Cyanobacteria, as microalgae or bacteria [5,6].

There are obvious distinctions between Cyanobacteria and microalgae—primarily, the presence of membrane-bound structures in eukaryotic cells acquired during evolution through endosymbiosis [7,11]. In fact, Cyanobacteria are primitive microorganisms that bear an intermediate structure between bacteria, characterized by the absence of a distinct membrane-bound nucleus, mitochondria, and chloroplasts, and plants by oxygenic photosynthesis. Secondly, eukaryotes are larger, more complex, more diverse, and can be either unicellular or multicellular [12], while prokaryotes are simple and microscopic single-celled organisms that have neither a distinct nucleus with a membrane nor other specialized organelles. Furthermore, Cyanobacteria contain chlorophyll a as a major photosynthetic pigment, which may be accompanied by carotenoids, xanthophylls, and phycobilisomes (mostly phycoerythrin and phycocyanin) as light collectors. It was estimated that around 10<sup>25</sup> cells of these blue-green microorganisms exist in the oceans, where they are responsible for the achievement of the fixation of photosynthetic carbon and nitrogen [13].

Microalgae are able to grow rapidly and live in a wide range of environmental conditions [14]. They are closely guided by abiotic factors (light, salinity, pH, temperature, and trophic limitation or excess) and biotic ones (trophic relationships) [15]. Microalgae generate themselves using photosynthesis by capturing sunlight, H<sub>2</sub>O, CO<sub>2</sub>, and inorganic nutrients and converting them into biomass, chemical energy, and O<sub>2</sub>, effectuating a full life cycle every few days [16]. They are more efficient at utilizing sunlight than terrestrial plants (10 times greater), with numerous additional technological advantages such as consuming damaging pollutants, having minimal resource requirements, and not competing with agriculture for precious resources [8].

The presence of microalgae is essential since they represent the basis of several food chains. It is estimated that microalgae and Cyanobacteria are around 200,000 to 800,000. Among them, the sole diatoms are estimated to be over 200,000 [17–19]. Currently, more than 164,800 species and infraspecific names exist in the database Algaebase (https://www.algaebase.org/, accessed on 24 november 2020). This appreciable diversity is associated with richness in lipids, proteins, vitamins, pigments, carbohydrates, and other compounds.

#### 2.2. Properties, Taxonomy, and Habitats

Microscopic in size and giant in their properties, eukaryotic microalgae and Cyanobacteria have a huge assortment of colors and different types of cell organization, e.g., unicellular, colonial, and filamentous. They possess different shapes: spherical (*Porphyridium*), droplet (*Chlamydomonas*), filament (*Oscillatoria*), spiral (*Arthrospira/Spirulina*), and even star shape (*Staurastrum*). With a diameter or length of approximately 3 to 10  $\mu$ m and a size ranging from 0.2 to 2  $\mu$ m, they could be classified into picophytoplankton (0.2–2  $\mu$ m), nanoplankton (2–20  $\mu$ m), microplankton (20–200  $\mu$ m), and mesoplankton (200  $\mu$ m– 5 mm) [20].

A microalgal cell contains within its cytoplasm many organelles necessary for sound functioning and metabolism. A single cell may contain one or more chloroplasts. They store their "food" as starch and fats (Table 1). Due to their simple reproduction, their cell growth systems, and their genetic adaptation amenability, they can quickly proliferate by forming phytoplankton "blooms" and survive long term in various harsh environmental conditions (e.g., higher temperature, light intensity, ultraviolet radiations, CO<sub>2</sub> levels, and salinity). They can survive short-term environmental stress by physiological acclimatization triggered by modifying their gene expression. Nevertheless, when environmental stress exceeds their physiological limits, adaptation will be acquired through the selection of several mutations with minor effects [21–23].

Diverse groups of microalgae have generated the remarkable wealth production of primary and secondary metabolites. These compounds, sometimes with high added value, are synthesized by several metabolic pathways such as photosynthesis, carotenogenesis, lipogenesis, and glycerol synthesis. Efficient coordination and regulation of the different metabolic pathways have been demonstrated, depending on the conditions and depth of culture in marine and freshwater environments. These autotrophic organisms are responsible for 40% of photosynthesis on earth [24]. They transform inorganic matter and light energy into chemical energy and different forms of reserve molecules:  $\alpha$ -(1-4)-D-glucans (such as starches)/ $\beta$ -(1-3)-D-glucan, fructans, carbohydrates (such as floridoside) and triacyglycerols [20]. They have three modes of reproduction: sexual by the union of gametes, vegetative by the division of a single cell or by fragmentation of a colony, and, finally, asexual by the production of mobile spores.

Cyanobacteria show enormous biodiversity, and a minority of the described species have been sequenced [25]. They can be either unicellular (*Chroococcus*) or multicellular (*Arthrospira*), with coccoid (*Chroococcus*) or branched filaments (*Anabaena*), singly or grouped into colonies, nearly colorless or intensely pigmented, autotrophic or heterotrophic, psychrophilic or thermophilic, acidophilic or alkalophilic, planktonic or barophilic, dwelling in fresh water or marine environments, including hypersaline environments (salt pans). They are found both free-living and as endosymbionts [26,27,28]. They constitute a phylogenetically coherent group of evolutionarily ancient phototrophic Gram-negative bacteria [13] that show a great deal of variety in size, ranging from small forms called picoplankton (less than 2–3 µm) to larger forms. Cyanobacteria do not contain chloroplast in the cells and keep their energy in reserve in the form of starch or lipids (Table 1). Cyanobacteria are also called blue-green algae due to the presence of Chl a and phycocyanin located in membrane vesicles called thylakoids, which give the cells a bluish color when present in sufficiently high concentrations. Phycoerythrin, a red accessory pigment, can be found as well in these organisms [27].

Cyanobacterial biomass can be estimated at around a thousand million tons (10<sup>15</sup> g) of wet biomass; therefore, they are considered among the most important organisms on Earth. Discovered as putative microfossils 2000–3500 million years ago, they are also considered among the oldest organisms in evolution and are believed to be ultimately responsible for the oxygenation of Earth's atmosphere during the episode of the "Great Oxidation Event" at the end of the Archean Aeon [28,13]. Thought to be amongst the first organisms to colonize the earth, these bacteria are predicted to be the photosynthetic ancestors of chloroplasts in eukaryotes such as plants and algae. In addition, Cyanobacteria

have the ability to perform photosynthesis, aerobic respiration, and CO<sub>2</sub> and (sometimes) nitrogen fixation. They perform oxygenic photosynthesis like plants and algae, utilizing solar energy and water to generate chemical energy (ATP) and reducing power (NADPH) to fix atmospheric CO<sub>2</sub> and synthesize carbohydrates [13,29]. Vermaas (2001) noted that photosynthesis and respiration in Cyanobacteria can take place simultaneously in the same compartment, represented by the thylakoid membrane, thus making this combination a bit unique for them [28]. During photosynthesis, Cyanobacteria use water as an electron donor and produce oxygen as a coproduct. Some filamentous Cyanophycae are characterized by the presence of specialized cells called heterocysts, which allow the fixation of atmospheric N<sub>2</sub> in anaeroby and, therefore, give Cyanobacteria the possibility of growth in inorganic nitrogen-depleted environments [30]. In the dark or under anoxic conditions, Cyanobacteria can perform fermentations for energy generation [31]. It may be due to this flexibility of metabolic pathways that these organisms are able to grow under a wide range of conditions.

Unlike eukaryotic organisms, the Cyanobacteria reproduce by vegetative, asexual processes, and sexual reproduction is completely absent. The commonest mode of reproduction among them is undergoing a binary split. Moreover, there have certain specialized structures such as akinetes, hormogonia, hormocysts, and spores, which are partly involved in the process of reproduction [25].

Microalgae and Cyanobacteria are extremely heterogeneous groups gathered around a common characteristic: oxygenic photosynthesis [32]. Among these groups, which exhibit significant taxonomic diversity, the evolution of microalgae was based on the theory of endosymbiosis, according to the characteristics of their mitochondria and plastids [7]. It was proposed that mitochondria originated from the endosymbiosis of  $\alpha$ -proteobacteria by a protoeukaryote, which thereby developed the complex eukaryote cell. The chloroplast, in turn, takes its origin through the primary endosymbiosis of a Cyanobacterium by this heterotrophic eukaryote, also called "protist". Endosymbiosis may be the origin of the Plantae kingdom, with primary plastids (formed with only two membranes) comprising phyla of Chlorophyta, Glaucophyta, and Rhodophyta. Therefore, photosynthetic membranes of plastids of red algae strongly evoke the thylakoids of photosynthetic bacteria. In addition, the pigment equipment of Rhodophyta (Chl a) and phycobiliproteins resemble that of Cyanobacteria. Other endosymbioses have arisen during the evolution of microalgae, giving rise to organisms with secondary and even tertiary plastids. However, uncertainties relating to the order of appearances and the independence of events still persist. The secondary endosymbiosis took place between a red algae and a non-photosynthetic host cell and gave rise to the ancestor of all Chromalveolata supergroup: Cryptophyta, Haptophyta, Ochrophyta (or Heterokontophyta), and Miozoa. This is the most controversial hypothesis in the literature postulated in 1999 by Cavalier-Smith under the name of "The Chromalveolate Hypothesis" [33]. By a similar process, Euglenophyta, Cercozoa, and green Miozoa acquired their plastids from ancestral Chlorophyta [34-36]. Nevertheless, the work described by Baurain et al. [37] has shown that the differences observed within these groups cannot be fully explained by this hypothesis. For this reason, another theory called "The Independent Acquisition Hypothesis" explains the origin of Chromalveolates by a series of endosymbioses.

Therefore, the classification of microalgae is very complex and ambiguous. As mentioned above, it could be based on various different criteria, such as the organization of photosynthetic membranes, but also other criteria such as the chemical nature of storage products and, most commonly, their pigmentation [38,39]. In addition to the morphological basis, Reviers Bruno de [40] and Groendahl et al. [41] have proposed a classification of microalgae and Cyanobacteria by means of molecular biology techniques based on the sequence alignments of genes encoding the 16S and 23SrRNAs for Cyanobacteria and 18S and 28SrRNAs for microalgae in order to establish family ties between these microorganisms. Other methods, such as the flow cytometer, ensuring semi-automated or fully automated classification, have been recently employed to identify and classify algal species [42]. Currently, there is no specific classification used by the taxonomists since the taxonomic organization may change over the years [43,44].

As early as their discovery, different kinds of classification of algae were proposed. For example, [5,45,46] classified microalgae and Cyanobacteria into 11 and 10 classes, respectively. Contrary to the classification of Fritsh, where the 11th class consists of Cyanobacteria, Chapman employed the Procaryota-Eucaryota concept in the classification of microalgae. Table 2 summarizes this classification by implementing the main characteristics of the different classes. Table 3 presents the classification of microalgae and Cyanobacteria are a mixture of different kingdoms: Eubacteria, Protozoa, Chromists, and Plants. This classification is based on data from Ruggiero et al., published in 2015 [47], who detailed the distribution of all living organisms.

Microalgae and Cyanobacteria are ubiquitous, and they occupy various types of ecological niches. Most of them thrive in fresh, saline, or brackish waters. Some of them are also capable of forming a sticky coating in the form of "biofouling" biofilms on various terrestrial habitats such as soil and rocks, tree trunks, or even architectural buildings [8,48] (Table 2). There are many parameters that influence the habitat of the microalgae and Cyanobacteria, mainly the salinity and the availability of substrates. The temperature and the quantity and quality of light are also among the principal factors influencing their habitat. At sea level, these two factors depend on the movement of the tides and the seacoast depth. Indeed, algae spread according to the sea depth, influencing the availability of the wavelengths that can be absorbed by their pigments. Some authors define this mechanism by the chromatic adaptation in algae [49]. The latter can be considered as a change in the absorption spectrum of an organism, activated by different spectral compositions and intensities of incident solar energy [50]. Green microalgae are present on the surface, adapted to red wavelengths. Brown microalgae are found between green and red microalgae, which are found in the deeper strata. In this depth, they exploit the green and blue wavelengths. However, this parameter does not prevent us from sometimes finding green microalgae living in deeper depths and red microalgae on the surface. This diversity of color allows microalgae to easily absorb the dominant rays in the solar radiation at various depths [51].

Table 1. Reserve molecules from photosynthesis products [20].

Reserve Form	Microalgae
Floridian starch (amylopectin subunits)	Red microalgae
Myxophyceal starch	Plus groop microslass
(amylopectin or glycogen-like subunits)	Blue-green microalgae
Cross-linked amylose-amylopectin starch	
Fructans (inulin-like fructose oligosaccharides)	Green microalgae
$\alpha$ -(1-4)-glucan	Cryptophyceae and Dinoflagellate
Oil or chrysolaminarin	Chrysophyceae
Chrysolaminarin	Bacillariophyceae
Denomy long $(1, 2)$ D alugan	Euglenophyta and
Paramyion: p-(1-5)-D-glucan	Prymnesionhyceae

Table 2. Classification of microalgae and Cyanobacteria [20,45,46].

