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► **To cite this version:**

David Garcia-Gragera, Carolina Arnau, Enrique Peiro, Claude-Gilles Dussap, Laurent Poughon, et al.. Integration of Nitrifying, Photosynthetic and Animal Compartments at the MELiSSA Pilot Plant. *Frontiers in Astronomy and Space Sciences*, 2021, 8, pp.177. 10.3389/fspas.2021.750616 . hal-03468640

HAL Id: hal-03468640

<https://uca.hal.science/hal-03468640>

Submitted on 7 Dec 2021

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Integration of Nitrifying, Photosynthetic and Animal Compartments at the MELiSSA Pilot Plant

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Specialty section:

This article was submitted to
Astrobiology,
a section of the journal
Frontiers in Astronomy and Space
Sciences

Received: 30 July 2021

Accepted: 22 September 2021

Published: 19 October 2021

Citation:

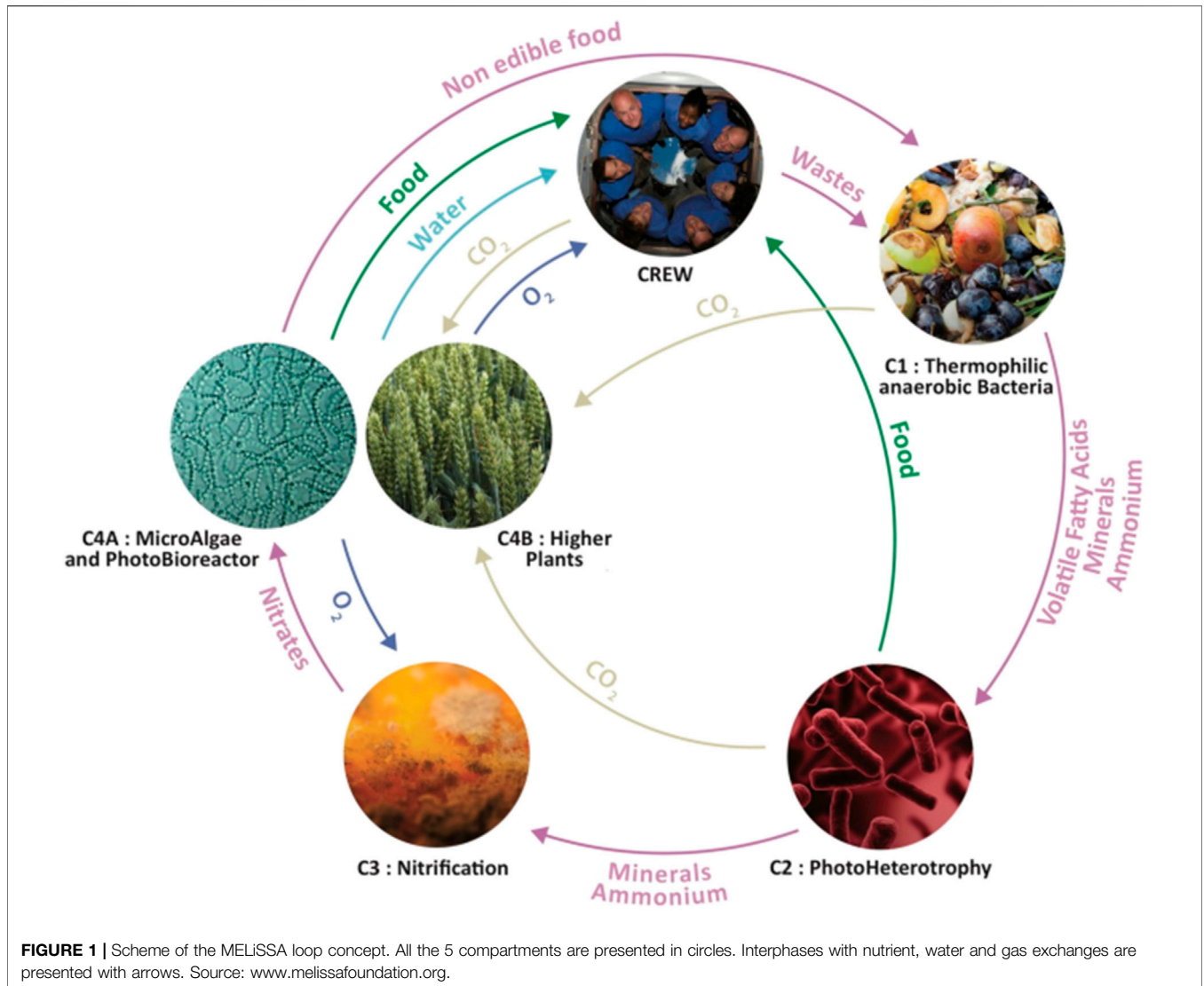
Garcia-Gragera D, Arnau C, Peiro E, Dussap C-G, Poughon L, Gerbi O, Lamaze B, Lasseur C and Godia F (2021) Integration of Nitrifying, Photosynthetic and Animal Compartments at the MELiSSA Pilot Plant. *Front. Astron. Space Sci.* 8:750616. doi: 10.3389/fspas.2021.750616

MELiSSA (Micro Ecological Life Support System Alternative) is developing bioregenerative Life Support technologies for long-term Space missions. The MELiSSA concept is conceived as a loop with several compartments, each one performing a specific function, providing all together edible material production, atmosphere regeneration and water recovery with a concomitant use of wastes, i.e., CO₂ and organic wastes. Each one of the compartments is colonized with specific bacteria or higher plants depending on its specific function. The MELiSSA Pilot Plant is a facility designed for the terrestrial demonstration of this concept, hosting laboratory rats as a crew mock-up mimicking the respiration of humans. Currently, the MELiSSA Pilot Plant focus on the integration of three compartments: Compartment 3 (nitrifying packed-bed bioreactor based on the co-culture of immobilized *Nitrosomonas europaea* and *Nitrobacter winogradsky*), compartment 4a (an air-lift photobioreactor for the culture of the edible cyanobacteria *Limnospira indica* with concomitant oxygen production) and Compartment 5 (an animal isolator with rats as mock-up crew). The output from these tests shows a high robustness and reliability and the performance of oxygen producing and oxygen consuming compartments is successfully demonstrated under transitory and steady-state conditions. This contribution reports on the current state of development of the MELiSSA Pilot Plant Facility and the most recent results of the integration work.

Keywords: Continuous operation, bioreactors, *Limnospira indica*, *Nitrosomonas europaea*, *Nitrobacter winogradsky*, Micro Ecological Life Support System Alternative, Environmental Control and Life Support System

INTRODUCTION

The MELiSSA (Micro-Ecological Life Support System Alternative) project focusses on the development and integration of a system providing life support for long-term human missions in space (Mergeay et al., 1988), such as a base on Moon or Mars, and is led by the European Space Agency, in the context of an international consortium with 15 partners (Lasseur et al., 2010). The project approach is inspired in an ecological system by reproducing its main functions in specific compartments (Hendrickx et al., 2006). In the proposed closed loop structure, first, Compartments 1



and 2 are dedicated to the degradation of wastes, with a first step based on thermophilic bacteria and a second complementary step of biotransformation of the wastes. Compartment 3 performs nitrification and it is an aerobic bioreactor working with an axenic pure co-culture of *Nitrosomonas* and *Nitrobacter* autotrophic bacteria. This co-culture transforms ammonium (NH_4^+) to nitrite (NO_2^-) and then to nitrate (NO_3^-), the last one being a more assimilable form of nitrogen for the photosynthetic elements of the loop. Some *Nitrospira* related microorganisms have been discovered over the past years that perform complete ammonium oxidation to nitrate in one organism, instead of the two-stage nitrification, such as *Nitrospira inopinata* (Daims et al., 2015). It is considered a promising discovery, but so far their application in a scenario of intensive bioprocessing has not yet been achieved to fulfill the MELISSA and urine nitrification requirements. Hence, the canonical two-stage nitrification as selected originally for MELISSA has been maintained. Compartment 4a and 4b are dedicated to oxygen, water and edible biomass production from the consumption of CO_2 , evapo-

transpiration and using light as energy source, and they are a photobioreactor (PBR) working with an axenic pure culture of the cyanobacteria *Limnospira indica* and a higher plant compartment for the culture of three representative plants [*Lactuca sativa* (lettuce), *Triticum aestivum* (wheat) and *Beta vulgaris* (red beet)], respectively. Finally, Compartment 5 corresponds to the animal isolator, in which a group of rats is used to mimic the crew. The animals are basically connected to the rest of the loop in the gas phase to demonstrate the atmosphere regeneration to support breathing. A complete illustration of the loop is shown in **Figure 1**. Those compartments are operated in continuous mode for long-term periods under controlled conditions, based on the use of mathematical models developed for each compartment, and are connected one to each other through various interfaces (gas, liquid and solid). Overall, the system should provide the essential functions of life support: food production, air regeneration, water reclamation and waste treatment when fully developed, and represents a unique effort in the development of life

support systems enhancing the self-sustainability of human activities in Space.

Taking into consideration the complexity of these proposed system, the MELiSSA Consortium has developed the MELiSSA Pilot Plant (MPP) with the main goal of demonstrating long-term continuous operation feasibility under the supervision of a control system (Gòdia et al., 2004; Poughon et al., 2009) with knowledge-based models that reproduce each compartment's individual characterization and intercompartment dynamics. Thus, in this pilot plant, the different compartments have been scaled-up to achieve the oxygen production equivalent to the respiration needs of one human ($0.84 \text{ kg}\cdot\text{d}^{-1}$) (Wieland, 2005), with 20–40% concomitant production of edible material. The MPP is developed using terrestrial conditions, with the main objective to demonstrate the feasibility of the MELiSSA loop concept, using an industrial approach based on systems engineering, operating under high quality standards, including clean room operation for the area hosting axenic compartments (Compartments 3 and 4a). This dedicated facility to MELiSSA loop demonstration is located at Universitat Autònoma de Barcelona and it is a European Space Agency external laboratory. To note, MELiSSA Consortium is performing in parallel studies to test MELiSSA compartments at Space conditions as well as studies on the application of the MELiSSA technology to Earth in order to foster circular systems.