Phylum	Class	Pigments	Reserve	Flagellation	Reproduction	Cell Envelope	Habitat
Cyanobacteri a	Cyanophyceae	Chl a, β-carotene, flavacene, Echinenone	Starch (granule) and glycogen	Unflagellated	Simple division, non-motile endospore, vegetative fragmentation	Lack of cellulose	Marine Freshwater Terrestrial

		isozea-, zea-,			No sexual		
		oscillaxanthin APC, C-PC, C-PE			reproduction		
Chlorophyta	Chlorophyceae Trebouxiophyc eae Ulvophyceae	Chl a, b, $\alpha$ and $\beta$ - carotene, lutein, zea-, viola-, loro- and neoxanthin	Starch Oil	From two to four flagella (mobile), isokont	Isogamy, anisogamy, oogamy Motile zoospore, non-motile spore	Cellulose+ mannans+ xylans Sitosterol Sometimes calcified	Marine Freshwater
Euglenophyt a	Euglenophycea e	Some colorless Chl a, b, diadinoxanthin	Paramylon Ergosterol	One or two flagella, one of which contains cilia and the second being reduced	Longitudinal division	No cell membrane Flexible periplast or pellicle	Marine Freshwater Terrestrial
Heterokonto phyta/ Ochrophyta	Xanthophyceae Eustigmatophy ceae	Chl a and c, β- carotene, heteroxanthin, diadinoxanthin (++)	Oil Leucosin Ergosterol	Two heterokont flagella and sometimes a single flagellum	Zoospore, aplanospores, statospores Isogamy, oogamy (Vaucheria)	Rich in pectic compounds Silicification during the rest period	Marine Freshwater Terrestrial
Miozoa	Dinophyceae	Chl a, c, β- carotene, diadinoxanthin, dinoxanthin, peridinins	Starch Lipids	Lateral biflagella	Zoospore Longitudinal oblique division Aniso or isogamy	With or without cellulose	Marine Freshwater
Heterokonto phyta/ Ochrophyta	Chrysophycea	Chl a, c, β-carot, Fuco-, Diato-, diadinoxanthin	Chrysolamina rin Fucosterol Porifasterol	Two heterokont flagella	Zoospore, statospore	Less cellulose with silicification or calcification of the membrane	Marine Freshwater
Haptophyta	Coccolithophyc eae Pavlovophycea e Rappephyceae	Chl a, c, β-carot, Fuco-, Diato-, diadinoxanthin	Chrysolamina rin Fucosterol Porifasterol	Two isokont flagella + haptonema	Zoospore, statospore	Less cellulose with silicification or calcification of the membrane	Marine Freshwater
Bacillariophy ta (Diatoms)	Bacillariophyce ae	Chl a, c, β-carot, Fuco-, Diato-, diadinoxanthin	Chrysolamina rin Oil	Without flagellum except for male gametes	Cell division Oogamie (centered diatoms) Amoeboid isogamy (pinnate diatoms)	Silica + pectic compounds	Marine Freshwater Terrestrial
Cryptophyta	Cryptophyceae	Chl a, c, Biliproteins, α- carot, Allo-, Croco-, Monado- xanthin	Starch (granule) Oil Carbohydrate s	Two unequal flagella except for <i>Bjornbergiella</i> and <i>Tetragonidium</i>	Zoospores	Proteinaceous periplast	Marine Freshwater
Heterokonto phyta/ Ochrophyta	Phaeophyceae	Chl a, c, β-carot, violaxanthin Fucoxanthin (++)	Mannitol Laminarin Lipids	Two heterokont flagella	Monospores, tetraspores, non- motile aplanospores	Cellulose + alginic acid + fucoidin	Marine Freshwater Terrestrial

					From isogamy to oogamy		
Rhodophyta	Porphyridioph yceae Bangiophyceae Florideophycea e	APC, C-PC, R- PC, B-PE, R-PE, phytochrome,α and βcarot, Chl a and d, zeaxanthin, lutein	Starch, floridoside, mannoglycer ate, sulfated polysaccharid es	No flagella	Oogamy complex	Carbohydrate s (xylose, cellulose, galactane)	Marine Freshwater Terrestrial

**Table 3.** Classification of microalgae and Cyanobacteria according to their phyla (based on data from Ruggiero et al. [47] and https://www.algaebase.org/, accessed on 24 November 2020).

	Kingd	om				Phylu	um			Class	
	0					<u> </u>			Class C	Class <b>Cvanophyceae</b>	
Kingdon	n	Subk	ingdom	m . Phyli		Phylum Cyanobacteria [= Cyanophyta]		[= Phycobacteria]			
EUBACTE	KIA	Negi	bacteria	2	5		2 9	1,5,5	Class Gloed	bacterophyceae	
						C 1	1 1 6	-1 1	Class I	Diplonemea	
						Subp	hylum (	Jycomonada	Class K	inetoplastea	
					-	Subph	ylum	Infraphylum	Class E	ntosiphonea	
				DL L		Euglei	noida	Entosiphona	Class Eu	glenophyceae	
	C		GDOM EUCLENO	Euclon	1m -	Subp	ohylum	Symbiontida	Class I	Postgaardea	
	S N	IGDO	ZOA	Euglen	020a -	т			Superclass Spirocuta	Class Peranemea	
T/1 1	E					Inf	raphylu	im Dipilida	Superclass	Class Ploeotarea	
Kingdon	n = E(	JZOA							Rigimonada	Class Stavomonadea	
PROTOZO	JA		INFRAKIN		1	Su	ıbphyluı	m Eolouka	Clas	s Jakobea	
			GDOM	Phylum Lo	oukozoa-	Sul	ophylun	n Neolouka	Class Ma	lawimonadea	
			EXCAVAT Phylum A Metamona		ım mada	Sul	ophylun	n Trichozoa	Superclass Eopharyngia	Class Trepomonadea	
			Phylum		Sub	phylum	Choanofila	Class Cho	oanoflagellatea		
				Choan	Choanozoa Subphylum		hylum C	Cristidiscoidea	Class C	ristidiscoidia	
					Pł	nylum I	Picozoa		Class P	icomonadea	
				Phylum P	ercolozoa	a Su	bphylur	n Tetramitia	Class H	eterolobosea	
					Phyl	um Cry	ptophyt	ta	Class Cı	yptophyceae	
	Subkir	adom	Uacrobia				Class Coccolithophyceae				
	SUDKI	iguoin	i i iaci obia	Phylum <b>Haptophyta</b>			[= Prymnesiophyceae]				
				-				Class Pavlovophyceae			
					Phyl	Phylum Subphylum		Class Ciliatea			
					Phylum		ora Intramacronucleata		Class Litostomatea		
				-	Chiop	nora	intrai	macronucleata	Class S	Spirotrichea	
								Infranhylum	Superclass	Class	
								Anicomplexa	Apicomonada	Apicomonadea	
Kingdom							_	присотрисли	Supercl	ass Sporozoa	
CHROMIST							Subph		_	Class Dinophyceae	
A			INFRAKIN	Superphy			ylum		Superclass -	Class Ellobiopsea	
	SUBKIN	NGDO	GDOM	lum	Phylum	Miozoa	Myzo	Infraphylum	Dinoflagellata -	Class Noctilucea	
	M HAI	ROSA	HALVARI	Alveolata			zoa	Dinozoa		Class Oxyrrhea	
			А							Class Syndinea	
									Superclass Perkinsozoa	Class Perkinsea	
				_		-		ubphylum otalveolata	Class C	Colponemea	
				-		Phylin	m Ochro	mbuta	Class Bo	olidophyceae	
					r_	rnyiui Hotoro	h Ocnře kontopi	priyta pyta n n l	Class Chry	somerophyceae	
						[= Heterokontopnyta p.p.]			Class Chrysophyceae		

					Class Eustigmatophyceae
					Class Pelagophyceae
					Class Dictyochophyceae
					Class Phaeophyceae
					Class Phaeothamniophyceae
					[= Aurophyceae]
					Class Picophagophyceae
					Class Pinguiophyceae
					Class Raphidophyceae
					(= Chloromonadophyta)
					Class Schizocladiophyceae
					Class Synchromophyceae
					Class Synurophyceae
					Class Xanthophyceae
					Class Bikosea
					Class Blastocystea
			Dhee	lum Diarma	Class Nanomonadea
			ГПу	Tulli Digyra	Class Opalinea
					Class Labyrinthulea
					Class Placididea [=Placidiophyceae]
					Class Chlorarachniophyceae
					[Chlorarachnea]
			Phylum C	ercozoa	Class Thecofilosea
	KIIZ/	ANIA			Class Filosa
					Class Imbricatea
			Phylum Bacillarionhyta	Subphylum	Class Bacillariophyceae
			r nyiuni bacmanopnyia	Bacillariophytina	[= Diatomeae]
		-	Phylum <b>Gla</b>	ucophyta	Class Glaucophyceae
				Subphylum	Class Cyanidiophyceae
			_	Cyanidiophytina	Clubb Cyulifulophyccuc
	SUBKIN	GDOM		Subphylum	Class Bangiophyceae
	BILIPH	ТА	Phylum <b>Rhodophyta</b> –	Eurhodophytina	Class Florideophyceae
	DILIIII		i nytum <b>mouopnyu</b>		Class Compsopogonophyceae
				Subphylum	Class Porphyridiophyceae
				Proteorhodophytina	Class Rhodellophyceae
					Class Stylonematophyceae
					Class Chlorodendrophyceae
Kingdom				Subphylum	Class Chlorophyceae
PLANTAF		INFRAKIN		Chlorophytina	Class Pedinophyceae
		GDOM	Phylum Chlorophyta	emorophymu	Class Trebouxiophyceae
		CHLOROP			Class Ulvophyceae
	SUBKINGDC	) HYTA		Subphylum	Class Nephrophyceae
	Μ			Prasipophytina	Class Mamiellophyceae
	VIRIDIPLAN	1		пазнорнуша	Class Pyramimonadophyceae
	TAE				Class Charophyceae
		INFRAKIN			Class Chlorokybophyceae
		GDOM	Superphylum	Charophyta:	Class Coleochaetophyceae
		STREPTOP	Phylum <b>Ch</b>	arophyta	Class Conjugatophyceae
		HYTA			Class Klebsormidiophyceae
			-		Class Mesostigmatonhyceae

## 2.3. Compositions and Technologies for Photoautotrophic Production of Microalgae and Cyanobacteria

Different morphological, physiological, and genetic traits confer on microalgae the ability to produce different biologically active metabolites. This implies biochemical diversity, which becomes the basis for many biotechnological and industrial applications. In fact, microalgae biomass contains important primary metabolites such as lipids, proteins, and carbohydrates, with food and nutraceutical applications. Nevertheless, bioactive compounds also can be synthesized from secondary metabolism with cosmetics and pharmaceutical aims [24,52]. Similarly, Cyanobacteria from marine and freshwater sources are known to produce a variety of bioactive compounds (secondary metabolites) that are derived from mixed biosynthetic pathways endowed with diverse biological activities, including antiviral, antibacterial, antifungal, antimalarial, antitumoral, and anti-inflammatory properties [53–55].

#### 2.3.1. Amino acids, peptides, and proteins

Microalgae and Cyanobacteria are considered to be rich and natural sources of vegetable proteins, mainly used as functional ingredients [56]. Indeed, they can produce proteins from 6% to 80% of their dry mass, depending on the species [57], e.g., *Chlorella vulgaris* (Chlorophyta) (51–58% DW), *Arthrospira maxima* (Cyanobacteria) (60–71% DW), *Isochrysis galbana* (Haptophyta) (50–56% DW), or *Porphyridium purpureum* (Rhodophyta) (28–39% DW). Moreover, bioactive peptides are convenient high added-value compounds for humans due to their therapeutic potential in the treatment of various diseases, in addition to providing nutritional benefits [56].

Besides their high protein content, microalgae and Cyanobacteria are also known for their high amino acid (Aas) content, in which the more abundant are aspartate and glutamate [39]. According to Sousa et al. [58], microalgae contain essential amino acids compared to egg and soybean. Microalgae and Cyanobacteria, such as those belonging to the species *Nostoc sphaeroides, Arthrospira platensis* (Cyanobacteria), and *Chlorella vulgaris* (Chlorophyta), show a large variability in protein digestibility, with values of 82%, 81%, and 76%, respectively [59]. Aminograms of *Arthrospira* extracts with high protein digestibility (83–90%) have highlighted all essential amino acids, particularly, leucine, valine, and isoleucine [60]. The increased digestibility of Cyanobacteria is explained by their thinner cell wall. Kose et al. [61] reported that other species, such as *Nannochloropsis oceanica* (Eustigmatophyceae) and *Chlorella sorokiniana* (Chlorophyta), may exhibit low digestibility rates of 50% and 55%, respectively, due to the rigidity and thickness of their cell walls, which are considered digestibility barriers.

Cyanobacteria represent an inexhaustible source of proteins and various compounds with promising antiviral activities. Cyanovirin-N is an 11 KDa protein produced by *Nostoc ellipsosporum* that displays viricidal activity against several viruses and, notably, the human immunodeficiency virus (HIV). Other microbial metabolites were equally reported to exhibit antiviral activities, such as Scytovirin, a single 95-amino acids protein, which was isolated from the Cyanobacteria *Scytonema varium*, with potent antiviral activity against HIV.

#### 2.3.2. Photosynthetic pigments

Photosynthetic pigments of microalgae and Cyanobacteria are composed of chlorophylls, carotenoids, and phycobiliproteins (phycoerythrin, phycocyanin, and allophycocyanin). These three groups confer on microalgae their green, yellow/orange, and red/blue colors, respectively [62]. Natural pigments derived from microalgae and Cyanobacteria are of great importance for the food, cosmetic, and pharmaceutical industries. They are not only responsible for capturing solar energy for photosynthesis but also play a role in photoprotective processes and display antioxidant, anticarcinogen, anti-inflammatory, antiobesity, antiangiogenic, and neuroprotective activities [24,63]. Chlorophylls, which represent 0.5% to 1.5% of dry matter of microalgae and Cyanobacteria, are green, lipid-soluble pigments that play a major role in photosynthesis. They are tetra-pyrrolic macrocycles that are metalized by the presence of a complexed magnesium atom in their center and substituted by a phytyl chain, and they may be found in algae, higher plants, and Cyanobacteria.

Chlorophyll a (Chl a) is present in all algae and has a major peak of light absorption at  $683 \pm 20$  nm due to the activity of photosystem II (PSII) and a second one at 700 to 750 nm due to both PSII and PSI [64]. This photosynthetic pigment is accompanied by accessory ones, including chlorophylls b, cl, c2 and d, phycobiliproteins, and carotenoids, which absorb at different wavelengths, thus expanding the spectrum of light-harvesting [20]. Chlorophyll b, c, and d are variants of the Chl a found in green algae, brown algae, and Rhodophyta, respectively [65].

Present as orange, red, or yellow colors, carotenoids (0.1% to 0.2% of dry matter) are responsible not only for the primary coloration of some algal groups but also for the mechanism of photosynthesis [66]. Carotenoids are chemically subdivided into deoxygenated carotenes, principally  $\beta$ -carotene, which is the most widely known of the carotenoids because of its antioxidant property as well as its dyeing power. Known as provitamin A (precursor of vitamin A),  $\beta$ -carotene is mainly produced by the hypersaline species *Du*naliella salina (Chlorophyta), which can accumulate up to 98.5% of this compound in relation to its total carotenoids and about 13% of its dry biomass [39]. The second major group is the red oxygenated pigment. The freshwater chlorophyceae Haematococcus lacustris (Chlorophyta) is considered the primary source of astaxanthin as it was found to have the highest production on the industrial scale and can accumulate up to 81% of astaxanthin of its total carotenoids and about 7% of its dry weight [67-69]. This pigment is commercially produced from *Chlorella zofingiensis*, *Chlorococcum* sp., and *Scenedesmus* sp. Astaxanthinis is described in the literature as having a higher antioxidant activity, health benefits, and dyeing power compared with those of  $\alpha$ - and  $\beta$ -carotenes, lutein and lycopene, canthaxanthin, and tocopherols [24,39,70]. Phycobiliproteins are colored (blue or red pigments) and water-soluble pigments organized in supramolecular complexes, called phycobilisomes, mainly present in Cyanophyta, Rhodophyta, Glaucophyta, and Cryptophyta. These complexes consist of 85% phycobiliproteins, of which 4 classes can be distinguished: allophycocyanin at the center of the complex, phycocyanin, followed by phycoerythrin or phycoerythrocyanin at the level of the radiary projections. The characteristics of these pigments have been detailed by Thomas (1989) [71]. They absorb light in the visible region of 450-650 nm. They are used as natural colorants in the food, cosmetic, and pharmaceutical industries as substitutes for synthetic dyes. The species most commonly used for the production of phycobiliproteins are Porphyridium sp. (Rhodophyta), Arthrospira/Spirulina sp., and Aphanizomenon flosaquae (Cyanobacteria) [39,70].

#### 2.3.3. Lipids and fatty acids

Lipids from microalgae and Cyanobacteria are classified into two categories, which are: (i) storage lipids, mainly triglycerides that are non-polar lipids produced by photosynthesis and stored in cells and (ii) structural lipids, such as phospholipids and sterols, which are plastid and cell membrane polar lipids [72]. Microalgae are able to accumulate up to 20–50% of their dry weight in lipids, and this content can reach 80% in response to changes in nutritional and environmental conditions (nitrogen concentration, salinity, or temperature). Furthermore, microalgae contain very rich and diverse fatty acid profiles. The latter are the primary metabolites with 14–20 carbons, belonging to the class of storage lipids mainly used for applications focusing on the production of biodiesel. Saturated and polyunsaturated (PUFAs) fatty acids with more than 20 carbon atoms are used in food as health food supplements and in the pharmaceutical industry for docosahexaenoic (DHA) and eicosapentaenoic (EPA) acids. In addition, the lipid composition of microalgae is very rich in essential PUFAs such as  $\omega_3$ , in particular eicosapentaenoic ( $\omega_3$ , EPA, C20: 5) and docosahexaenoic ( $\omega_3$ , DHA, C22: 6) and  $\omega$ -6 fatty acids. The fatty acid composition of

microalgae and Cyanobacteria depends essentially on the species. The Chlorella species are known for their high content of DHA and EPA (3.24% and 8.9% of total fatty acid methyl esters (FAMEs), respectively, in addition to saturated fatty acids). The fatty acids extracted from Chlorella vulgaris are mainly composed of oleic, palmitic, and linolenic acids. The green algae *Haematococcus* are characterized by its short-chain fatty acids profile with antimicrobial activity, while the Cyanobacterium Arthrospira/Spirulina represents a good source of y-linolenic acid, sterols, and fatty acids such as palmitic, lauric, oleic acids, and DHA. Diatoms are considered to be a promising source of EPA (15% to 30% of total fatty acids) but not DHA, contrary to dinoflagellates, which have a high potential for use in the commercial production of this compound, which ranges from 12% to 51% of total fatty acids [39,72–74]. Different antimicrobial, antiviral, anticancer, and antiprotozoal compounds have been reported from Cyanobacteria. Singh et al. [75,76] mentioned that lipopeptides from Cyanobacteria have cytotoxic (41%), antitumor (13%), antiviral (4%), antibiotic (12%) activities, and the remaining 18% include antimalarial and antimycoticsone activities. Some Cyanobacteria also have the potential to produce intracellular and extracellular bioactive compounds with potent antibacterial and antifungal activities against many fungi and Gram-positive and Gram-negative bacteria. However, some of the antimicrobial compounds isolated from Cyanobacteria exhibit cytotoxicity and may be ineffective for use in pharmaceuticals [77].

Cyanobacteria are a source of value-added products such as isoprene, which is an unsaturated, energy-rich hydrocarbon widely used as a biofuel by the chemical industry. Polyhydroxyalkanoates are biodegradable and biocompatible polyesters produced in nature by numerous Cyanobacteria, including *Aphanothece* sp., *Oscillatoria limosa, Arthrospira/Spirulina*, and the thermophilic strain *Synechococcus* sp. MA19. This polymer has gained immense attention for industrial applications, such as replacements for conventional non-biodegradable plastics, as well as in the biomedical or pharmaceutical field [78]. Additionally, Cyanobacteria produce a broad spectrum of compounds with anticancer activity. These compounds show potential benefits against cancer, including cell cycle arrest by inhibition of cell growth using apoptotic processes and mitochondrial dysfunctions. For example, some compound extracts from the genus *Arthrospira/Spirulina* resulted in the inhibition of homogenous leukoplakia. Others repressed chemically induced carcinogenesis in model hamster buccal pouches [76].

#### 2.3.4. Carbohydrates

Microalgae have been suggested as a promising source of carbohydrates, which are located not only inside the chloroplast but also in the cytosol and extracellular compartments. These mono-, oligo-, and polysaccharides are produced during photosynthesis, and their contents can reach 50% of the dry weight of some microalgae, depending on the species and the cultivation and environmental conditions [39,78]. Cyanobacteria and red microalgae store, respectively, glycogen ( $\alpha$ -(1,4-1,6)-D-glucan) and floridean starch (hybrid between starch and glycogen), while green microalgae accumulate amylopectin-like polysaccharides [39,78] (Table 3).

Carbohydrates are classified as simple or complex compounds. The former contains just one or two sugars and are called monosaccharides, while the latter has two or more sugars, called di-, oligo-, and polysaccharides. The latter are polymers of carbohydrates (glycans), and they are extracted from many species and genera of microalgae, mainly from *Chlorella* sp., *Tetraselmis* sp. (Chlorophyta), *Isochrysis* sp. (Haptophyta), *Porphyridium purpureum*, and *Dixoniella grisea* (Rhodophyta) [65,78,79]. Some microalgae also produce, depending on species, structural polysaccharides qualified as fibrillary, which participate in the formation of the cell wall and could be used in pharmaceutical, cosmetic, and food industries. They also have the potential of applications such as thickening or gelling agents as well as bioactive compounds, citing antiviral, anti-inflammatory, anticoagulant and antithrombotic, antitumor, and immunomodulatory activities [80]. The second class is known as energy reserve polysaccharides and exocellular polysaccharides (EPSs),

mainly used for biofuel production. Finally, polysaccharides can also be involved in cell communication [40]. Carbohydrates such as Nostoflan, cyclic depsipeptides, microvirin, ichthyopeptins A and B, and many others compound extracts from Cyanobacteria also show strong antiviral properties [13,76,77].

#### 2.3.5. Other compounds

Microalgae have also been recognized as an excellent source of vitamins and minerals, some of them having antioxidant properties such as the hydrosoluble vitamins (C, B1, B2, B3, B5, B6, B9, and B12) and fat-soluble ones (provitamin A and vitamins E, D, and K). They also contain nicotinate, biotin, folic acid, and pantothenic acid as well as macrominerals (Na, K, Ca, and Mg) and microminerals (Fe, Zn, Mn, and Cu) [81–84]. Parts of prokaryotic microalgae are known for their large-scale production of vitamins of commercial interest, such as vitamins B and E, and *Spirulina* is part of this. In fact, this Cyanobacterium is known to be a rich source of vitamin B12,  $\beta$ -carotene, thiamin, and riboflavin [77].

Microalgal biomass also contains an abundance of a wide variety of secondary bioactive substances. Within this bioactive diversity, phenolic compounds constitute one of the most numerous and abundant groups of phytochemicals, which exhibit a wide range of biological properties, such as antioxidant, anti-inflammatory, anticancer, antiallergic, antidiabetes, antiaging, and antimicrobial properties [39]. Moreover, microalgae and Cyanobacteria have great potential for synthesizing enzymes such as cellulases, galactosidases, amylases, proteases, lipases, phytases, laccases, phosphatases, and antioxidant enzymes, with high potential for several applications [77,84]. Microalgae also produce phytohormones such as auxin, abscisic acid, cytokinin, ethylene, gibberellins, polyamines, and jasmonides, mainly known to regulate the main physiological processes of plants, terrestrial or aquatic organisms [79,85,86], phytosterols [39], and others.

2.3.6. Technologies for photoautotrophic production of microalgae and Cyanobacterial biomass

Given the great interests of algae and their broad spectrum of applications, the world is invited to push and encourage the production and cultivation of these organisms. Furthermore, until 2011, global algal exploitation remained marginal, with 15 million tons/year of macroalgae and 4 billion tons/year of terrestrial plants against 10,000 tons/year of microalgae [38]. New estimates show that between 2016 and 2018, the production of microalgae, including Cyanobacteria, bounded by 19,000 and 20,000 tons per year. By 2024, their expected commercial production will be 27,500 tons per year, with a market value of USD 1.1 billion [87]. The challenge nowadays will be to improve current cultivation methods to scale up this production. Macroalgae are cultivated in a natural marine environment, while microalgae and Cyanobacteria are cultivated in reactors, either in photoautotrophy (raceways or photobioreactors) or in heterotrophy [38]. These two production systems remain, in fact, the most practical, as mentioned next.

Open ponds are the most exploited processes for the industrial culture of microalgae and Cyanobacteria due to their ease of use and low cost. These systems generally use natural light and vary according to their shape, the type of materials used, as well as the mixing system. They are mainly classified into two types: circular and "raceway" ponds [88]. The first open ponds, qualified as natural basins, were without agitation and generally had low cell productivity. In order to overcome this problem, mixing systems with a circular shape and centrally pivoted rotating agitator were then put in place for the production of biomass, animal feed (aquaculture), and dietary supplements based on *Arthrospira/Spirulina* (Cyanobacteria) or *Chlorella* (Chlorophyta) [89]. Open raceway tanks later appeared, characterized by an elongated shape, and they are currently the most widely used artificial systems for microalgal cultivation. This design consists of a closed-loop flow channel with a typical culture depth of approximately 12 to 50 cm, in which the mixing and circulation of the microalgae culture are promoted by paddle wheels [90]. Light intensity is considered among the parameters that strongly influence the growth rates of microalgae in outdoor cultures. Moreover, with such a mixing system, microalgae have better access to sunlight, which can come only from the top. Besides light, dissolved CO<sub>2</sub> concentration is another important parameter for productivity in these systems. Indeed, atmospheric CO<sub>2</sub> uptake by microalgae and Cyanobacteria is not sufficient for optimal growth. In open reactors, supplementary to light and CO<sub>2</sub> availabilities, factors such as temperature and the mixing and availability of nutrients are involved in reaching optimal growth and are strongly affected by the culture depth [90].

Photobioreactors (PBRs) are closed systems that consist either of horizontally or vertically arranged tubes or vertically arranged panels, wherein microalgae or Cyanobacteria are grown. This technology offers better control of all culture parameters such as pH, temperature, oxygen, and agitation. They are well suited for microalgae sensitive to contamination, such as extremophilic microalgae. Depending on location and metabolite production, they may or may not be artificially illuminated. During the last decades, the marketing of industrial PBRs with different designs and efficiencies has increased the yields of biomass production. Several shapes are currently available, such as sleeves, tubular reactors, annular reactors, flat reactors, rigid panel type reactors, and "airlift" type reactors (bubble column) [88,91].