The MELiSSA Pilot Plant is developed in a stepwise approach. First, the individual compartments have been designed according to the final scenario of the MPP based on the research work done previously to characterize them and develop their mathematical models. Once manufactured and installed in the MPP site, the compartments, associated instrumentation and auxiliary equipment have been operated and fully characterized for a wide range of operational conditions, while testing simultaneously the monitoring and control elements developed for each one of them. Second, the so called integration phase has followed. This phase implies the progressive connection of the compartments, involving gas, liquid and solid phases. Today, the integration work is at the level of connecting Compartment 3 (nitrification), Compartment 4a (photosynthesis) and Compartment 5 (crew compartment), both in gas and liquid phase. The results of the integration are reported in this contribution and have been obtained in three consecutive steps. First, compartments 4a and 5 have been connected in close loop in the gas phase. The results from this first connection have been reported previously (Alemany et al., 2019). Second, compartments 3 and 4a have been connected in the liquid phase. Third, the two previous connections have been combined, so, compartments 5 and 4a connected in gas phase and 3 and 4a connected in liquid phase. Each integration step has been operated for long-term periods (several months of continuous operation) under different operational conditions, including several steady-state conditions and the corresponding transitory phases. In the following, the results obtained in the last two steps of integration are discussed, representing a step forward in the integration of the building blocks that should lead to the demonstration of the MELiSSA loop concept

and envisage how it would contribute to human exploration missions in Space.

MATERIALS AND METHODS

Nitrifying Reactor (Compartment 3)

Description

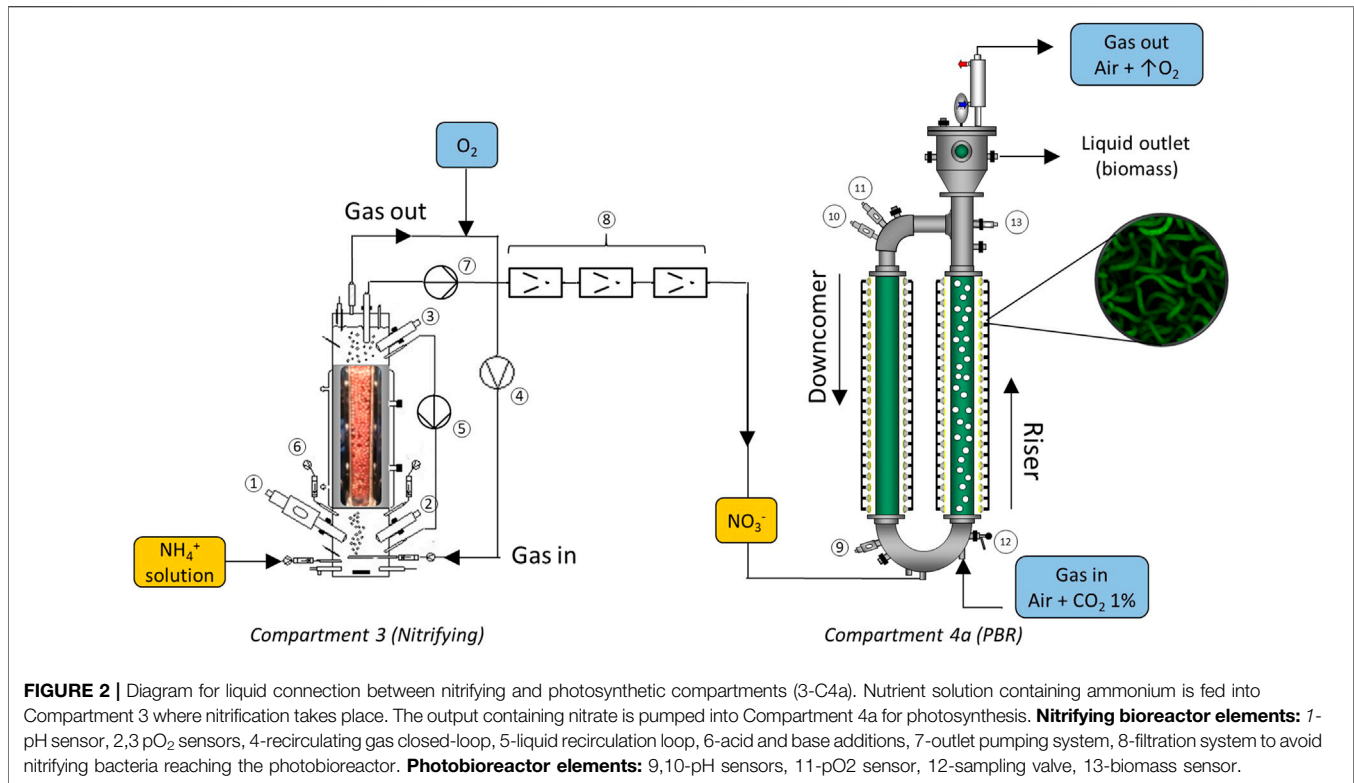
The nitrification compartment (Compartment 3) is a cylindrical packed-bed bioreactor of 7 L operation volume (SNC-Lavalin, Brussels, Belgium and Bioprocess Technologies, Madrid, Spain). It is packed with polystyrene beads and colonized by a co-culture of *Nitrosomonas europaea* and *Nitrobacter winogradsky* that grow forming a biofilm. *N. europaea* is an Ammonia Oxidizing Bacteria (AOB) capable to oxidize ammonia into nitrite. Then, complete nitrification is achieved by the oxidation of nitrite into nitrate by the Nitrite Oxidizing Bacteria (NOB) *N. winogradsky*. 2 mol of oxygen are needed for the complete oxidation of 1 mol of N-NH_4^+ making it an aerobic process. On top of the central packed-bed section, the reactor has a bottom section, mechanically stirred, where fresh liquid feeding, liquid recirculation inlet and gas sparging take place. There is as well a top section after the packed-bed for gas-liquid separation, and liquid and recirculation outlets. Both top and bottom section hold the instrumentation for on-line monitoring of the compartment (pH, T, pO_2 , conductivity). It is operated in a recirculation closed gas-loop mode regulated by means of a mass flow-meter (Bronkhorst, F-202D-FA, Bronkhorst, Ruurlo, Netherlands). pH is measured by means of two sterilizable glass pH probe connected to a pH amplifier (Mettler Toledo, Inpro 3253, Greifensee Switzerland). pO_2 is measured by means of two Clark amperometric sensor (Mettler Toledo, InPro6950i/12/320, Greifensee, Switzerland) (see Figure 2).

Photobioreactor (Compartment 4a)

Description

The photobioreactor compartment (Compartment 4a) is an 83L external-loop gas lift photobioreactor (Bioengineering Ag, Wald, Switzerland). It is used for *Limnospira indica* cultivation. The reactor consists of two glass cylindrical tubes with 15 cm diameter and 1.5 m height with an illuminated volume of 55 L. Illumination is provided by 350 halogen lamps (MR16HM, (12 V, 20 W), Sylvania, Mississauga, Ontario, Canada) around the glass tubes and can be adjusted by the control system of the loop in order to regulate oxygen production. The upper and lower parts of the tubes are connected by U-shaped stainless steel sections that allow improved liquid circulation and heat exchange through an external jacket. The gas phase is injected through the bottom of the right column (riser) and exits from the gas separation section situated at the top of that same column, which creates a difference of density that allows the fluid to return through the left column (downcomer) (see Figure 2).

For continuous operation of the bioreactor during the experiment, the inlet feed medium is pumped by a variable speed gear pump (Lewa, EEC0002S11, Hispania S.L., Leonberg, Germany) followed by a $0.2 \mu\text{m}$ filtration



(MCY4440DFLPH4, Pall Corporation, New York, United States). Outlet flow is regulated with a second variable speed gear pump (Lewa Hispania S.L., EEC0002S11, Leonberg, Germany) Inlet and outlet air supply is regulated and measured by means of three flow-meters and controllers (Bronkhorst, F-202D-FA, Bronkhorst, Ruurlo, Netherlands). pH is measured by means of a conventional sterilizable glass pH probe connected to a pH amplifier (Mettler Toledo, Inpro 3253, Switzerland). Control of pH is performed by addition of NaOH (2 M) or H₂SO₄ (2 M) as required. The composition of the outlet gas is continuously monitored on-line by a IR analyser for CO₂ and a paramagnetic analyser for O₂ (600 series, CAI, United States).

Animal Compartment Description

The crew mock-up compartment (Compartment 5) is an animal isolator of 1550 L volume (Hosokawa Micron Ltd., Runcorn, UK). It is divided in three zones: main chamber, transfer airlock and recirculation loop. It is designed to host the animal crew with artificial light at periods of 12–12 h day-night cycles. It includes six 250 mm round glove ports at the front of the isolator to perform the animal support operations.

In the main chamber, an overpressure of 200 Pa is maintained by pressure control through two pressure transmitters (Rosemount, 2051 CG1, Shakopee, Minnesota, United States). An external 250 L buffer tank is connected to the isolator to compensate the effects of atmospheric pressure changes. Temperature and humidity in the isolator are monitored by the use of two transmitters (Vaisala, HMT337, Helsinki, Finland) and controlled at 22°C and 55%, respectively. A gas

recirculation loop ensures a homogenous distribution of air in the isolator using a recirculation flow of 90 m³·h⁻¹. An active carbon filter is installed (Bi-On[®] ACPA, Bioconservación, Barcelona, Spain) in order to avoid accumulation of contaminants during long operational periods. CO₂ and O₂ are measured on-line through an IR and paramagnetic analyzers respectively (SICK-Maihak, GSM810, Hamburg, Germany). Finally, the transfer airlock operates with an independent pressure control to allow the entry and exit of materials (cages, food, bedding, etc.) without affecting the main isolator variables.

Three Wistar rats (Charles River Laboratories, Domaine des Oncins, France) with an age of 12 weeks were selected as the suitable crew model for inhabiting the isolator and running the experiments. The rationale for the animals selection has been described previously (Alemany et al., 2019). A different group of rats is used for each experimental condition where different liquid flow rates of the liquid loop are tested. The rats are fed with an *ab libitum* diet (Teckad 2914, Envigo, United States). Weekly monitoring of the rats weight is performed.

Cell Strains and Culture Media

Nitrobacter winogradsky and *Nitrosomonas europaea* were provided by DSMZ, (Germany) for nitrifying bioreactor culture. *Limnospira indica* PCC 8005 was provided by SKC CEN (Mol, Belgium) for PBR cultivation. The strains were grown axenically during the experiment in their corresponding bioreactor. Culture media was a combination of a modified Zarrouk medium (Cogne et al., 2003) and the medium defined in the literature for nitrifying cultures (Pérez et al., 2004):

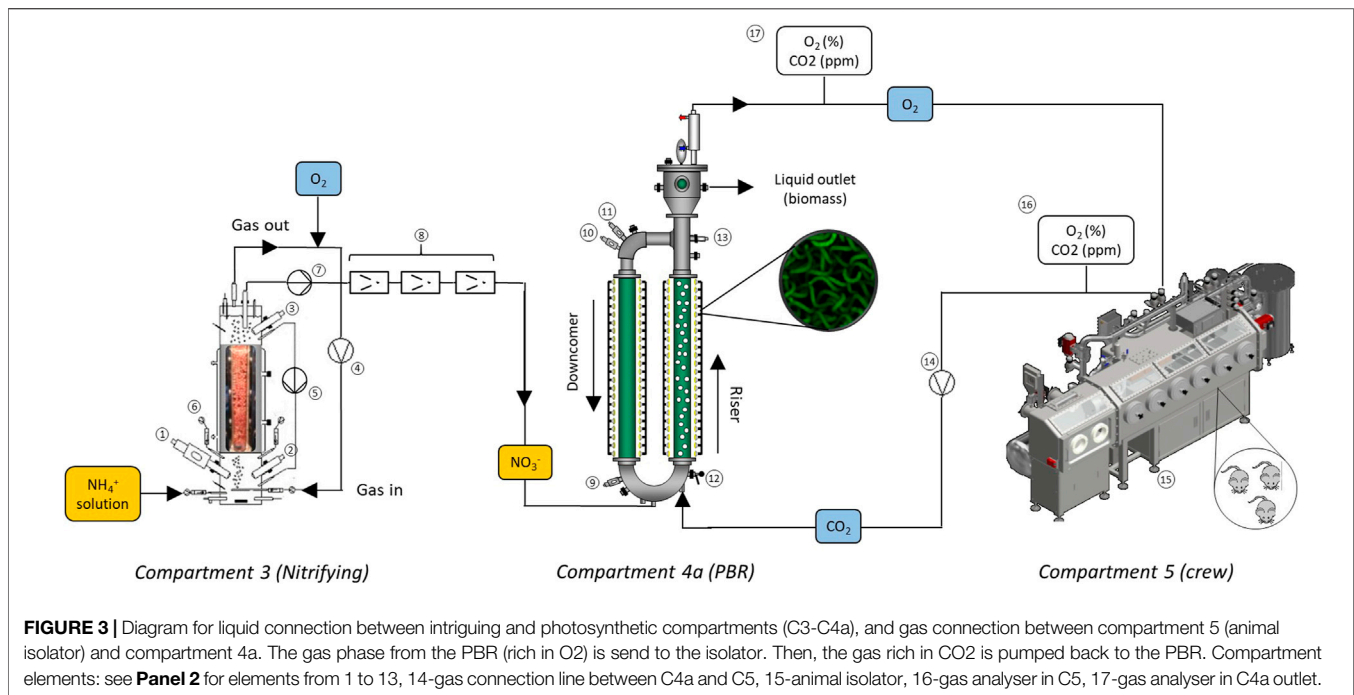


FIGURE 3 | Diagram for liquid connection between intriguing and photosynthetic compartments (C3-C4a), and gas connection between compartment 5 (animal isolator) and compartment 4a. The gas phase from the PBR (rich in O_2) is sent to the isolator. Then, the gas rich in CO_2 is pumped back to the PBR. Compartment elements: see **Panel 2** for elements from 1 to 13, 14-gas connection line between C4a and C5, 15-animal isolator, 16-gas analyser in C5, 17-gas analyser in C4a outlet.

1.358 $\text{g}\cdot\text{L}^{-1}$ of $(\text{NH}_4)_2\text{SO}_4$, 1 $\text{g}\cdot\text{L}^{-1}$ of K_2SO_4 , 1 $\text{g}\cdot\text{L}^{-1}$ of NaCl , 0.816 $\text{g}\cdot\text{L}^{-1}$ of NaHCO_3 , 0.71 of Na_2HPO_4 , 0.68 of KH_2PO_4 , 0.5 of K_2HPO_4 , 0.1 $\text{g}\cdot\text{L}^{-1}$ of $\text{MgSO}_4\cdot 7\text{H}_2\text{O}$, 0.18 $\text{g}\cdot\text{L}^{-1}$ of $(\text{NH}_4)_6\text{Mo}_7\text{O}_{24}\cdot 4\text{H}_2\text{O}$, 0.08 $\text{g}\cdot\text{L}^{-1}$ $\text{EDTA}\cdot 2\text{Na}\cdot 2\text{H}_2\text{O}$, 0.01 $\text{g}\cdot\text{L}^{-1}$ of $\text{FeSO}_4\cdot 7\text{H}_2\text{O}$, 0.04 $\text{g}\cdot\text{L}^{-1}$ of $\text{CaCl}_2\cdot 2\text{H}_2\text{O}$, 1.81 $\text{mg}\cdot\text{L}^{-1}$ of $\text{MnCl}_2\cdot 4\text{H}_2\text{O}$, 0.22 $\text{mg}\cdot\text{L}^{-1}$ of $\text{ZnSO}_4\cdot 7\text{H}_2\text{O}$, and 0.079 $\text{mg}\cdot\text{L}^{-1}$ of $\text{CuSO}_4\cdot 5\text{H}_2\text{O}$.

Liquid Connection Between Compartment 3 and 4a

A liquid stream was constantly supplied from a reservoir tank to the inlet of the nitrifying bioreactor. Liquid effluent from nitrifying bioreactor was constantly processed by a three-step filtration to maintain axenicity in both bioreactors. This was a critical operation to avoid any cells detached from Compartment 3 biofilm reaching Compartment 4a, with the associated contamination effects. The effluent from nitrifying bioreactor was pumped to the inlet of the PBR with a gear pump (Lewa, EEC0002S11, Hispania S.L., Leonberg, Germany). A variable peristaltic pump (Watson-Marlow 323Du, Watson Marlow Fluid Technology Group, Wilmington, United States) at the nitrifying bioreactor outlet was used to keep an overpressure in the liquid connection line at 200 mbarg. Finally, the PBR effluent was continuously collected in a harvesting tank (see **Figure 2**).

A set of different liquid flow rates corresponding to three NH_4^+ loads ($\text{ppm}\cdot\text{d}^{-1}$) into compartment 3 were initially tested: 10, 20 and 30 $\text{L}\cdot\text{d}^{-1}$. A combination of testing conditions was defined by setting two illumination levels (q_o): 120 and 285 $\text{W}\cdot\text{m}^{-2}$. So, a total of six conditions were tested. Operational conditions in the PBR were established as follows:

pH was set at nominal value of 8.5, CO_2 was constantly injected at 1% composition in the air inlet at a total gas flow of 2.8 $\text{L}\cdot\text{min}^{-1}$ and temperature was set at 36°C. Conditions in Compartment 3 were as follows: pH was controlled at 8.1 by the addition of H_2SO_4 (0.1 M) and KOH (1.9 M), pO_2 was maintained at 80% thanks to the injection of external pure O_2 into the recirculation closed gas-loop, a recirculation liquid flow between the top and bottom section of the bioreactor was set at 4.5 $\text{L}\cdot\text{h}^{-1}$ to improve the homogeneity and temperature was set at 30°C.

Gas Connection Between Compartments 4a and 5

The animal isolator was connected to the PBR at the entrance and output of the compartment so that closed gas loop was established by using a diaphragm vacuum pump (GAST, 22D1180-202-1005, Benton Harbor, Michigan, United States).

After completion of previous tests, involving Compartments 3 and 4a in the liquid phase, a new series of experiments was carried out. Now, the animal isolator was connected in the gas phase to the PBR, and liquid connection between nitrifying bioreactor and PBR was maintained (see **Figure 3**). Therefore, the three compartments were connected. In this scenario, the liquid flow-rate was increased to 20, 30 and 40 $\text{L}\cdot\text{d}^{-1}$ and the $\text{N}\text{-NH}_4^+$ concentration was maintained at 300 ppm at 20 and 30 $\text{L}\cdot\text{d}^{-1}$, while it was decreased to 240 ppm at 40 $\text{L}\cdot\text{d}^{-1}$. The main control variable of these experiments was the light intensity of the PBR. The control system adjusted the light intensity to maintain an O_2 concentration set-point in the animal isolator and compensating the animals oxygen needs along the day/night cycles. For each liquid loop flow rate condition, the O_2 set-point in the isolator was sequentially changed following this

sequence: 21–19–20–21%. The rationale between this operation schedule was to test the capacity of the control system to adjust the illumination conditions in the PBR each time a new O₂ set-point was fixed in the animal isolator. Additionally, the experimental period at a given flow rate between the nitrifying bioreactor and PBR coincided with a new group of rats in the isolator.

Biomass Concentration Determination

Off-line monitoring of biomass in the PBR was carried out by cell dry weight (CDW) determination and OD measurement. For CDW, 25–50 ml of culture broth were filtered through a 47 mm-glass microfiber (GMFC-52047, Scharlab, Barcelona, Spain), dried at 105°C until constant weight. OD measurement was performed at 750 nm in an optical spectrophotometer (UV-VIS DR6000, Hach, Düsseldorf, Germany).