Microalgae and Cyanobacteria can be harvested by mechanical, chemical, biological, and electrical methods. They are based on the principles of separation by the density difference or cells sizes. For the first strategy, we cite sedimentation, flocculation-settling, flotation, and centrifugation techniques, while the second one is based on membrane processes such as micro- or ultrafiltration [38]. The choice of the used technique depends on the chosen microalgae morphology, specifications of the final product, the ability to reuse the culture medium, as well as the density and size of the microalgal cell [92]. Likewise, to improve harvesting efficiencies, the process should not only be effective for the majority of microalgae and allow for a higher biomass concentration but should also require moderate costs of operation energy and maintenance. Therefore, the harvesting process cost is also very important as it usually accounts for about 20–30% of the total microalgal production cost. Thus, it is better to apply a combination of reliable and low-cost processes such as coagulation/flocculation by mechanical, chemical, and biological methods, which are the most used to reduce operation and maintenance charges. Filtration-, centrifugation-, and electrical-based processes remain the least used due to their high energy and equipment costs.

#### 3. Current Uses of Microalgae and Cyanobacteria in Industry

Thanks to their important and diverse high-added-value contents, industrial microalgae and Cyanobacteria production has increased in these last years. By 2050, the world population will exceed 9 billion, which will compel humanity to find and use new sources for food. In fact, 70% more food will be needed, according to Food and Agriculture Organization (FAO) organization (www.fao.org, accessed on 10 November 2020). In addition to this increase, demand for food and fuel production will be affected by climate change and the decline of ordinary resources. Microalgae and Cyanobacteria could represent a natural alternative for the resolution of these problems. The National Center for Biotechnology Information expects that, by 2050, algae could provide 18% of protein sources (www.fortunebusinessinsights.com, accessed on 7 December 2020). Currently, algae production is rising as their compound annual growth rate (CAGR) will increase by 4.3% in the period 2020–2027, according to www.strategyr.com, accessed on 11 November 2020). It was estimated that in 2020, dyeing agents are at the head of the list of microalgae products on the market, with USD 800 million, followed by pharmaceuticals/chemicals (USD 500 million), nutraceuticals (USD 300 million), and, finally, cosmetics (USD 30 million) [39].

Microalgae and Cyanobacteria provide to the consumer an unrepeatable source of high-value compounds. Therefore, they are mostly commercialized as health products, diet supplements, functional food, and nutraceuticals in capsules, tablets, powders, pills, and syrups forms [93]. Table 4 presents different industries producing food supplements and nutraceuticals from microalgae and Cyanobacteria. *Chlorella* (Chlorophyta) and *Ar*-*throspira/Spirulina* (Cyanobacteria) are the most popular genus used as diet supplements. They constitute 80% of the global microalgae food market, thanks to their composition (www.fortunebusinessinsights.com, accessed on 14 February 2021).

Spirulina is known as an interesting source of proteins, phycobiliproteins, vitamin B12, minerals,  $\gamma$ -linolenic acid, and others. Its production and consumption are in continuous growth. In a previous assessment, the global production of dried Spirulina was 12,000 tons.year<sup>-1</sup>. In 2014, FAO estimated a Spirulina global production of 86,000 tons [94]. According to persistence market research, over 128,000 tons of Spirulina were globally consumed in 2016 (www.persistencemarketresearch.com/, accessed on 14 February 2021). Its global market size was USD 348 million in 2018 and is projected to reach USD 779 million by 2026, growing at a CAGR of 10.5% from 2019 to 2026 (www.alliedmarketresearch.com, accessed on 14 February 2021). In their report, Persistence Market Research affirmed that the powder form of Spirulina was the highest form used as, in 2016, this segment represented 73.5%, and it is expected that its consumption will record a CAGR of 10.2% in the period 2019–2026. In second place, the demand for tablets and capsules will increase to a CAGR of 8.8% in the same period. By 2026, more than 250,000 tons of Spirulina will be consumed globally in the production of nutraceuticals (www.persistencemarketresearch.com/, accessed on 14 February 2021). The aqueous extract of this bluegreen microalgae, rich in natural phycocyanin colorant, was authorized by the Food and Drug Administration (FDA) at a maximum level of 2 mg.kg<sup>-1</sup> to be safely used in coloring confections and in dietary supplement tablets and capsules (www.accessdata.fda.gov, accessed on 10 December 2020). Different companies are currently producing phycobiliprotein around the world, such as Parry Nutraceuticals (Pudukkottai District, Unaiyur, Tamil Nadu, India), Kyanos nutrition (Toulouse, France), and Greensea (Mèze, France), which produce C-PC, R-PC, APC, R-PE, and B-PE. Algosource Technology (Saint Nazaire, France) is developing different extracts from Arthrospira platensis (Cyanobacteria), which are Spirulysat, an aqueous extract rich in phycocyanin and Spirugrass, a fraction from the biorefining of Spirulina, rich in amino acids, iron, vitamin K, and beta-carotenes. Spirulina diet supplements are advantageous for athletes, women during pregnancy and lactation, malnourished children, and even for cosmonauts in long space travels.

*Chlorella* is also considered an exceptional source of proteins, especially in its hot water extract, known as *Chlorella* growth factor (CGF), rich in amino acids, vitamins, peptides, minerals, and nucleic acids (DNA and RNA). It was demonstrated to promote growth and healing, stimulate the immune system, and control body weight and serum lipid content [95]. *Chlorella* production has doubled, as it was 2000 tons.year<sup>-1</sup> in 2003 [96], to be over 5000 tons.year<sup>-1</sup> [39]. The largest *Chlorella* producers are situated in Taiwan. Its commercial market value is around one billion US dollars [94].

**Table 4.** Uses of microalgae and Cyanobacteria for food supplement and nutraceutical production by some industries.

	Industry	Product Name	Microalgae/Microfungi Strain	Specificity/Description
		CHLORELLA Green Gem®		Food nutritional supplement
Food supplement	Nutriphys (Enghien, Belgium) https://nutriphys.com/, accessed on 17 March 2021	FLORIL	Auxenochlorella pyrenoidosa (Chlorophyta)	Food supplement: stimulates the intestinal flora while eliminating toxins and heavy metals
		DIETIMAG	-	Food supplement: reduces exhaustion and regain energy

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	VITENE		Dietary supplement: helps to fight against the signs of aging and premature aging
	CHLORELLA PLUS®	- - -	Food supplement with antioxidant properties: revitalizing action, detoxifying the body, toning, acid-base regulator, stimulating natural defenses, helps regulate intestinal transit
	NUbiocell ® TONIC PLUS		Food supplement: improves sportive performance; treats herpes, reduces fatigue; stimulates the immune system, tones the body
Algavia (Oran, Algeria) https://algavia.com/, accessed on 26 January 2021	AlgaVia® Protein- Rich Whole Algae	Auxenochlorella pyrenoidosa (Chlorophyta)	Protein-rich product
AlgaSpring (Almer, Netherlands) https://www.algaspring.nl/, accessed on 3 February 2021	NutriSpring	Microchloropsis gaditana (Eustigmatophyceae)	Food supplement
Fitoplancton Marino (Cadiz, Spain) https://www.tetrasod.com/, accessed on 10 March 2021	TetraSOD	Tetraselmis chuii (Chlorophyta)	Powerful antioxidant and nutritious marine phytoplankton ingredient
Flora (Burnaby, British Columbia, USA) https://www.florahealth.com/, accessed on 17 March 2021	Omega Brain+	<i>Schizochytrium</i> spp. (Fungi)	Vegan rich in EPA and DHA: maintains good health and brain function, supports cardiovascular health, healthy blood triglycerides levels, and cognitive health in an aging population
DSM (Heerlen, Netherlands) https://www.dsm.com/ 23 March 2021	DHASCO-B®	<i>Schizochytrium</i> sp. (Fungi)	Nutritional oil from microalgae containing min. 38% DHA
Sun Chlorella (Torrance, USA) https://www.sunchlorellausa.co m/, accessed on 17 March 2021	Sun Wakasa Gold Plus	<i>Chlorella</i> (Chlorophyta)	Food supplement: extract from the nucleus containing CGF
Microphyt (Baillargues, France) http://www.microphyt.eu/, accessed on 17 March 2021	BrainPhyt	Phaeodactylum tricornutum (Bacillariophyta)	Functional food: Phycoprostans, Xanthophylls, omega-3 fatty acids, sterols Prevent the effects of cognitive decline
Kyanos nutrition (Toulouse, France) https://kyanos- nutrition.com/, accessed on 28 March 2021	KyanosBrain	Aphanizomenon flos- aquae and Arthrospira platensis (Cyanobacteria)	Food supplement: phenylethylamineandphycocyanin
	Spirulina 720	Arthrosnira nlatensis	Food supplement with plant and microalgae powder
	Spirulina 5-Flowers	(Cyanobacteria)	Food supplement with Bach flowers, microalgae and vitamin B5
BlueBioTech/Dr. Peter Hartig (Holstein/Germany)	Astaxanthin	Haematococcus lacustris (Chlorophyta)	Dietary supplement with astaxanthin
nttps://dr-peterhartig.de/, accessed on 28 March 2021	Chlorella coriander 900	Auxenochlorella pyrenoidosa (Chlorophyta)	Food supplement: microalgae powder
	Sunny 200	Dunaliella salina (Chlorophyta)	Food supplement: beta-carotene-rich microalgae powder

		Power 22® double pack	Arthrospira platensis (Cyanobacteria), Auxenochlorella pyrenoidosa (Chlorophyta)	Dietary supplement: microalgae powder
	AlgaTechnologies Ltd. (Hevel Eliot, Israel)	FucoVital™	Phaeodactylum tricornutum (Bacillariophyta)	Fucoxanthin
	accessed on 29 March 2021	AstaPure	Haematococcus lacustris (Chlorophyta)	Astaxanthin
	Fuji Chemical Industries (Toyama, Japan) http://www.fujichemical.co.jp/, accessed on 29 March 2021	AstaReal	Haematococcus lacustris (Chlorophyta)	Astaxanthin
	Cyanotech (Kailua-Kona, Hawai) https://www.cyanotech.com/, accessed on 17 March 2021	BioAstin	Haematococcus lacustris (Chlorophyta)	Astaxanthin
Nutraceutica	Progress Biotech (Capelle aan den IJssel, Netherlands) https://www.progressbiotech.c om/, accessed on 6 March 2021	DHA algal oil	Schizochytrium (Fungi)	DHA (omega-3)
S	Parry nutraceuticals (Onnaiyur, India) https://www.parrynutraceutical s.com/, accessed on 6 January 2021	Organicphycocyanin	Arthrospira/Spirulina (Cyanobacteria)	Phycocyanin
	Kyanos nutrition (Toulouse, France) https://kyanos- nutrition.com/, accessed on 12 March 2021	Kyanos Blue	Arthrospira/Spirulina (Cyanobacteria)	Phycocyanin
-	BASF (Ludwigshafen, Germany) https://nutrition.basf.com/, accessed on 2 February 2021	Betatene	Dunaliella salina (Chlorophyta)	β-carotene
	Algosource (Saint Nazaire, France) https://algosource.com/, accessed on 12 March 2021	Spirulysat	Arthrospira/Spirulina (Cyanobacteria)	Extract of phycocyanin with polysaccharides, amino acids

The global algae protein market will exceed USD 1 billion by 2026; algae protein capsules demand, mainly by sports athletes, will increase by over 6% by the same year. In 2018, dietary supplements represented about 42% of the global algae protein market, simultaneously with the expansion of the nutraceuticals industry in China, Japan, and India. Other microalgae or their extracts, such as Dunaliella, Haematococcus, Scenedesmus (Chlorophyta), Aphanizomenon (Cyanobacteria), Odontella (Bacillariophyta), Porphyridium (Rhodophyta), Schizochytrium, and Ulkenia (Marine Fungi), have been accepted in food and health-food markets [93]. Between 2017 and 2019, three microalgae/microfungi (Euglena gracilis-Euglenozoa, Tetraselmis chui-Chlorophyta, and Aurantiochytrium limacinum-Fungi) were recommended for full Qualified Presumption of Safety status, according by the European Food Safety Authority [97]. Table 5 presents examples of the authorized microalgae and Cyanobacteria and their respective allowed levels for specified foods given by the union list of novel foods in accordance with Regulation European Union (EU) 2015/2283 of the European Parliament and of the Council on novel foods (https://eur-lex.europa.eu/legal-content/, accessed on 27 November 2020) and also by the FDA [98]. Schizochytrium sp. oil, extracted in specified conditions, was authorized as a

novel food by the EFSA [99]. Many companies are producing this oil, rich in DHA, such as DSM (Heerlen, The Netherlands), Progress Biotech (Capelle aan den Ijssel, Netherlands), Bioplus Life Sciences PvT (Bengaluru, Karataka, India), Mara Renewables Corporation (Dartmouth, NH, Canada), and BASF (Ludwigshafen, Germany) (Table 4). It was proposed as an alternative to fish DHA.

Carotenoid production is also in perpetual growth, with a CAGR estimated at 4.2%. Its annual market size is projected to grow from USD 1.5 billion in 2019 to USD 2 billion in 2026 (www.marketsandmarkets.com/, accessed on 26 January 2021). This rise goes together with scientific acceptance and authorization. Betatene, produced by BASF, is an extract of β-carotene from Dunaliella salina used as dietary supplements as well as functional foods and beverages. Dunaliella production also increased from 1200 tons.year-1 to 3000 tons.year<sup>-1</sup>. The *D. salina* global market is estimated to reach a CAGR of 5.1% and will evolve from USD 472.3 million in the year 2020 to US D 671 million by 2027 (www.researchandmarkets.com, accessed on 26 January 2021). The EFSA has accepted the use of an AST-rich oleoresin from Haematococcus lacustris (Chlorophyta) algae as a food supplement up to 40–80 mg.day<sup>-1</sup> (Table 5) [100]. AST was shown to be a powerful dietary antioxidant. There are also many companies producing AST, such as Fuji Chemical Industries Co., Ltd. (Toyama, Japan), Cyanotech (Kailua-Kona, Hawai, USA), Algalif (Reykjanesbaer, Iceland), and Microphyt (Baillargues, France). Its global market is expected to reach USD 770 million by 2024, where the global nutraceuticals industry will be assessed to have consumed 54.8% of H. lacustris microalgae natural AST in 2017 to reach 190 metric tons by 2024 (www.businesswire.com, accessed on 14 February 2021). The H. lacustris market is estimated to grow at a CAGR of 13.2% from 2020 to 2027 to reach USD 148.1 million by 2027 (www.globenewswire.com, accessed on 14 February 2021).

**Table 5.** Microalgae and Cyanobacteria authorized for incorporation into food products according to the European Union ("EUR-Lex-02017R2470-20200827-EN-EUR-Lex," n.d.) and the Food and Drug Administration (Nutrition, 2020).

Microalgae/ Microfungi	Fraction	Specified Food Category	Maximum Dose	Authorized by
		Bakery products (breads, rolls, and sweet cookies)	200 mg.100 g <sup>-1</sup>	
Ulkenia sp.	Extracted oil	Cereal bars	500 mg.100 g <sup>-1</sup>	
(Marine Fungi)		Non-alcoholic drinks (including milk drinks)	60 mg.100 mL <sup>-1</sup>	
Haematococcus lacustris (Chlorophyta)	extracted Oleoresin rich in astaxanthin	Food supplements	40–80 mg oleoresin per day, equivalent to ≤8 mg astaxanthin per day	_
		Flavored pasta	1.5%	
		Fish soups	1%	
Odontella aurita		Marine terrines	0.5%	
(Bacillariophyta)		Court bouillon preparations	1%	EU
		Crackers	1.5%	_
		Frozen breaded fish	1.5%	
		Food supplements, intended for the adult population, excluding pregnant or breastfeeding women	3000 mg.day <sup>-1</sup>	
Schizochytrium sp. (Marine Fungi)	Extracted oil with high	Food supplements, intended for pregnant or breastfeeding women	450 mg.day <sup>-1</sup>	
	DITA and EFA content	Substitutes for the total daily ration for weight control	250 mg	
		Milk-based beverages and similar products intended for young children	200 mg.100 g <sup>-1</sup>	_

		Cereal-based preparations and baby food intended for infants and young children Foods suitable for intense muscular expenditure (for athletes)		
		Dairy product substitutes, except beverages	600 mg.100 g <sup>-1</sup> for cheeses; 200 mg.100 g <sup>-1</sup> for soy milk and imitation milk products (except beverages)	
		Sauces	20% or 250 mg.day <sup>-1</sup>	
Tetraselmis chuii	Dried microalgae	Special salts	1%	
(Chlorophyta)	Difee incrouigae	Condiments	250 mg.day <sup>-1</sup>	
		Food supplements	250 mg.day <sup>-1</sup>	
Arthrospira/Spirulina (Cyanobacteria)	Extract (color additive)	Coloring confections, frostings, ice cream and frozen desserts, dessert coatings and toppings, beverage mixes and powders, yogurts,		FDA
<i>Haematococcus</i> sp. (Chlorophyta)	Meal	Salmonid fish feed only	Total astaxanthin from all astaxanthin color additive sources ≤80 mg per kg of finished feed	FDA

The introduction of microalgae as food ingredients and the demand for high-addedvalue products are increasing despite some concerns of the algae powder consistence, color, fishy smell, and production cost [101]. Some companies have incorporated *Chlorella* in noodles (Sun Chlorella (Torrance, CA, USA), Paldo (Seoul, Korea)), bars and cookies (Allma (Lisboa, Portugal)), and in mayonnaise to replace eggs (Fooding Company (Sucyen-Brie, France)) (Table 6).

Microalgae constitute equally an important feed for animals, especially with the expansion of aquaculture, the reduction of land fertility, and scientific evidence on their positive effect on animal health. For example, carotenoids have been incorporated as a pigment in egg yolks, broiler skin, fishes, and crustaceans. Microalgae feeds are largely used in aquaculture and poultry as a source of DHA and proteins. Nannostar, based on the microalgae *Microchloropsis gaditana* (Eustigmatophyceae), is produced by AlgaSpring (Almere, The Netherlands) for aquaculture purposes. It is rich in EPA (20:5 omega-3) and other fatty acids such as C14:0, C16:0, C16:1, C18:1, and C18:2. It contains  $\beta$ -carotene, violaxanthin, tocopherols, various sterols, and valuable minerals. Some industries use a combination of microalgae, as in the case of Easybooster (Fitoplancton marino, Cádiz, Spain), commercialized as a liquid phytoplankton extract combining 33% of *Isochrysis* (T-ISO), 31% of *Nannochloropsis*, 18% of *Tetraselmis*, and 18% *Phaeodactylum* (Table 6).

Microalgae are an important food for some marine bivalve mollusks, the larval stages of abalone and conch gastropods, the larvae of several marine fish species, penaeid shrimp, and zooplankton [102]. For adult fish, the use of microalgae is limited as fishmeal is still cheaper [101]. Oostlander estimated the cost of live microalgae production for feed in aquaculture hatcheries as 300–600 EUR.kg<sup>-1</sup> biomass. This cost could be reduced using larger-scale production [103].

Table 6. Microalgae exploitation in food and feed domains.

	Industry	Product Name	Microalgae Strain	Description
		Bars with microalgae—Chlorella		Bars
Food htt	Allma (Pataias, Portugal) https://www.allmicroalgae.com/en/, accessed on 26 January 2021	Chlorella Cookies	Chlovalla	Cookies
		Soods Crup chy Bar Bar with	(Chlorophyta)	Seeds
		seeds Crunchy Bar. Bar with		Crunchy
		Interoargae		Bar

	Sun Chlorella (Torrance, USA) https://www.sunchlorellausa.com/, accessed on 17 March 2021	Chlorella Udon Noodles	<i>Chlorella</i> (Chlorophyta)	Noodles
	Paldo (Seoul, South Korea) http://www.paldofood.com/, accessed on 18 March 2021	Green Tea Chlorella Noodles	<i>Chlorella</i> (Chlorophyta)	Noodles
-	Fooding Company (Sucy-en-Brie, France) http://foodingcompany.com/, accessed on 18 March 2021	The good spoon	<i>Chlorella</i> (Chlorophyta)	Mayonnaise
	eChlorial (L'isle-d'Abeau, France) https://www.echlorial.fr/, accessed on 19 March 2021	TetraselmischuiieChlorial	Tetraselmis chuii (Chloroophyta)	Sea spice, condiment
	The Algae Factory (Wageningen, Netherlands) http://thealgaefactory.com/the-milk-chocolate/, accessed on 17 March 2021	The dark chocolate	Arthrospira/Spirulin a (Cynaophya)	Chocolate
Feed	AlgaSpring (E. Heimansweg Almere, The Netherlands) https://www.algaspring.nl/nannochloropsis-gaditana- micro-algae/, accessed on 17 March 2021	NannoStar	Microchloropsis gaditana (Chlorophyta)	Aquacultur e feed
	TomAlgae (Belgium) https://www.tomalgae.com/, accessed on 19 March 2021	TomAlgae (ThalaPure)	freeze-dried algae	Aquacultur e feed
	Solazyme (San Francisco, USA) https://algaprime.com/, accessed on 17 March 2021	AlgaPrime™ DHA	Microalgae	Companion animals and in aquaculture feed

#### 3.2. Use of Microalgae and Cyanobacteria for Cosmetic Applications

Microalgae and Cyanobacteria have been incorporated into face, skin, and hair care products, in addition to sun protection compounds such as antioxidants, thickening, and water-binding agents [104]. Some industries in this field develop their own microalgae culture, but others tend to develop collaborations with industries producing microalgae, such as Rejuvasea<sup>®</sup> (NV, USA), which commercializes products containing active compounds from marine microalgae provided by Fitoplancton Marino (Cadiz, Spain). Some brands incorporate in their products extracts from more than one microalga in combination with other plant extracts, as Estee Lauder (New York, NY, USA) and Algenist, which is Solazyme's skincare line. The latter produces a whole range of cosmetic products based on microalgae (Table 7). In the literature, several works have been carried out to evaluate the effect of these extracts by in vitro and in vivo studies. Rumin et al. [105] reported that in 2015 and 2017, the number of publications was about 25 per year, where France, Spain, and Germany were the top publishers. They focus mainly on the antioxidant and antiinflammatory activities of microalgae extracts and the potential of bioactive compounds in the treatment of skin problems such as aging, tanning, and pigment disorders [106]. For instance, the effect of Dermochlorella D, produced by CODIF, an aqueous extract of *Chlo*rella vulgaris rich in amino acids, was COSMOS approved. This label is managed by a notfor-profit, international, and independent association, and it represents a consumer guarantee for organic and natural cosmetics. The effect of this extract was studied by in vitro test on fibroblasts and keratinocytes. It was shown that the mentioned extract ameliorated the skin structure by an increase in collagen-1, -3, and -4 and also laminin-6 and elastin expressions by human fibroblasts, as well as collagen-7, laminin-5, and elafin expressions by keratinocytes. It also improved skin protection against free radicals by increasing the expression of thioredoxin-1 and thioredoxin-2, which could reduce skin process aging. Furthermore, it protects Langerhans cells against ultraviolet exposure [106]. Phormiskin from CODIF Company took the second prize at the "BSB Innovation Awards 2010" for the most innovative ingredient focused on natural products.

Industry	Product	Microalgae	Fraction Used	Effect	
	PEPHA®-CTIVE	Dunaliella salina (Chloroophyta)	aqueous extract rich in	Skin care: reloads skin with	
			amino acids, minerals,	new energy and improves its	
		(1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	and carbohydrates	radiance.	
DSM	) PEPHA®-TIGHT	Microchloropsis	aqueous extract rich in	Skin care: Tightening effect	
(Heerlen, The Netherlands)		gaditana	vitamin C, vitamin B12,	and long-term skin firming	
https://www.dsm.com/,		(Chlorophyta)	and polysaccharides	effect	
accessed on 26 January 2021	PEPHA-AGE	Halochlorella rubescens (Chlorophyta)	aqueous extract rich in	Chine mucha stien from hisso	
			(B2) algal saccharidas	light and LW	
			and minerals (7n)	light and 0 v	
Evonik (Frankfurt Germany)	/ TEGO® Stemlastin	Cyanidium caldarium (Rhodophyta)	n.d.	Skin care: retains a youthful	
https://corporate.evonik.com/				skin appearance and reduces	
en, accessed on 26 January				the signs of chronological skin	
2021				aging	
				Skin care: against photo-aging	
	Phormiskinbioprotech g	Pseudanabaena persicina	Concentrate microalgae	of the skin, makes skin	
				pigmentation uniform and	
		(Cyanobacteria)		adds luster to the skin tone	
		<i>Rhodella</i> (Rhodophyta)	Concentrate microalgae	Skin care: improves tissue	
CODIE technologie naturalle				oxygenation and luminosity	
(SaintMalo, France)	Detoxondria			of the skin, reduces the	
http://www.codif-tp.com/				susceptibility of the skin to	
accessed on 12 January 2021				fatigue and signs of fatigue	
accessed on 12 January 2021	Rosacea	Rhodella violacea	The complete extract	Skin care: hydrates the skin	
	Dermochlorella D	(Rhodophyta)	1		
		Chlorella vulgaris	aqueous extract rich in	Skin care: reactivates collagen	
		(Chlorophyta)	amino acid concentrate	synthesis	
	Goldella	(Chlorophyta)	Latein	Skin care: Anti-Aging	
Sun Chlorella (Torrance,	Astarella Primetime	Chlorella	futchi	Skin care: ensures healthy-	
USA)	Skin Cream	(Chlorophyta)	Astaxanthin and CGF	looking skin	
https://www.sunchlorellausa.		(Chiefephyta)	la 20% CGF		
com/, accessed on 26 January	Sun Chlorella Cream	Chlorella		Skin care: Nourishes and	
2021		(Chlorophyta)		hydrates the skin	
Kalia Naturel (Bondy, France)		Arthrospira/Spirulina	Microalgae powder	Hair care: strengthens the hair	
https://kalianature.com/fr/,	Protect My Hair				
accessed on 2 March 2021		(Cyanobacteria)		and accelerates their growth	
	BLUE ALGAE VITAMIN C™ Dark	Arthrospira/Spirulina (Cyanobacteria)	Extracted vitamin C	Skin care: reduces the	
				appearance of dark spots &	
	Spot Correcting Peel			discolorations	
Algenist (Torrance, CA, USA)	ELEVATE Advanced	Dunaliella salina	n.d.	Skin care: anti-aging	
https://www.algenist.com/,	Lift Contouring Cream	(Chlorophyta)			
accessed on 2 March 2021	GENIUS Liquid Collagen®	Parachlorella	Parachlorella beijerinckii Exopolysaccharides and Chlorella protothecoides Oil	Skin care: enhances skin's bounce and resilience	
		beijerinckii and			
		Auxenochlorella			
		protothecoides			
Estas laudar (Naw York		(Chlorophyta)			
Estee lauder (New Tork,	Nutritious Micro-Algae, Pore Purifying Cleansing Jelly	(Chlorena ouigaris	Chlorella vulgaris extract and Spirulina platensis	Chin care, col to romano	
UJA) https://www.estaoloudor.fr		Arthroenira nlatancio		makeup and impurities.	
accessed on 17 March 2021		(Cvanobacteria)	powder		
		Phaeodactulum		Skin care: Anti-pollution anti-	
Givaudan (Vernier,	Depollutine	tricornutum	Peptidic extract	ageing	
Switzerland)	Sensityl	(Bacillariophyta)	Water extract	Skin care: calm the skin	
	<i>,</i>	· · · /			