TIC Total Inorganic Carbon and Nitrogen Determination

Filtered fractions for biomass determination in the PBR were used for NO₃⁻ and TIC analysis. Samples were diluted 1:100 with milli-Q H₂O and NO₃⁻ was analysed by spectrophotometry with LCK 339 kits (Hach, Weinheim, Germany). For TIC determination a 1:5 dilution with milli-Q H₂O was required. 1.5 ml of sample were loaded to a multi N/C2100S equipment (Analytik Jena, Jena Germany). 200 µL from the sample were injected into the acidic TIC reactor and 0.5 ml of H₃PO₄ at 10% were added to the chamber. CO₂ generated from the acidification was detected by the NDIR detector and quantified by triplicate.

In Compartment 3, NO₃⁻ at the bioreactor outlet was analysed following the same method as in Compartment 4a. NH₄⁺ and NO₂⁻ were analysed with LCK 303/304 and LCK 342 (Hach, Weinheim, Germany).

Gas Analysis, O₂ Production and CO₂ Consumption

Outlet gas composition from the PBR was measured on-line by an IR analyser for CO₂ coupled to a paramagnetic analyser for O₂ (600 series CAI, Orange, California, United States). In the case of Compartment 4a and 3 liquid connection, oxygen volumetric production rate (g·L⁻¹·h⁻¹) was calculated on-line based on continuous monitoring of outlet gas composition in the PBR:

$$r_{O_2} = ((F_{in} - F_{CO_2}) \cdot 0.2089 - (F_{out} \cdot O_{2out}/100)) / (22.4 \cdot MW_{O_2} \cdot VR) \quad (1)$$

where F_{in} is the inlet air flow rate, F_{CO_2} is the inlet CO₂ flow rate, F_{out} is the total outlet gas flow rate, O_{2out} is the outlet mole fraction of O₂ in %, MW_{O_2} is the molecular weight of O₂ (32 g·mol⁻¹). The flow rates are expressed at normal conditions (T = 273.15 K; p = 1 atm). In the case of the second integration step (Compartments 5 and 4a connected in the gas phase and 3 and 4a connected in the liquid phase), the inlet gas composition of PBR is analysed by means of the gas

analyser in compartment 5. Then, oxygen volumetric production rate in PBR is calculated as follows:

$$r_{O_2} = ((F_{in} \cdot O_{2in} - (F_{out} \cdot O_{2out}/100)) / (22.4 \cdot MW_{O_2} \cdot VR) \quad (2)$$

where O_{2in} is the inlet mole fraction of O₂ (%) in the PBR.

PBR CO₂ consumption was calculated for each condition considering on-line gas analysis and off-line TIC concentration in the steady state. In such situation, no carbon accumulation takes place and CO₂ can be calculated in g·L⁻¹·h⁻¹ by means of the following mass balance:

$$r_{CO_2} = CO_{2outlet} - (CO_{2gas\ transfer}) \quad (3)$$

$$r_{CO_2} = C_{Lout} \cdot D + F_{in} (X_{CO_{OUT}} - X_{CIN}) / 22.4 \cdot MW_{CO_2} \quad (4)$$

where C_{Lout} is the carbon concentration (g·L⁻¹) in the liquid outlet converted into CO₂ by means of molecular weight, D is the dilution rate (h⁻¹), F_{in} (L·h⁻¹) is the bioreactor gas flow rate, $X_{CO_{OUT}}$ is the molar fraction of CO₂ in the outlet gas, X_{CIN} is the molar fraction of CO₂ in the inlet gas and MW_{CO_2} is the molecular weight of O₂ (44 g·mol⁻¹). When PBR and animal compartment were connected in the gas phase, X_{CIN} was the corresponding CO₂ concentration in the animal compartment.

Considering that O₂ was the controlled variable in the animal compartment, O₂ consumption in animal compartment was assumed to be the same as O₂ production in the PBR. In the case of CO₂ consumption (g·h⁻¹) of the rats, calculation was as follows:

$$r_{CO_2} = F_{in} (X_{CO_{OUT5}} - X_{CIN5}) / 22.4 \cdot MW_{CO_2} \quad (5)$$

where $X_{CO_{OUT5}}$ and X_{CIN5} are the molar fractions of CO₂ at the outlet and inlet of animal compartment respectively.

Statistical Analysis

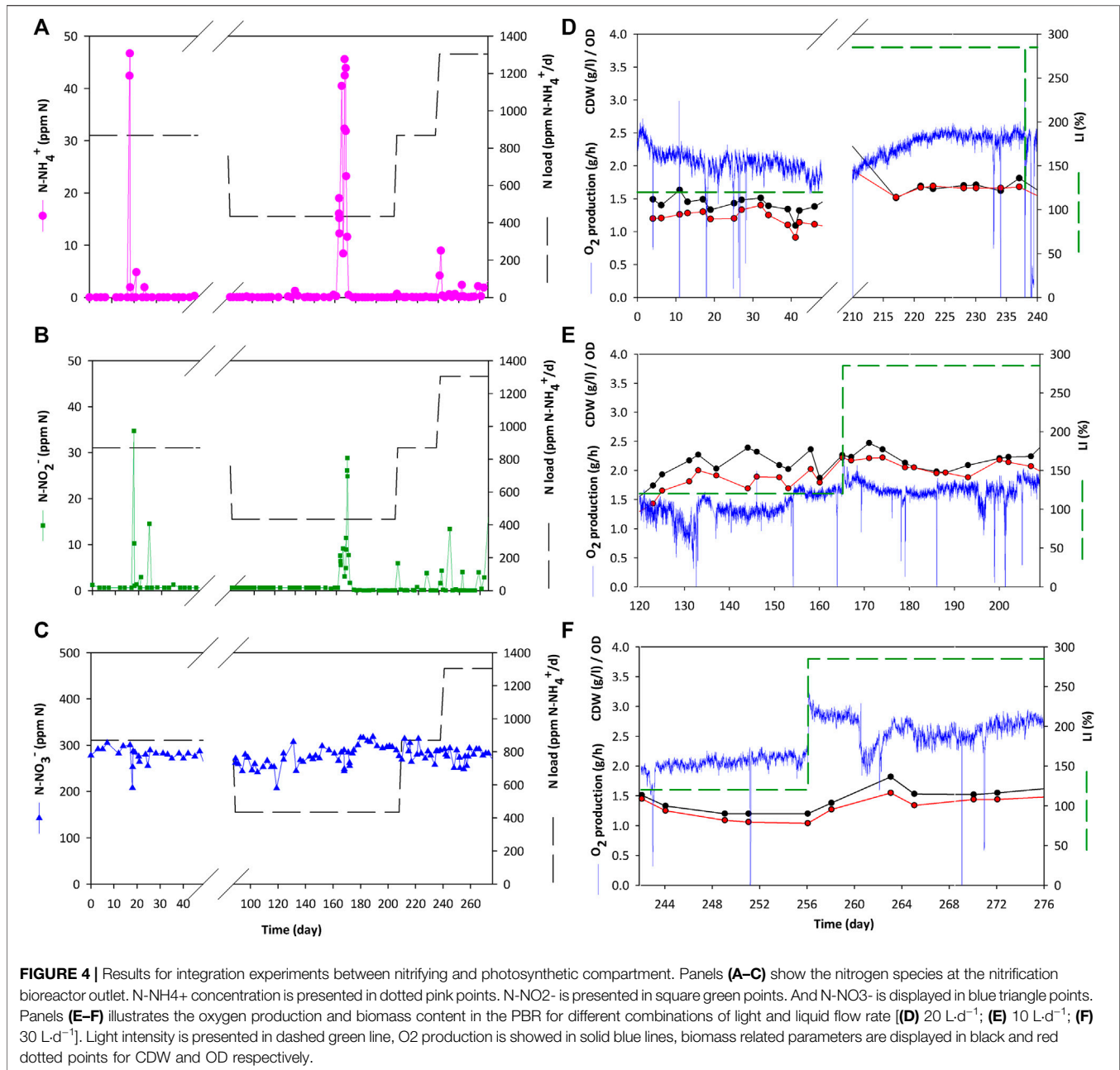
One-way ANOVA analysis assuming differences in variances was performed to assess variability of O₂ generation of the system. This statistical approach was also used to study the differences in body weight of the animals. A significance value of $p < 0.05$ was considered in all cases.

RESULTS

Compartment 3 and 4a Liquid Connection

Nitrifying bioreactor and PBR are connected in the liquid phase in continuous operation mode for a total period of 276 days. The nitrification compartment provides the necessary nitrogen source for the growth of cyanobacteria (Clauwaert et al., 2017). A total of six different conditions were tested corresponding to three different ammonium loads: 435, 870, and 1300 ppm·d⁻¹ into Compartment 3 at 10, 20, and 30 L·d⁻¹ respectively. These flow rate conditions are combined with two levels for q_o in the PBR: 120 and 285 W·m⁻².