 Table 7. Microalgae and Cyanobacteria exploitation in the cosmetic field.

https://www.givaudan.com/, accessed on 17 March 2021	Costalane	Skeletonema costatum	Poly-unsaturated fatty acid rich in omega-3	Skin care: restore skin homeostasis and promotes epidermal differentiation
	Grevilline <sup>™</sup> PF	(Bacinanophyta)	Peptidic extract	Anti-inflammatory, Anti- redness
	Hydrintense	Porphyridium purpureum (Rhodophyta)	Exopolysaccharide	Skin care: hydrates the skin
	Mariliance	<i>Rhodosorus marinus</i> (Rhodophyta)	n.d.	Neurosoother
	blue algae life water	Arthrospira/Spirulina (Cyanobacteria)	Extract rich in vitamins, minerals, trace elements, and essential amino-acid	Skin care: stimulates and revitalizes the skin
	ReinaPhyt	Chlamydomonas	overno ot might in	Skin care: protects the skin
	MonaPhytcarotenes rich extract	<i>reinhardtii</i> (Chlorophyta)	carotenoids	Skin care: rejuvenates the skin
Microphyt (Baillargues, France) http://www.microphyt.eu/,	Luteana	<i>Tisochrysis lutea</i> (Haptophyta, Coccolithophyceae)	lipidic fractions rich in xanthophylls	Skin care: soothing effect and anti-aging
accessed on 20 March 2021	Renouvellance	Porphyridium purpureum (Rhodophyta)	Phycoerythrin and EPS	Skin care: Anti-aging, anti- pollution, exposomial protection, radiance enhancer and sun care
Expanscience (Epernon, France) with microphyt (Baillargues, France) https://www.expanscience.co m/fr/, accessed on 25 March 2021	ALGAENIA	Chlamydomonas acidophila (Chlorophyta)	peptide concentrate	Skin care: protects sensitive skin
GREENSEA (Mèze, France) http://greensea.fr/, accessed on 27 March 2021	Silidine®	Porphyridium purpureum (Rhodophyta)	mixture of oligosaccharides and trace elements	Skin care: corrects skin dysfunctions
Sumrise (Holzmindon	SymControl	Tetraselmis suecica (Chlorophyta)	n.d.	Skin care: reduce sebum overproduction, soothing the skin and strengthening the skin barrier.
Germany) https://www.symrise.com/,	SymBronze®1659	Isochrysis Galbana	n.d.	Skin care: increases skin pigmentation and accelerates the natural tanning process
accessed on 29 March 2021	SymHair® Force 1631	(Haptophyta, Coccolithophyceae)	n.d.	Hair care: prevents hair loss, improves volume, and makes hair stronger, healthier, and more vital
algoVita (Tunis, Tunisia)	CONFORTING SOOTHING CREAM	Porphyridium purpureum (Rhodophyta)	n.d.	Skin care: provides intense and long-lasting hydration, soothes, and promotes skin repair.
https://www.algovita.tn/, accessed on 19 January 2021	LIPOFILLER SERUM	<i>Nannochloropsis</i> (Eustigmatophyceae)	n.d.	Skin care: treat older stains
· · ·	DEPICLEAR SERUM	Haematococcus lacustris (Chlorophyta)	astaxanthin	Skin care: unifies skin complexion while reducing spots and imperfections.

3.3. Use of Microalgae and Cyanobacteria for Biofuel Applications

The use of microalgae in recent years for the production of "third-generation" biofuel is increasing as this renewable biosource is not in conflict with critical food production. Indeed, it does not compete with agriculture or at the expense of food, as is the case with first- and second-generation biofuels. In addition, biofuel production from microalgae does not cause deforestation or habitat destruction. These microorganisms are suitable for biofuel production thanks to their high content of lipids, which can reach over 84% of dry cell weight among some species. Lipids produced by microalgae are converted into biofuels, biogas, dihydrogen (H2), and bioelectricity. The term biofuels encompass biodiesel, bioethanol, and biomethane. From microalgae containing 30% oil by weight of dry biomass, 587,000 L of oil per hectare can be obtained [12]. Additionally, their richness of carbohydrates is convertible to fermentable sugars, making them convenient for bioethanol production [107]. Moreover, the production of biofuel by microalgae has the advantage of reducing greenhouse gas emissions and preserving the environment. It has been reported that, during their growth, a production of 100 kg of algae consumes 182 kg of CO<sub>2</sub>, which has a positive effect on the climate; in addition, some species could use SOx and NOx as a nutrient source [38,105].

The global biofuel algae market is expected to have a CAGR of 8.8% and to reach USD 10.73 billion by 2025, with a dominance of their use in transportation to account for over 70% of the total demand (www.grandviewresearch.com/, accessed on 19 January 2021).

However, until now, there have been many challenges and difficulties of this production, such as the enormous water demand and the necessity of maintaining algae cultivation across all climates. If wastewater is used to produce oil, the barrel of algae oil could cost USD 332, according to the estimation of Lundquist et al. [108]. This price is too high compared to crude oil, which was USD 80 in 2010 as the average closing price and USD 56 in 2021. The proposed solutions are to capitalize the production and develop technologies for larger-scale productions. Some industries use genetically modified microalgae or Cyanobacteria to produce biofuel, as in the case of Algenol, which uses modified Cyanobacteria for ethanol production [109]. European Aeronautic Defence and Space (EADS), the parent company of Airbus, used, in 2010, 100% derived biofuel microalgae to fly a Diamond DA42NG for 4 days. With the aim of testing and certification, Solazyme (San Francisco, CA, USA) produced over 36,000 L of 100% algae-derived biofuel destined for the U.S. Navy in 2010. In the next few years, it is awaited that North America will be the largest market, accounting for over 30% (www.grandviewresearch.com/, www.grandviewresearch.com/, accessed on 19 January 2021). In addition to Solazyme, there are many industries producing energy based on algae in the United States, such as Algenol, Blue Marble Production, Sapphire Energy, Culture Biosystems, Genifuels, Seambiotic, Solix Biofuels, and Algae Production Systems, and in other countries, such as Proviron in Belgium (Hemiksem, Belgium).

#### 4. Conclusions

Microalgae is a group of eukaryotic photosynthetic microorganisms that thrive in many habitats and present significant taxonomic diversity. They present obvious properties distinguishing them from Cyanobacteria (sometimes called prokaryotic microalgae or blue-green microalgae). Typically, microalgae are photoautotrophic microorganisms producing biomass with great efficiency using water, solar energy, and CO<sub>2</sub>, implying biochemical diversity, which becomes the basis for many biotechnological, commercial, and industrial sectors to harness the growing demands of such an unexplored natural resource.

Despite the huge use of microalgal and Cyanobacterial biomass in many industrial applications, as presented in this review, some bioactive compounds from these biomasses still depict an entirely passive role because of the photoproduction, extraction, and purification costs. Moreover, the selection of algal strains after screening from collections or isolation from different environments as well as their cultivation in appropriate culture

media always require numerous developments. Thus, the development of novel and adapted extraction and purification methodologies applied to algal biomass might initiate the turning event toward wider industrial utilization. Moreover, the improvement of biomass production in photobioreactors after process intensification could, probably in the future, significantly increase biomass yields by capturing CO<sub>2</sub>. Indeed, photoproduction in dilute aqueous media using natural or artificial light remain currently inappropriate for large-scale biomass production and low-value applications. Additionally, more work needs to be done to fully utilize the potential of microalgae and Cyanobacteria biomass for their application in large-scale production. Since the key challenges appear to be the high cost of operation, infrastructure, maintenance, mass production, bioproduct accumulation, and extraction, a proposed solution is the cultivation of microalgae in wastewaters, thanks to their ability to assimilate organic and inorganic carbon as well as inorganic nitrogen (N) and phosphorous (P), in order to reduce the cost of cultures.

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#### References

- 1. Jacob-Lopes, E.; Maroneze, M.M.; Queiroz, M.I.; Zepka, L.Q. Handbook of Microalgae-Based Processes and Products: Fundamentals and Advances in Energy, Food, Feed, Fertilizer, and Bioactive Compounds; Academic Press: Cambridge, MA, USA, 2020.
- Gonçalves, A.L.; Rodrigues, C.M.; Pires, J.C.M.; Simões, M. The Effect of Increasing CO<sub>2</sub> Concentrations on Its Capture, Biomass Production and Wastewater Bioremediation by Microalgae and Cyanobacteria. *Algal Res.* 2016, 14, 127–136. https://doi.org/10.1016/j.algal.2016.01.008.
- 3. Alvarez, A.L.; Weyers, S.L.; Goemann, H.M.; Peyton, B.M.; Gardner, R.D. Microalgae, Soil and Plants: A Critical Review of Microalgae as Renewable Resources for Agriculture. *Algal Res.* **2021**, *54*, 102200. https://doi.org/10.1016/j.algal.2021.102200.
- 4. Abu-Ghosh, S.; Dubinsky, Z.; Verdelho, V.; Iluz, D. Unconventional High-Value Products from Microalgae: A Review. *Bioresour. Technol.* 2021, 329, 124895. https://doi.org/10.1016/j.biortech.2021.124895.
- Chapman, V.J.; Chapman, D.J. Classification. In *The Algae*; Chapman, V.J., Chapman, D.J., Eds.; Macmillan Education: London, UK, 1973; pp. 1–12.
- Palinska, K.A.; Surosz, W. Taxonomy of Cyanobacteria: A Contribution to Consensus Approach. *Hydrobiologia* 2014, 740, 1–11. https://doi.org/10.1007/s10750-014-1971-9.
- Rockwell, N.C.; Lagarias, J.C.; Bhattacharya, D. Primary Endosymbiosis and the Evolution of Light and Oxygen Sensing in Photosynthetic Eukaryotes. *Front. Ecol. Evol.* 2014, 2, 66. https://doi.org/10.3389/fevo.2014.00066.
- 8. Mostafa, S.S.M. Microalgal Biotechnology: Prospects and Applications; IntechOpen: London, UK, 2012.
- 9. Lauersen, K.J. Eukaryotic Microalgae as Hosts for Light-Driven Heterologous Isoprenoid Production. *Planta* **2019**, 249, 155–180. https://doi.org/10.1007/s00425-018-3048-x.
- 10. Lee, R.E. Phycology; Cambridge University Press: Cambridge, UK, 1989.
- Pulz, O.; Gross, W. Valuable Products from Biotechnology of Microalgae. Appl. Microbiol. Biotechnol. 2004, 65, 635–648. https://doi.org/10.1007/s00253-004-1647-x.
- Suresh, K.S.; Suresh, P.V.; Kudre, T.G. 4—Prospective Ecofuel Feedstocks for Sustainable Production. In Advances in Eco-Fuels for a Sustainable Environment; Azad, K., Ed.; Woodhead Publishing Series in Energy, Woodhead Publishing: Sawston, UK, 2019; pp. 89–117.
- Garcia-Pichel, F. Cyanobacteria. In *Encyclopedia of Microbiology*, 3rd ed.; Schaechter, M., Ed.; Academic Press: Cambridge, MA, USA, 2009; pp. 107–124.
- Mata, T.M.; Martins, A.A.; Caetano, Nidia.S. Microalgae for Biodiesel Production and Other Applications: A Review. *Renew. Sustain. Energy Rev.* 2010, 14, 217–232. https://doi.org/10.1016/j.rser.2009.07.020.
- 15. de Vargas, C.; Audic, S.; Henry, N.; Decelle, J.; Mahé, F.; Logares, R.; Lara, E.; Berney, C.; Le Bescot, N.; Probert, I.; et al. Ocean Plankton. Eukaryotic Plankton Diversity in the Sunlit Ocean. *Science* **2015**, *348*, 1261605. https://doi.org/10.1126/science.1261605.