Figures 4D–F presents the operation profile of the PBR in terms of oxygen production and biomass cell density for the complete series of experiments. In all the tested conditions steady state in the PBR is achieved as it is maintained with stable parameters for a period of at least 3 Hydraulic Residence



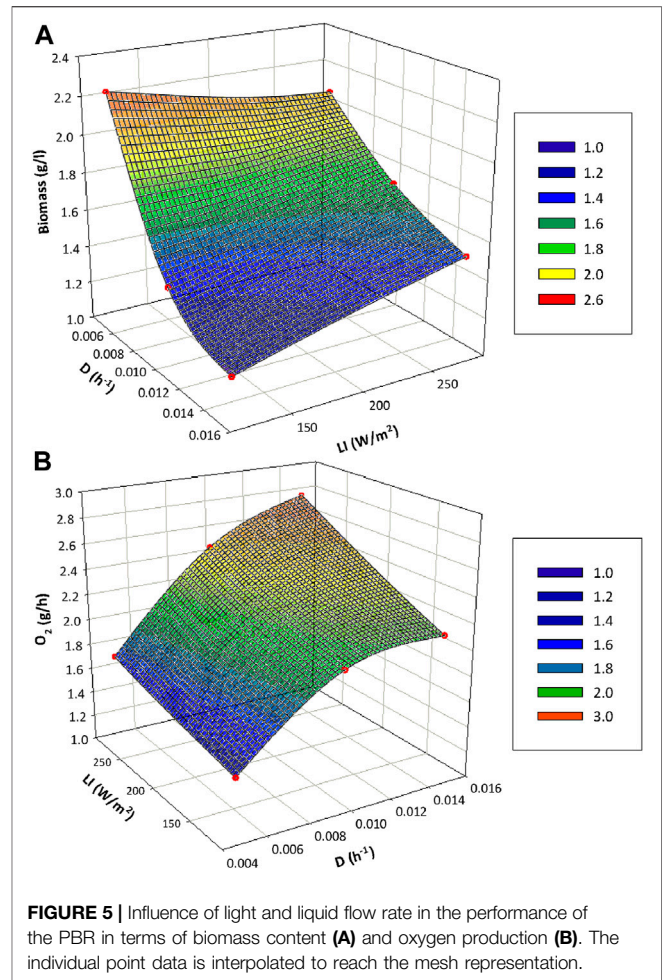
Times (HRT) for each condition. The highest biomass concentration is obtained for 10 L·d⁻¹ condition (Figure 4E) with values between 2.1 and 2.2 g·L⁻¹, while the lowest cell density is reached at 30 L·d⁻¹ (Figure 4F) with values between 0.9 and 1.1 g·L⁻¹. On the contrary, the highest biomass productivity results from 30 L·d⁻¹ conditions, followed by 20 and 10 L·d⁻¹ respectively. On-line oxygen production monitoring is also presented in Figure 4. The dynamics of the system can be observed for each q_o increase followed by a stabilisation profile. The highest oxygen production is reached at 30 L·d⁻¹ condition (between 2.1 and 2.7 g·h⁻¹) while the lowest production occurs at 10 L·d⁻¹ (1.4–1.7 g·h⁻¹). The same pattern is also observed from specific production values (q_{O_2}). Detailed data are presented in

Table 1. Figure 5 reflects the influence of light and flow in the performance of the PBR in terms of oxygen and biomass production. A Pareto's ANOVA was performed to study the influence of those factors. The liquid flow is the factor showing the greatest effect on biomass and oxygen production. The influence of flow on biomass concentration is 85%, while it is 69% for oxygen production. In contrast, light intensity shows little influence in biomass concentration (4%), while its contribution increases up to 21% regarding oxygen production. The interaction effect between factors in both cases is at 11 and 10% respectively.

Values from carbon dioxide consumption in the PBR are also presented in Table 1, showing that carbon dioxide consumption

TABLE 1 | Tested conditions and results in PBR for liquid connection between Compartment 3 (nitrification) and 4 (PBR).

Condition	Inlet NH ₄ ⁺ (N-ppm)	N-NH ₄ ⁺ load (ppm·d ⁻¹)	q _o (W·m ⁻²)	Liquid flow (L·d ⁻¹)	CDW (g·l ⁻¹)	Prod _x (g·h ⁻¹)	Prod _{o₂} (g·h ⁻¹)	qO ₂ (mmol·g ⁻¹ ·h ⁻¹)	Cons _{CO₂} (g·h ⁻¹)	qCO ₂ (mmol·g ⁻¹ ·h ⁻¹)	Q _P (mol _{O₂} ·mol _{CO₂} ⁻¹)
#1	300	870	120	20	1.41 ± 0.12	1.18 ± 0.10	2.03 ± 0.15	0.54	-1.55	-0.30	1.80
#2	300	435	120	10	2.24 ± 0.15	0.93 ± 0.06	1.43 ± 0.18	0.24	-1.51	-0.18	1.30
#3	285	870	285	10	2.15 ± 0.15	0.89 ± 0.06	1.6 ± 0.14	0.30	-1.61	-0.21	1.44
#4	285	1300	285	20	1.70 ± 0.07	1.42 ± 0.06	2.44 ± 0.13	0.54	-1.95	-0.31	1.72
#5	285	1300	120	30	1.20 ± 0.02	1.50 ± 0.02	2.11 ± 0.08	0.66	-1.70	-0.39	1.71
#6	285	1300	285	30	1.50 ± 0.08	1.87 ± 0.10	2.76 ± 0.10	0.69	-2.33	-0.43	1.63

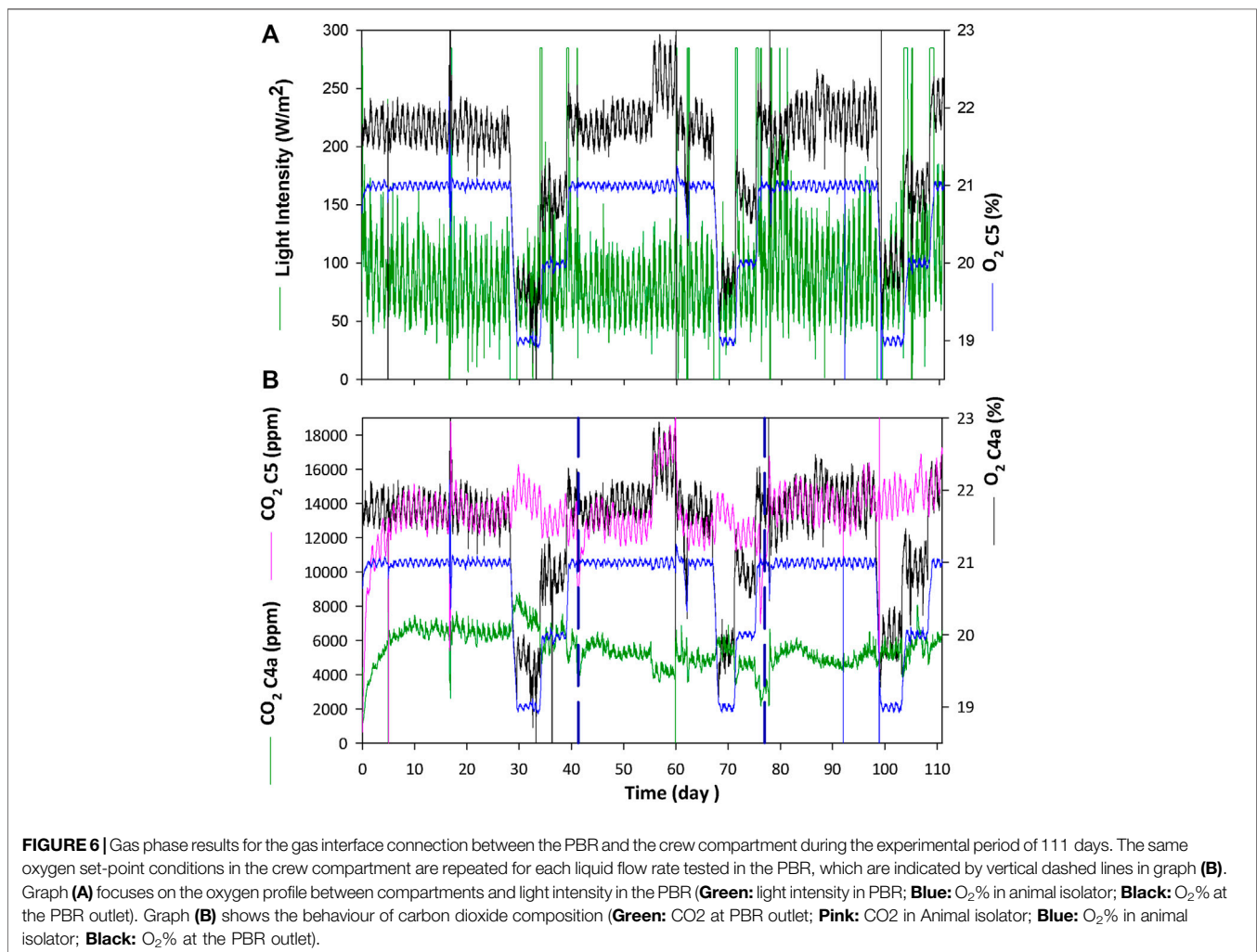


by *L. indica* increases in correspondence with oxygen production. 2.3 g CO₂·h⁻¹ are consumed when the cells are exposed to the maximum q_o of 285 W·m⁻² at a continuous flow of 30 L·d⁻¹. The minimum carbon dioxide consumption is found at the lowest flow rate of 10 L·d⁻¹ and minimum light intensity (120 W·m⁻²). Photosynthetic quotient ranges between 1.3 and 1.8 molO₂·molCO₂⁻¹, which is in agreement with theoretical values between 1.2 and 1.4 when NO₃⁻ is used as nitrogen source according to the stoichiometry (Cornet, 2007).

In terms of N balance, all the nitrogen supplied to compartment 4 is in form of NO₃⁻ previously converted from NH₄⁺ in compartment 3. The ammonium conversion is maintained at 99% during most of the experiment (Figures 4A–C). Only minor transitory peaks occur because of operation activities in the bioreactor focused on biomass removal with an external circulation system coupled with filtration for biomass retention. During these short periods the maximum N-NH₄⁺ and N-NO₂⁻ concentration in the PBR is 0.1 and 1 ppm respectively (data not shown) and therefore can be considered negligible. The outlet N-NO₃⁻ concentration from the nitrifying bioreactor is between 275 and 300 N-ppm (Table 2). Yield values are higher than 90%, whilst conversion of ammonium is at 99% during all steady states (transitory peaks

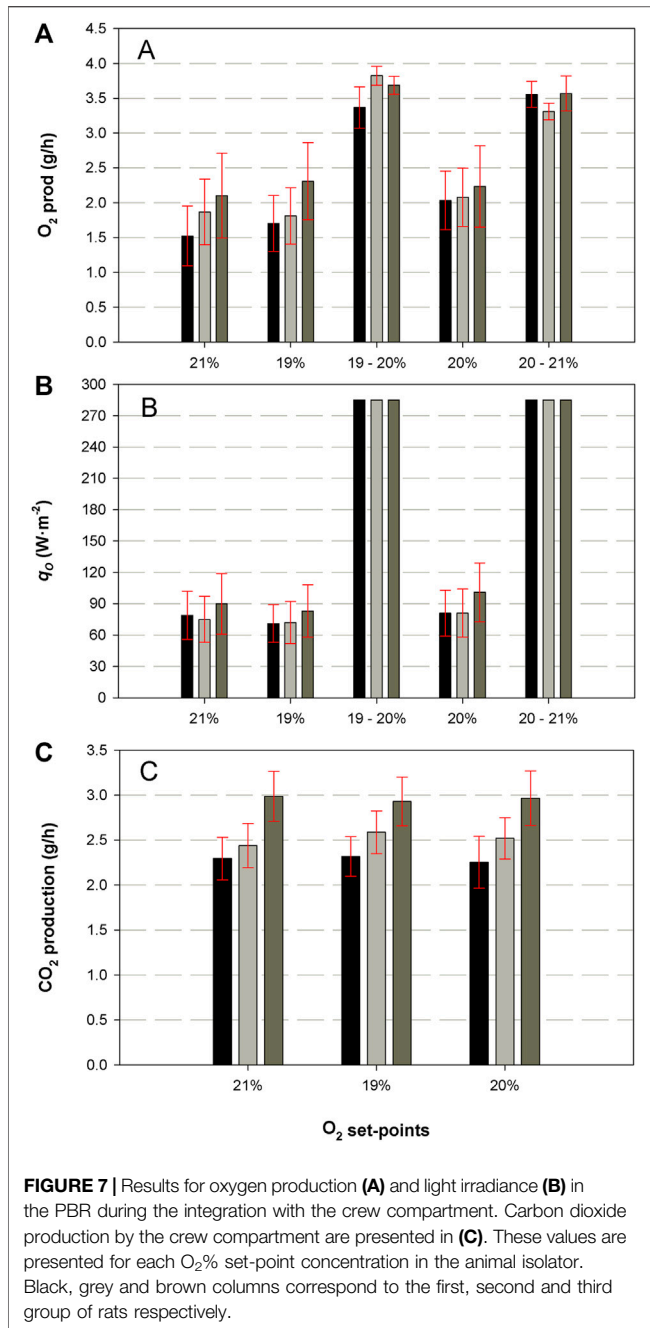
TABLE 2 | Nitrogen balances observed in nitrifying bioreactor for every experimental condition during liquid connection experiments between nitrifying and photosynthetic compartment. The outlet of NO_3^- from nitrifying bioreactor is considered as the inlet concentration in the PBR.

Cond	Nitrifying compartment						PBR			
	Inlet NH_4^+ (N-ppm)	Load (N-ppm-d ⁻¹)	NO_3^- outlet (N-ppm)	NH_4^+ outlet (N-ppm)	NH_4^+ removal (%)	Conversion (%)	NO_3^- outlet (N-ppm)	$Y_{N/X}$ (g-g ⁻¹)	Y_{N/O_2} (g-g ⁻¹)	
#1	300 N- NH_4^+	870	274.8	0.01	100.00	91.6	116.06	0.11	0.065	
#2		435	288.5	0.05	99.98	96.2	66.35	0.10	0.064	
#3				297.7	0.02	99.99	99.2	65.71	0.11	0.058
#4		870	279.1	0.10	99.97	93.0	95.15	0.11	0.063	
#5		1300	274.7	0.20	99.93	91.6	152.80	0.10	0.072	
#6				276.8	1.00	99.67	92.3	121.60	0.10	0.070



are not considered). The yield is higher (96–99%) with the lowest ammonium load of 435 N-ppm-d⁻¹, but the yield decreases (91–93%) when the load increases to 870 and 1300 ppm-d⁻¹. Inlet N-NH₄⁺ into nitrifying bioreactor was maintained constant at 300 N-ppm for all conditions, resulting in different ammonium loads depending on the liquid flow rate. The output from

nitrifying bioreactor was the inlet of the PBR, so $Y_{N/X}$ was calculated considering that input. Results show that $Y_{N/X}$ is maintained constant no matter the experimental condition used. 0.10–0.11 g of N-NO₃⁻ are needed to generate 1 g of biomass. In terms of oxygen as a product, Y_{N/O_2} is between 0.06 and 0.07 g-g⁻¹ in all tested conditions. The yields obtained fit



with the reaction stoichiometry described in previous works (Cornet et al., 1998).

Compartment 3 and 4 Liquid Connection Combined with Compartment 4a and 5 Gas Connection Gas Phase Performance

The connection between the PBR and the animal isolator was conducted during 110 days. Three female Wistar rats were connected in the gas phase with the PBR, which provided the oxygen needed in the animal isolator to maintain the oxygen

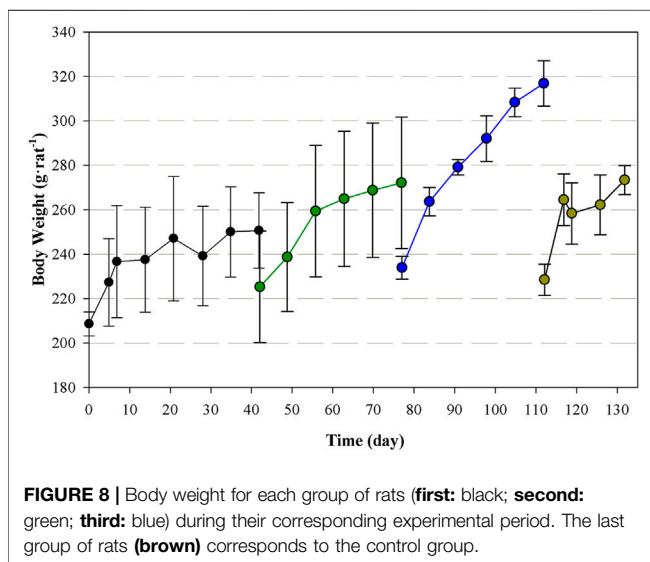
atmospheric concentration at the target value (21–19 to 20–21%). On the other hand, the carbon dioxide produced by the rats was supplied to the PBR to cover the carbon needs for photosynthetic activity. In parallel, the nitrogen needs of the PBR were supplied by the nitrifying bioreactor after complete removal of ammonium and conversion into nitrate. Different liquid flows between nitrifying bioreactor and the PBR (20–30–40 L·d⁻¹, each one with a different group of rats in the animal isolator for age reasons) were tested in order to assess the effect of hydraulic changes in the PBR.

Figure 6A provides the gas phase evolution of the animal isolator and the PBR. The oxygen concentration of the isolator is controlled first at the set-point of 21% following the sequential changes (21–19 to 20–21%) for each animal group. A repetitive cyclic pattern in the oxygen concentration profile is observed, caused by the circadian activity (day and night cycles) of the rats. Light irradiance in C4a changes likewise to adapt the oxygen production to the animals needs. Changes in oxygen set-point concentration are made when 6 HRT are reached in the PBR. During oxygen set-point decrease (from 21 to 19%), the control system turns off the lights and it takes 30, 26 and 20 h for the first, second and third group of rats respectively to reach the target value. In transition periods where the oxygen set-point is increased from 19 to 20%, the lights are set by the control system to the maximum intensity (285 W·m⁻²) and it takes 8, 7.8 and 19.5 h for each group respectively to reach the target set-point. Finally, when oxygen set-point is increased from 20 to 21% the transition takes 8.5, 8.3 and 19.8 h respectively. **Figure 6B** focuses in the carbon dioxide concentration of the animal isolator and the PBR. Carbon dioxide in the animal isolator is kept between 12,000 and 16,000 ppm (1.2–1.6%) during all the test, except for the period between day 55–60, when it reaches 18,000 ppm because of a perturbation in the gas flow exchange between the PBR and animal isolator. However, normal values are recovered afterwards. The cyclic biological activity of the rats during day and night is also observed in carbon dioxide dynamics.

Results for oxygen production in the PBR, carbon dioxide production by the crew and light intensity for each group of animals and for each oxygen concentration in the isolator are presented in **Figures 7A–C**. The average oxygen needs of the rats are different for each group and consequently the oxygen production. The average oxygen production by the PBR is 1.5, 1.9 and 2.1 g·h⁻¹ for the first, second and third group respectively during the 21% set-point. The same pattern is observed for 19 and 20% concentration. Regarding carbon dioxide generation of the rats, 2.3, 2.4 and 3.0 g·h⁻¹ are produced by each group at 21% O₂ concentration. RQ of the rats at 21% is 1.0, 0.9 and 0.97 mol_{O₂}·mol_{CO₂}⁻¹ for each group (data not shown). Average light intensity (\bar{q}_o) provided by the control system to the PBR does not correlate with oxygen production as \bar{q}_o in the second group (75 W·m⁻²) is lower than in the first group (79 W·m⁻²). During the third group \bar{q}_o increases to 90 W·m⁻². Focusing on transition periods (19–20% and 20–21%) where the \bar{q}_o is at maximum, the oxygen generation in the PBR are statistically different in both transitions ($p < 0.05$). During the

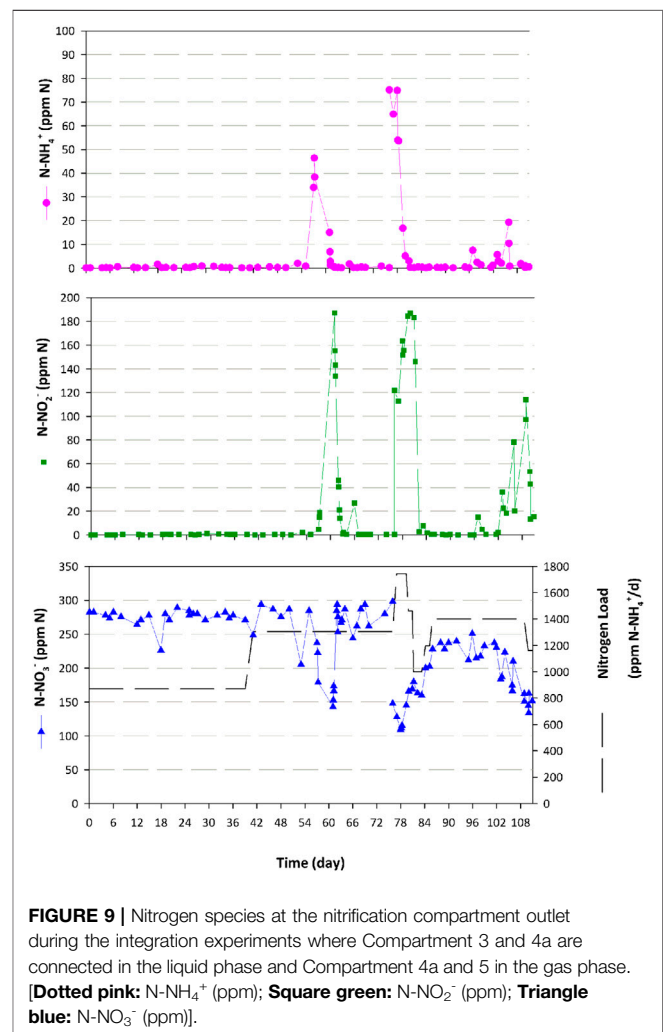
TABLE 3 | PBR performance results in terms of O₂ and CO₂ generation and biomass concentration reached for every condition. CO₂ uptake is only expressed at 21% O₂ set-point as liquid phase is not stable at 19 and 20% leading to variability in TIC concentration.