- Sheehan, J.; Dunahay, T.; Benemann, J.; Roessler, P. Look Back at the U.S. Department of Energy's Aquatic Species Program: Biodiesel from Algae; Close-Out Report; National Renewable Energy Lab.: Golden, CO, USA, 1998.
- 17. Norton, T.A.; Melkonian, M.; Andersen, R.A. Algal Biodiversity. *Phycologia* **1996**, *35*, 308–326. https://doi.org/10.2216/i0031-8884-35-4-308.1.
- 18. Richmond, A. Handbook of Microalgal Culture: Biotechnology and Applied Phycology; Blackwell Publishing: Hoboken, NJ, USA, 2004.
- 19. Guiry, M.D. How Many Species of Algae Are There? J. Phycol. 2012, 48, 1057–1063. https://doi.org/10.1111/j.1529-8817.2012.01222.x.
- 20. Metting, F.B. Biodiversity and Application of Microalgae. J. Ind. Microbiol. 1996, 17, 477–489. https://doi.org/10.1007/BF01574779.
- García-Balboa, C.; Baselga-Cervera, B.; García-Sanchez, A.; Igual, J.M.; Lopez-Rodas, V.; Costas, E. Rapid Adaptation of Microalgae to Bodies of Water with Extreme Pollution from Uranium Mining: An Explanation of How Mesophilic Organisms Can Rapidly Colonise Extremely Toxic Environments. *Aquat. Toxicol. Amst. Neth.* 2013, 144–145, 116–123. https://doi.org/10.1016/j.aquatox.2013.10.003.
- Yong, W.K.; Tan, Y.H.; Sze-Wan, P.; Lim, P.E. Response of Microalgae in a Changing Climate and Environment. *Malays. J. Sci.* 2016, 35, 167–187. https://doi.org/10.22452/mjs.vol35no2.7.
- Suparmaniam, U.; Lam, M.K.; Uemura, Y.; Lim, J.W.; Lee, K.T.; Shuit, S.H. Insights into the Microalgae Cultivation Technology and Harvesting Process for Biofuel Production: A Review. *Renew. Sustain. Energy Rev.* 2019, 115, 109361. https://doi.org/10.1016/j.rser.2019.109361.
- 24. de Morais, M.G.; da Vaz, B.S.; de Morais, E.G.; Costa, J.A.V. Biologically Active Metabolites Synthesized by Microalgae. *BioMed Res. Int.* 2015, 2015, 835761. https://doi.org/10.1155/2015/835761.
- Dvořák, P.; Casamatta, D.A.; Hašler, P.; Jahodářová, E.; Norwich, A.R.; Poulíčková, A. Diversity of the Cyanobacteria. In Modern Topics in the Phototrophic Prokaryotes: Environmental and Applied Aspects; Hallenbeck, P.C., Ed.; Springer International Publishing: Cham, Switzerland, 2017; pp. 3–46.
- 26. Thajuddin, N.; Subramanian, G. Cyanobacterial Biodiversity and Potential Applications in Biotechnology. *Curr. Sci.* 2005, *89*, 47–57.
- 27. Whitton, B.; Potts, M. The Ecology of Cyanobacteria: Their Diversity in Time and Space; Springer: Berlin/Heidelberg, Germany, 2002.
- 28. Vermaas, W.F. Photosynthesis and Respiration in Cyanobacteria. In *eLS*; American Cancer Society: Atlanta, GA, USA, 2001.
- 29. Whitton, B. Diversity, Ecology, and Taxonomy of the Cyanobacteria. *Photosynth. Prokaryotes* **1992**, 1–51, https://doi.org/10.1007/978-1-4757-1332-9\_1.
- 30. Vachard, D. Cyanobacteria. In *Reference Module in Earth Systems and Environmental Sciences*; Elsevier: Amsterdam, The Netherlands, 2019.
- 31. Stal, L.J.; Moezelaar, R. Fermentation in Cyanobacteria. *FEMS Microbiol. Rev.* **1997**, *21*, 179–211. https://doi.org/10.1111/j.1574-6976.1997.tb00350.x.
- 32. Andersen, R.A. Diversity of Eukaryotic Algae. Biodivers. Conserv. 1992, 1, 267–292. https://doi.org/10.1007/BF00693765.
- Cavalier-Smith, T. Principles of Protein and Lipid Targeting in Secondary Symbiogenesis: Euglenoid, Dinoflagellate, and Sporozoan Plastid Origins and the Eukaryote Family Tree1,2. J. Eukaryot. Microbiol. 1999, 46, 347–366. https://doi.org/10.1111/j.1550-7408.1999.tb04614.x.
- 34. Raven, J.A.; Allen, J.F. Genomics and Chloroplast Evolution: What Did Cyanobacteria Do for Plants? Genome Biol. 2003, 4, 209.
- 35. Jeffrey, S.W.; Wright, S.W.; Zapata, M. Microalgal Classes and Their Signature Pigments. *Phytoplankton Pigment*. 2011, https://doi.org/ 10.1017/CBO9780511732263.004
- 36. Archibald, J.M. Endosymbiosis and Eukaryotic Cell Evolution. *Curr. Biol.* 2015, 25, R911–R921. https://doi.org/10.1016/j.cub.2015.07.055.
- Baurain, D.; Brinkmann, H.; Petersen, J.; Rodríguez-Ezpeleta, N.; Stechmann, A.; Demoulin, V.; Roger, A.J.; Burger, G.; Lang, B.F.; Philippe, H. Phylogenomic Evidence for Separate Acquisition of Plastids in Cryptophytes, Haptophytes, and Stramenopiles. *Mol. Biol. Evol.* 2010, *27*, 1698–1709. https://doi.org/10.1093/molbev/msq059.
- 38. Person, J. Livre Turquoise—Algues, Filière Du Future; Actes Colloq. Adebiotech: Romainville, France, 2011.
- Levasseur, W.; Perré, P.; Pozzobon, V. A Review of High Value-Added Molecules Production by Microalgae in Light of the Classification. *Biotechnol. Adv.* 2020, 41, 107545. https://doi.org/10.1016/j.biotechadv.2020.107545.
- 40. de Reviers, B. Biologie et Phylogénie des Algues; Belin Editeur: Paris, France, 2002.
- 41. Groendahl, S.; Kahlert, M.; Fink, P. The Best of Both Worlds: A Combined Approach for Analyzing Microalgal Diversity via Metabarcoding and Morphology-Based Methods. *PLoS ONE* **2017**, *12*, e0172808. https://doi.org/10.1371/journal.pone.0172808.
- Correa, I.; Drews, P.; Botelho, S.; de Souza, M.S.; Tavano, V.M. Deep Learning for Microalgae Classification. In Proceedings of the 2017 16th IEEE International Conference on Machine Learning and Applications (ICMLA), Cancun, Mexico, 18–21 December 2017; pp. 20–25.
- Heimann, K.; Huerlimann, R. Chapter 3—Microalgal Classification: Major Classes and Genera of Commercial Microalgal Species. In *Handbook of Marine Microalgae*; Kim, S.-K., Ed.; Academic Press: Cambridge, MA, USA 2015; pp. 25–41.
- 44. Malcata, F.X.; Pinto, I.S.; Guedes, A.C. *Marine Macro- and Microalgae: An Overview*; Malcata, F.X., Pinto, I.S., Guedes, A.C., Eds.; CRC Press: Boca Raton, FL, USA, 2018.
- 45. Fritsch, F.E. The Structure and Reproduction of The Algae, 1st ed.; Cambridge University Press: Cambridge, UK, 1922; Volume 1.
- 46. Fritsch, F.E The Structure and Reproduction of The Algae, 1st ed.; Cambridge University Press: Cambridge, UK, 1923; Volume 2.

- Ruggiero, M.A.; Gordon, D.P.; Orrell, T.M.; Bailly, N.; Bourgoin, T.; Brusca, R.C.; Cavalier-Smith, T.; Guiry, M.D.; Kirk, P.M. A Higher Level Classification of All Living Organisms. *PLoS ONE* 2015, 10, e0119248. https://doi.org/10.1371/journal.pone.0119248.
- 48. Sialve, B.; Steyer, J.-P. Les microalgues, promesses et défis. Innov. Agron. 2013, 26, 25–39.
- 49. Rochet, M.; Legendre, L.; Demers, S. Photosynthetic and Pigment Responses of Sea-Ice Microalgae to Changes in Light Intensity and Quality. J. Exp. Mar. Biol. Ecol. **1986**, 101, 211–226. https://doi.org/10.1016/0022-0981(86)90264-9.
- 50. Pazderník, B.M. Light Harvesting Complexes and Chromatic Adaptation of Eustigmatophyte Alga Trachydiscus Minutus. Ph.D. Thesis, Faculty of Science, University of South Bohemia, České Budějovice, Czech Republic, 2015.
- Dutton, H.J.; Juday, C. Chromatic Adaptation in Relation to Color and Depth Distribution of Freshwater Phytoplankton and Large Aquatic Plants. *Ecology* 1944, 25, 273–282. https://doi.org/10.2307/1931276.
- 52. Hentati, F.; Tounsi, L.; Djomdi, D.; Pierre, G.; Delattre, C.; Ursu, A.V.; Fendri, I.; Abdelkafi, S.; Michaud, P. Bioactive Polysaccharides from Seaweeds. *Molecules* **2020**, *25*, 3152. https://doi.org/10.3390/molecules25143152.
- Ben Hlima, H.; Bohli, T.; Kraiem, M.; Ouederni, A.; Mellouli, L.; Michaud, P.; Abdelkafi, S.; Smaoui, S. Combined Effect of Spirulina Platensis and Punica Granatum Peel Extacts: Phytochemical Content and Antiphytophatogenic Activity. *Appl. Sci.* 2019, 9, 5475. https://doi.org/10.3390/app9245475.
- 54. Elleuch, F.; Hlima, H.B.; Barkallah, M.; Baril, P.; Abdelkafi, S.; Pichon, C.; Fendri, I. Carotenoids Overproduction in Dunaliella Sp.: Transcriptional Changes and New Insights through Lycopene β Cyclase Regulation. *Appl. Sci.* 2019, *9*, 5389. https://doi.org/10.3390/app9245389.
- Elleuch, F.; Baril, P.; Barkallah, M.; Perche, F.; Abdelkafi, S.; Fendri, I.; Pichon, C. Deciphering the Biological Activities of Dunaliella Sp. Aqueous Extract from Stressed Conditions on Breast Cancer: From in Vitro to in Vivo Investigations. *Int. J. Mol. Sci.* 2020, *21*, 1719. https://doi.org/10.3390/ijms21051719.
- Ibañez, E.; Cifuentes, A. Benefits of Using Algae as Natural Sources of Functional Ingredients. J. Sci. Food Agric. 2013, 93, 703– 709. https://doi.org/10.1002/jsfa.6023.
- 57. Becker, E.W. Micro-Algae as a Source of Protein. *Biotechnol. Adv.* 2007, 25, 207–210. https://doi.org/10.1016/j.bio-techadv.2006.11.002.
- Sousa, I.; Gouveia, L.; Batista, A.P.; Raymundo, A.; Bandarra, N.M. Microalgae in Novel Food Products. *Food Chem. Res. Dev.* 2008, 75–112.
- Niccolai, A.; Chini Zittelli, G.; Rodolfi, L.; Biondi, N.; Tredici, M.R. Microalgae of Interest as Food Source: Biochemical Composition and Digestibility. *Algal Res.* 2019, 42, 101617. https://doi.org/10.1016/j.algal.2019.101617.
- 60. Vaz, B. da S.; Moreira, J.B.; de Morais, M.G.; Costa, J.A.V. Microalgae as a New Source of Bioactive Compounds in Food Supplements. *Curr. Opin. Food Sci.* 2016, 7, 73–77. https://doi.org/10.1016/j.cofs.2015.12.006.
- Kose, A.; Ozen, M.O.; Elibol, M.; Oncel, S.S. Investigation of in Vitro Digestibility of Dietary Microalga Chlorella Vulgaris and Cyanobacterium Spirulina Platensis as a Nutritional Supplement. *3 Biotech* 2017, *7*, 170. https://doi.org/10.1007/s13205-017-0832-4.
- 62. Siqueira, S.F.; Queiroz, M.I.; Jacob-Lopes, L.Q.Z.E. Introductory Chapter: Microalgae Biotechnology A Brief Introduction. *Microalgal Biotechnol.* **2018**. https://doi.org/10.5772/intechopen.73250.
- 63. Borowitzka, M.A. Microalgae as Sources of Pharmaceuticals and Other Biologically Active Compounds. J. Appl. Phycol. **1995**, 7, 3–15. https://doi.org/10.1007/BF00003544.
- 64. Pedrós, R.; Moya, I.; Goulas, Y.; Jacquemoud, S. Chlorophyll Fluorescence Emission Spectrum inside a Leaf. *Photochem. Photobiol. Sci.* 2008, *7*, 498–502. https://doi.org/10.1039/B719506K.
- Gaignard, C.; Gargouch, N.; Dubessay, P.; Delattre, C.; Pierre, G.; Laroche, C.; Fendri, I.; Abdelkafi, S.; Michaud, P. New Horizons in Culture and Valorization of Red Microalgae. *Biotechnol. Adv.* 2019, 37, 193–222. https://doi.org/10.1016/j.biotechadv.2018.11.014.
- Vílchez, C.; Forján, E.; Cuaresma, M.; Bédmar, F.; Garbayo, I.; Vega, J.M. Marine Carotenoids: Biological Functions and Commercial Applications. *Mar. Drugs* 2011, 9, 319–333. https://doi.org/10.3390/md9030319.
- 67. Molino, A.; Rimauro, J.; Casella, P.; Cerbone, A.; Larocca, V.; Chianese, S.; Karatza, D.; Mehariya, S.; Ferraro, A.; Hristoforou, E.; et al. Extraction of Astaxanthin from Microalga Haematococcus Pluvialis in Red Phase by Using Generally Recognized as Safe Solvents and Accelerated Extraction. J. Biotechnol. 2018, 283, 51–61. https://doi.org/10.1016/j.jbiotec.2018.07.010.
- Rammuni, M.N.; Ariyadasa, T.U.; Nimarshana, P.H.V.; Attalage, R.A. Comparative Assessment on the Extraction of Carotenoids from Microalgal Sources: Astaxanthin from H. Pluvialis and β-Carotene from D. Salina. *Food Chem.* 2019, 277, 128–134. https://doi.org/10.1016/j.foodchem.2018.10.066.
- 69. de Raposo, M.F.J.; de Morais, A.M.M.B.; de Morais, R.M.S.C. Carotenoids from Marine Microalgae: A Valuable Natural Source for the Prevention of Chronic Diseases. *Mar. Drugs* **2015**, *13*, 5128–5155. https://doi.org/10.3390/md13085128.
- Cuellar-Bermudez, S.P.; Aguilar-Hernandez, I.; Cardenas-Chavez, D.L.; Ornelas-Soto, N.; Romero-Ogawa, M.A.; Parra-Saldivar, R. Extraction and Purification of High-Value Metabolites from Microalgae: Essential Lipids, Astaxanthin and Phycobiliproteins. *Microb. Biotechnol.* 2015, *8*, 190–209. https://doi.org/10.1111/1751-7915.12167.
- Thomas, J.-C. L'antenne Collectrice d'énergie Lumineuse à Phycobiliprotéines Chez Les Cyanobactéries. Bull. Société Bot. Fr. Actual. Bot. 1989, 136, 31–49. https://doi.org/10.1080/01811789.1989.10826916.
- 72. Hamed, I. The Evolution and Versatility of Microalgal Biotechnology: A Review. Compr. Rev. Food Sci. Food Saf. 2016, 15, 1104–1123. https://doi.org/10.1111/1541-4337.12227.

- Dammak, M.; Haase, S.M.; Miladi, R.; Ben Amor, F.; Barkallah, M.; Gosset, D.; Pichon, C.; Huchzermeyer, B.; Fendri, I.; Denis, M.; et al. Enhanced Lipid and Biomass Production by a Newly Isolated and Identified Marine Microalga. *Lipids Health Dis.* 2016, 15. https://doi.org/10.1186/s12944-016-0375-4.
- Dammak, M.; Hadrich, B.; Barkallah, M.; Hentati, F.; Ben Hlima, H.; Pichon, C.; Denis, M.; Fendri, I.; Michaud, P.; Abdelkafi, S. Modelling Tetraselmis Sp. Growth-Kinetics and Optimizing Bioactive-Compound Production through Environmental Conditions. *Bioresour. Technol.* 2018, 249, 510–518. https://doi.org/10.1016/j.biortech.2017.10.028.
- 75. Singh, S.; Kate, B.N.; Banerjee, U.C. Bioactive Compounds from Cyanobacteria and Microalgae: An Overview. *Crit. Rev. Biotechnol.* 2005, 25, 73–95. https://doi.org/10.1080/07388550500248498.
- Kumar, J.; Singh, D.; Tyagi, M.B.; Kumar, A. Chapter 16–Cyanobacteria: Applications in Biotechnology. In *Cyanobacteria*; Mishra, A.K., Tiwari, D.N., Rai, A.N., Eds.; Academic Press: Cambridge, MA, USA, 2019; pp. 327–346.
- Lau, N.-S.; Matsui, M.; Abdullah, A.A.-A. Cyanobacteria: Photoautotrophic Microbial Factories for the Sustainable Synthesis of Industrial Products. *BioMed Res. Int.* 2015, 2015. https://doi.org/10.1155/2015/754934.
- Markou, G.; Angelidaki, I.; Georgakakis, D. Microalgal Carbohydrates: An Overview of the Factors Influencing Carbohydrates Production, and of Main Bioconversion Technologies for Production of Biofuels. *Appl. Microbiol. Biotechnol.* 2012, 96, 631–645. https://doi.org/10.1007/s00253-012-4398-0.
- Mourelle, M.L.; Gómez, C.P.; Legido, J.L. The Potential Use of Marine Microalgae and Cyanobacteria in Cosmetics and Thalassotherapy. Cosmetics 2017, 4, 46. https://doi.org/10.3390/cosmetics4040046.
- de Raposo, M.F.J.; de Morais, R.M.S.C.; de Morais, A.M.M.B. Bioactivity and Applications of Sulphated Polysaccharides from Marine Microalgae. *Mar. Drugs* 2013, 11, 233–252. https://doi.org/10.3390/md11010233.
- Christaki, E.; Florou-Paneri, P.; Bonos, E. Microalgae: A Novel Ingredient in Nutrition. Int. J. Food Sci. Nutr. 2011, 62, 794–799. https://doi.org/10.3109/09637486.2011.582460.
- Galasso, C.; Gentile, A.; Orefice, I.; Ianora, A.; Bruno, A.; Noonan, D.M.; Sansone, C.; Albini, A.; Brunet, C. Microalgal Derivatives as Potential Nutraceutical and Food Supplements for Human Health: A Focus on Cancer Prevention and Interception. *Nutrients* 2019, *11*, 1226. https://doi.org/10.3390/nu11061226.
- Škrovánková, S. Seaweed Vitamins as Nutraceuticals. Adv. Food Nutr. Res. 2011, 64, 357–369. https://doi.org/10.1016/B978-0-12-387669-0.00028-4.
- 84. Brasil, B.; Siqueira, F.; Salum, T.; Zanette, C.; Spier, M. Microalgae and Cyanobacteria as Enzyme Biofactories. *Algal Res.* **2017**, 25, 76–89. https://doi.org/10.1016/j.algal.2017.04.035.
- Lu, Y.; Xu, J. Phytohormones in Microalgae: A New Opportunity for Microalgal Biotechnology? *Trends Plant Sci.* 2015, 20, 273–282. https://doi.org/10.1016/j.tplants.2015.01.006.
- Bajguz, A. Brassinosteroids in Microalgae: Application for Growth Improvement and Protection Against Abiotic Stresses. In Brassinosteroids: Plant Growth and Development; Hayat, S., Yusuf, M., Bhardwaj, R., Bajguz, A., Eds.; Springer: Berlin/Heidelberg, Germany, 2019; pp. 45–58.
- Amorim, M.L.; Soares, J.; dos Coimbra, J.S.R.; de Leite, M.O.; Albino, L.F.T.; Martins, M.A. Microalgae Proteins: Production, Separation, Isolation, Quantification, and Application in Food and Feed. *Crit. Rev. Food Sci. Nutr.* 2020, *61*, 1976–2002. https://doi.org/10.1080/10408398.2020.1768046.
- 88. Pruvost, J.; Cornet, J.-F.; Pilon, L. Large-Scale Production of Algal Biomass: Photobioreactors. In *Algae Biotechnology: Products and Processes*; Bux, F., Chisti, Y., Eds.; Green Energy and Technology Springer: Cham, Switzerland, 2016; pp. 41–66.
- 89. Costa, J.A.V.; de Morais, M.G. Chapter 1—An Open Pond System for Microalgal Cultivation. In *Biofuels from Algae*; Pandey, A., Lee, D.-J., Chisti, Y., Soccol, C.R., Eds.; Elsevier: Amsterdam, The Netherlands, 2014; pp. 1–22.
- Costa, J.A.V.; Freitas, B.C.B.; Santos, T.D.; Mitchell, B.G.; Morais, M.G. Chapter 9–Open Pond Systems for Microalgal Culture. In *Biofuels from Algae*; 2nd ed.; Pandey, A., Chang, J.-S., Soccol, C.R., Lee, D.-J., Chisti, Y., Eds.; Biomass, Biofuels, Biochemicals; Elsevier: Amsterdam, The Netherlands, 2019; pp. 199–223.
- 91. Jerney, J.; Spilling, K. Large Scale Cultivation of Microalgae: Open and Closed Systems. In *Biofuels from Algae: Methods and Protocols*; Spilling, K., Ed.; Methods in Molecular Biology; Springer: New York, NY, USA, 2020; pp. 1–8.
- 92. Singh, G.; Patidar, S.K. Microalgae Harvesting Techniques: A Review. J. Environ. Manage. 2018, 217, 499–508. https://doi.org/10.1016/j.jenvman.2018.04.010.
- 93. Chacón-Lee, T.L.; González-Mariño, G.E. Microalgae for "Healthy" Foods Possibilities and Challenges. *Compr. Rev. Food Sci. Food Saf.* 2010, 9, 655–675. https://doi.org/10.1111/j.1541-4337.2010.00132.x.
- 94. Mobin, S.; Alam, F. Some Promising Microalgal Species for Commercial Applications: A Review. *Energy Procedia* 2017, *110*, 510–517. https://doi.org/10.1016/j.egypro.2017.03.177.
- An, B.-K.; Kim, K.-E.; Jeon, J.-Y.; Lee, K.W. Effect of Dried Chlorella Vulgaris and Chlorella Growth Factor on Growth Performance, Meat Qualities and Humoral Immune Responses in Broiler Chickens. *SpringerPlus* 2016, 5, 718. https://doi.org/10.1186/s40064-016-2373-4.
- 96. Milledge, J.J. Microalgae Commercial Potential for Fuel, Food and Feed. *Food Sci. Amp Technol.* **2012**, *26*, 28–30.
- 97. Koutsoumanis, K.; Allende, A.; Alvarez-Ordóñez, A.; Bolton, D.; Bover-Cid, S.; Chemaly, M.; Davies, R.; Cesare, A.D.; Hilbert, F.; Lindqvist, R.; et al. Scientific Opinion on the Update of the List of QPS-Recommended Biological Agents Intentionally Added to Food or Feed as Notified to EFSA (2017–2019). EFSA J. 2020, 18, e05966. https://doi.org/10.2903/j.efsa.2020.5966.
- 98. Nutrition, C. for F.S. and A. Summary of Color Additives for Use in the United States in Foods, Drugs, Cosmetics, and Medical Devices. *FDA* **2020**.

- Turck, D.; Castenmiller, J.; Henauw, S.D.; Hirsch-Ernst, K.I.; Kearney, J.; Maciuk, A.; Mangelsdorf, I.; McArdle, H.J.; Naska, A.; Pelaez, C.; et al. Safety of Schizochytrium Sp. Oil as a Novel Food Pursuant to Regulation (EU) 2015/2283. EFSA J. 2020, 18, e06242. https://doi.org/10.2903/j.efsa.2020.6242.
- 100. Turck, D.; Castenmiller, J.; de Henauw, S.; Hirsch-Ernst, K.I.; Kearney, J.; Maciuk, A.; Mangelsdorf, I.; McArdle, H.J.; Naska, A.; Pelaez, C.; et al. Safety of Astaxanthin for Its Use as a Novel Food in Food Supplements. *EFSA J.* 2020, 18, e05993. https://doi.org/10.2903/j.efsa.2020.5993.
- Voort, M.P.J. van der; Vulsteke, E.; de Visser, C.L.M. Macro-Economics of Algae Products: Output WP2A7.02; EnAlgae Swansea University: Swansea, UK, 2015.
- 102. Baert, P.; Bosteels, T.; Sorgeloos, P. Manual on the Production and Use of Live Food for Aquaculture. Pond Prod. 1996, 196–251.
- Oostlander, P.C.; van Houcke, J.; Wijffels, R.H.; Barbosa, M.J. Microalgae Production Cost in Aquaculture Hatcheries. *Aquaculture* 2020, 525, 735310. https://doi.org/10.1016/j.aquaculture.2020.735310.
- Rizwan, M.; Mujtaba, G.; Memon, S.A.; Lee, K.; Rashid, N. Exploring the Potential of Microalgae for New Biotechnology Applications and beyond: A Review. *Renew. Sustain. Energy Rev.* 2018, 92, 394–404. https://doi.org/10.1016/j.rser.2018.04.034.
- 105. Rumin, J.; Nicolau, E.; de Junior, R.G.O.; Fuentes-Grünewald, C.; Picot, L. Analysis of Scientific Research Driving Microalgae Market Opportunities in Europe. Mar. Drugs 2020, 18. https://doi.org/10.3390/md18050264.
- 106. Morvan, P.-Y.; Vallee, R. Effects of Chlorella Extract on Skin. Pers. Care. 2007, 7, 57–64.
- 107. Khan, M.I.; Shin, J.H.; Kim, J.D. The Promising Future of Microalgae: Current Status, Challenges, and Optimization of a Sustainable and Renewable Industry for Biofuels, Feed, and Other Products. *Microb. Cell Factories* 2018, 17. https://doi.org/10.1186/s12934-018-0879-x.
- 108. Lundquist, T.; Woertz, I.; Quinn, N.; Benemann, J. A Realistic Technology and Engineering Assessment of Algae Biofuel Production. *Energy* **2010**, 1.
- 109. Legere, E.; Roessler, P.; Miller, H.; Belicka, L.; Yuan, Y.; Chance, R.; Dalrymple, K.; Porubsky, W.; Coleman, J.; Sweeney, K.; et al. *Recovery Act—Integrated Pilot-Scale Biorefinery for Producing Ethanol from Hybrid Algae*; Algenol Biotech LLC: Fort Myers, FL, USA, 2017.