Rat's group	O ₂ SP (%)	q _o (W·m ⁻²)	X (g·L ⁻¹)	O ₂ prod (g·h ⁻¹)	CO ₂ uptake (g·h ⁻¹)	qO ₂ (mmol·g ⁻¹ ·h ⁻¹)	qCO ₂ (mmol·g ⁻¹ ·h ⁻¹)	PQ (rO ₂ :rCO ₂ ⁻¹)
Liquid flow								
#1	21	79	1.24	1.52	2.18	0.46	0.48	0.96
—	19	71	1.07	1.70	—	0.60	—	—
20 L/d	20	78	1.22	1.81	—	0.63	—	—
#2	21	75	0.98	1.87	1.92	0.72	0.54	1.33
—	19	72	0.77	1.81	—	0.89	—	—
30 L/d	20	81	0.89	2.08	—	0.88	—	—
#3	21	90	0.81	2.10	2.08	0.98	0.71	1.39
—	19	83	0.72	2.31	—	1.21	—	—
40 L/d	20	101	0.83	2.23	—	1.01	—	—



transition from 19 to 20%, the oxygen production is 3.4, 3.8 and 3.7 g·h⁻¹ for the first, second and third groups respectively. In the second transition step the oxygen generation is 3.6, 3.3 and 3.6 g·h⁻¹.

Table 3 shows the results obtained in the PBR. Biomass concentration is different depending on the group (and flow) tested. In the steady state, which takes place at 21% oxygen set-point, the highest concentration is 1.24 g·L⁻¹ when the system works at 20 L·d⁻¹. The lowest concentration, 0.81 g·L⁻¹ is observed in the last group of rats when inlet liquid flow rate in the PBR is 40 L·d⁻¹. Changes within the same group are also observed for each oxygen set-point. Carbon dioxide uptake is only presented at 21% set-point condition as stability of TIC concentration in the liquid phase of the PBR is only achieved at 21% after 6 HRT. Maximum carbon dioxide demand is 2.18 g·h⁻¹ at 20L·d⁻¹ (first group). Similar uptake is observed for the other tested groups (1.92 and 2.08 g·h⁻¹). Specific oxygen production rates and carbon dioxide consumption rates (q_{O₂} and q_{CO₂}) increases for each group of rats



sequentially, indicating a higher efficiency in oxygen production, but also a higher carbon dioxide demand. PQ ranges between 1 and 1.4.

TABLE 4 | Nitrogen balances observed in nitrifying bioreactor and photobioreactor for every experimental condition during liquid connection between nitrifying and photosynthetic compartments and gas connection between photosynthetic and crew compartment. The outlet of NO_3^- from nitrifying bioreactor is considered as the inlet concentration in the PBR.

Liquid flow (L·d ⁻¹)	Nitrifying compartment					PBR		
	Load (N·ppm·d ⁻¹)	NO_3^- inlet (N·ppm)	NO_3^- outlet (N·ppm)	NH_4^+ removal (%)	Conversion (%)	NO_3^- outlet (N·ppm)	$Y_{N/X}$ (g·g ⁻¹)	Y_{N/O_2} (g·g ⁻¹)
20	870	300	274.8	99	91.6	160	0.09	0.064
30	1,304	300	289	99	96.3	202	0.09	0.059
40	1,400	241	227	99	94.2	155	0.09	0.057

Finally, the daily monitoring of the rats (**Figure 8**) shows that the weight gain differs between groups. Average weight gain of the first group is $0.77 \text{ g}\cdot\text{d}^{-1}\cdot\text{rat}^{-1}$, followed by the second group ($1.35 \text{ g}\cdot\text{d}^{-1}\cdot\text{rat}^{-1}$) and the third group ($1.30 \text{ g}\cdot\text{d}^{-1}\cdot\text{rat}^{-1}$). The fourth group of rats, which corresponds to the control group, showed a weight gain of $1.75 \text{ g}\cdot\text{d}^{-1}\cdot\text{rat}^{-1}$. Pearson's correlation shows a positive correlation between the starting weight of the rats and the weight gain rate ($r = 0.95$; $p < 0.05$). All the animal groups did not show any relevant alteration in their physiological parameters, behaviour or clinical evaluation. Food and water intake are normal in all the groups (data not shown).

Liquid Phase Results

As mentioned, nitrogen in form of NO_3^- was provided to the PBR through the liquid phase, which was connected to the outlet of the nitrifying bioreactor. Nitrification profile during the whole experiment is presented in **Figure 9**. Ammonium conversion is maintained at 100% and no accumulation of nitrite nor ammonium takes place during the first hydrodynamic condition ($20 \text{ L}\cdot\text{d}^{-1}$) at $870 \text{ ppm}\cdot\text{d}^{-1}$. In the second experimental period ($30 \text{ L}\cdot\text{d}^{-1}$) a peak of nitrite ($187 \text{ N}\cdot\text{ppm}$) and ammonium ($46 \text{ N}\cdot\text{ppm}$) is measured at the outlet of nitrifying bioreactor on day 60, probably caused by the biomass removal in the packed-bed as a nominal operation (excess biomass accumulation is removed periodically from the packed-bed to avoid clogging). Recovery of the complete nitrification takes place within 6 days. In the last hydraulic condition ($40 \text{ L}\cdot\text{d}^{-1}$) the tested nitrogen load of 1740 results in ammonium ($78 \text{ N}\cdot\text{ppm}$) and nitrite ($186 \text{ N}\cdot\text{ppm}$) accumulation at the bioreactor outlet, therefore showing that the maximum load capacity was achieved. Then, the system is recovered after decreasing the nitrogen load to $1400 \text{ ppm}\cdot\text{d}^{-1}$, reaching total nitrification as in previous conditions. Finally, at the end of the same hydraulic condition, a third and last nitrite peak ($113 \text{ N}\cdot\text{ppm}$) occurs due to a second operation to control biomass in the bioreactor to avoid clogging. Those peaks are translated in a slight presence of nitrite in the PBR: maximum concentration of 78, 58 and $36 \text{ N}\cdot\text{ppm}$ respectively. No presence of ammonium is detected in the PBR. In the first and second cases, the presence of nitrite in the PBR is translated into a light intensity increase reaching maximum q_o during night periods (data not shown), except in the last group.

Nitrogen balances during steady state of the PBR (corresponding to O_2 concentration in the animal isolator at 21%) are presented in **Table 4**. Ammonium is not detected in the liquid outlet of the nitrifying bioreactor, so ammonium removal is at 99% for all conditions. Conversion of ammonium into nitrate is at 91.6, 96.3 and 94.2% for each condition respectively, which is in line with the performance of the bioreactor in previous integration experiment (*Compartment 3 and 4a Liquid Connection*). The outlet nitrate concentration in the PBR depends on the hydraulic condition. Higher concentration is measured at $30 \text{ L}\cdot\text{d}^{-1}$ ($202 \text{ N}\cdot\text{ppm}$) than at $20 \text{ L}\cdot\text{d}^{-1}$ ($160 \text{ N}\cdot\text{ppm}$). It is not the same behaviour as at $40 \text{ L}\cdot\text{d}^{-1}$ ($155 \text{ N}\cdot\text{ppm}$) because of the inlet concentration reduction to diminish the ammonium⁺ load into the system. Results show that the same $Y_{N/X}$ is obtained in all conditions. 0.09 g of $\text{N}\cdot\text{NO}_3^-$ are needed to generate 1 g of biomass. In contrast, around 0.06 g of $\text{N}\cdot\text{NO}_3^-$ are needed to generate 1 g of oxygen. These values are in the same range as those obtained in the first series of experiments.

DISCUSSION

Compartment 3 Ad 4a Liquid Connection

The obtained results prove that liquid connection between nitrifying and photosynthetic bioreactors was successful. Nitrogen requirements of *L. indica* were met thanks to the nitrate supplied by the nitrifying bioreactor after a complete nitrification process performed by the co-culture of *N. europaea* and *N. winogradsky*. Indeed the operation of the photobioreactor was very stable and robust in the range of $10\text{--}30 \text{ L}\cdot\text{d}^{-1}$ and q_o of $120 \text{ W}\cdot\text{m}^{-2}$ and $285 \text{ W}\cdot\text{m}^{-2}$. Stability for each condition was maintained at steady-state to obtain reliable data. In terms of nitrification activity, the system has been proved to be robust enough to maintain the ammonium conversion at 100% for 200 days no matter the load. There were minor accumulation peaks that were rapidly recovered and had no impact on photosynthetic activity of *L. indica*. These peaks were most likely caused after biomass removal from the packed-bed reducing the amount of active biomass. Then, once the previous active levels of biomass were reached, the nitrification activity recovered to 100%. Additionally, residual $\text{N}\cdot\text{NO}_2^-$ concentration in the PBR supernatant was below the limit for drinking water (World Health Organization, 2017).

The effect of light and dilution rate have been proved to contribute in a different degree to the performance of the PBR. Liquid flow, and consequently dilution rate, is the factor having the greatest effect on biomass and oxygen productivity. It is an indication that what really influences the behaviour of cyanobacteria is light availability rather than the amount of light. **Figure 5** shows that light availability was directly proportional to the amount of light, but inversely proportional to the amount of cells. Results in terms of oxygen production were very relevant for the definition of the next integration step including the gas connection of the animal compartment to the PBR. Animal compartment in the MELISSA loop aims to mimic the human activity. In this case the rats are the mock crew. As reported previously (Alemany et al., 2019), the oxygen needs for 1 rat is 0.48–0.7 g·h⁻¹. The analysis of the obtained results for Compartment 3 and 4a liquid connection shows that depending on the liquid flow between bioreactors, different number of rats could be used in the experimental connection with the animal compartment. When working at 10, 20 or 30 L·d⁻¹, the maximum oxygen productions achieved in the steady state were 1.6, 2.44 and 2.76 g·h⁻¹ respectively. Under this scenario, the maximum number of rats that could be selected in the animal compartment is 4 with a theoretical minimum and maximum oxygen demand of 1.92 and 2.8 g·h⁻¹ respectively. Nevertheless, under this scenario the system would be at the maximum capacity to meet the maximum oxygen requirements. It is for this reason that 3 rats were selected for the experiments where nitrifying bioreactor and the PBR were connected in the liquid phase, and the PBR was connected through the gas phase with animal compartment. This solution provides some margin to the system to react against potential perturbations or changes imposed by the user in the gas phase composition of the animal isolator. The selection of 3 rats imposes a theoretical oxygen demand into the system of 1.44–2.1 g·h⁻¹ for minimum and maximum values. In order to meet this demand, the minimum and maximum carbon dioxide needs of the PBR would be 1.32 and 1.93 g·h⁻¹ respectively assuming a middle range experimental Q_p of 1.5 molO₂·molCO₂⁻¹ (**Table 1**). The animal compartment would provide 1.78–2.60 g·h⁻¹ of carbon dioxide if a theoretical RQ of 0.9 molCO₂·molO₂⁻¹ is considered (McGregor and Lee, 1998). So, the carbon needs of the PBR would be met. In terms of N needs, no lack of nitrate is expected in the PBR to meet the O₂ requirements if the same N-NH₄⁺ loads in nitrifying bioreactor are maintained. It has been demonstrated that according to yields displayed in **Table 2**, the amount of N-NO₃⁻¹ needed in the PBR would be between 0.09 and 0.12 g·h⁻¹. Hence, any of the 3 tested loads conditions in the nitrifying bioreactor are acceptable.

According to previous rationale, the best hydrodynamic conditions for the next integration phase would be selecting a liquid flow of 20 and 30 L·d⁻¹ between nitrifying bioreactor and the PBR. The liquid flow of 10 L·d⁻¹ should be discarded as the system could be in the limit in terms of oxygen production, especially during high activity periods of the rats. Additionally, the exploration of a wider experimental liquid flow domain could provide of relevant information for further optimisation of the

MELISSA loop. This is why a liquid flow of 40 L·d⁻¹ was also selected as an experimental condition for the second phase tests.

Compartment 3 and 4 Liquid Connection Combined With Compartment 4a and 5 Gas Connection

After a 110 days experiment, where the PBR and the animal isolator were connected in the gas phase, it has been demonstrated that the system is able to cover the oxygen needs of the rats and carbon needs of *L. indica* while operating in a closed gas loop. This condition is in agreement with previous results (Alemany et al., 2019). The added value of the current work is that nitrogen requirements of *L. indica* to cover the oxygen demand, have been covered by the continuous operation of a nitrifying bioreactor. This is particularly relevant since it represents a further step in the completion of the overall closed-life support system.

Focusing on the gas phase dynamics, the system could maintain a constant oxygen concentration in the animal isolator. Oscillations ($\pm 0.04\%$) in the measured value are the direct cause of circadian activity of the rats. In all oxygen set-points (21, 19, 20%), the activity of the rats was higher during night periods because of its photoperiod (Antle and Mistlberger, 2005). In all cases the RQ of the rats ranged between 0.9 and 1.0 molO₂·molCO₂⁻¹, which is consistent with the experimental data reported in the literature (McGregor and Lee, 1998; Jørgensen et al., 2010).

Transition periods, where oxygen set point was increased, took less than 10 h in two groups of rats and less than 20 h in the last group. This proves the fast response of the system to adjust to a change in the parameters of the crew compartment. It should be reminded that the aim of Compartment 4a (PBR) in the loop is to respond to oxygen demand variations thanks to its fast dynamics. The explanation of a higher time period for the transition in the last group of rats is the higher oxygen demand of the rats. It was demonstrated by the average oxygen produced in the PBR for each rat's group (**Figure 7**), which increased up to 2.1 g·h⁻¹ in that case. Indeed, the body weight gain of the rats, which could be an indication of the physiological activity of the rats, has been correlated with the oxygen demand of the system. In contrast, the transition periods for the first group of rats was not lower than in the second group, even a lower oxygen demand (1.5 vs. 1.9 g·h⁻¹). This effect could be explained by the differences in the PBR capabilities. In the first case, the biomass concentration in the PBR was higher than in the second case (1.24 g·L⁻¹ vs. 0.98 g·L⁻¹), potentially limiting the light availability to the cells and decreasing the photosynthetic activity at the same light irradiance. This hypothesis is plausible considering the average \bar{q}_o , which is higher in the first group (79 W·m⁻²) than in the second the group of rats (75 W·m⁻²) even the lower oxygen need. From this observation, it is confirmed that the cells were in a “physical limitation regime” where the light is the limitation parameter of the growth kinetics (Cornet, 2007). The higher light intensity observed in the last group could be explained by the high oxygen demand.

In terms of carbon dioxide concentration, the levels were always maintained below 15,000 ppm. This was below the acceptance limit of 3% for rats (Krohn and Hansen, 2002; Krohn et al., 2003). However, future experiments in the system should consider lowering this level and make it closer to the limits accepted for humans (<0.5%). This point will address in future integration steps of the MELISSA loop.

Results from liquid connection experiments between Compartment 3 and 4a showed that maximum oxygen production in the PBR was around 2.4–2.8 g·h⁻¹ with a flow rate of 20–30 L·d⁻¹. However, during the second phase of the experiments (liquid connection between Compartment 3 and 4a and gas connection between Compartment 4a and 5) the system has proved to be over the previous determined limits. Light irradiance was maintained around 30% of its capacity with enough margin to meet the oxygen demand during transition periods. Indeed, the maximum oxygen production in those transition phases exceeded by 25% the reported values of the first experimental phase. This difference evidences that transition phases should be treated differently than steady state. Comparing biomass concentration at maximum light intensity condition in both experimental phases shows that cell density is lower in the second phase, leading to a higher light availability. From a biological perspective this effect might be explained by the molecular composition of the cells. The photosynthetic capacity of *L. indica* is influenced by the pigment content. When light availability is increased, it takes some hours (Tomaselli et al., 1997) to reduce the pigment content to protect the cells against potential photoinhibition (Muramatsu and Hihara, 2012; Weiwen Zhang, 2018). It is during that transition that the closed loop benefits from the still high pigment content to exploit the maximum photosynthetic activity of the cells. It can be concluded that the capacity of the system in transition steps cannot be extrapolated at long-term, when the system would evolve to reach a new steady state with lower oxygen generation capacity.

Regarding nitrogen needs of *L. indica* to meet the oxygen requirements of the animal crew, it has been demonstrated that nitrifying bioreactor is capable to fulfil them. Indeed, nitrate concentration in the PBR outlet was always kept higher than 20 N-ppm, which is considered a potentially limiting concentration (Depraetere et al., 2015). This scenario avoided any metabolic drift in *L. indica*, allowing an optimal performance in terms of air revitalisation. However, the limiting capacity of the nitrifying bioreactor was reached in the last condition (40 L·d⁻¹), so ammonium load had to be reduced in order to avoid nitrogen species accumulation. One of the outputs from this work is that the maximum capacity of nitrifying bioreactor in the MELISSA loop is 1400 ppm·d⁻¹, which should be taken into consideration when it comes to introduce the urea in the loop. Once the ammonium load was decreased, the full nitrification capacity was fully recovered. Although complete nitrification was dominant during most of the test, three partial nitrification events occurred. Nitrite and ammonium reached the PBR resulting in different consequences. Whilst nitrite was present at detectable levels, no ammonium was detected at the PBR outlet. This result shows

that *L. indica* can use ammonium as nitrogen source as previously reported in the literature (Sachdeva et al., 2018). Even nitrite concentration in the PBR reached up to 78 N-ppm during peaks periods, no irreversible effects were detected in *L. indica* culture. In the first and second accumulation events (78 and 58 N-ppm), the presence of nitrite had an effect on photosynthetic activity. The control system adjusted the lights up to the maximum q_o of 285 W·m⁻² during 2–3 h in order to cover the oxygen needs of the animal compartment. This behaviour was not observed in the last accumulation event (36 N-ppm) suggesting that a concentration higher than 40 N-ppm of nitrite have an inhibitory effect on *L. indica* photosynthetic activity. Nevertheless, this event shows the robustness of the system in such a wide range of conditions and in front of potential perturbations. The main function of air revitalisation in Compartment 4a was never at risk and oxygen needs of the rats were fulfilled during 110 days of continuous operation.

CONCLUSION

The work presented here has demonstrated the feasibility of integrating three compartments of the MELISSA loop: Compartment 4a (photosynthesis) and 5 (crew compartment) in the gas phase and Compartment 3 (nitrification) and 4a in the liquid phase. The step-wise approach has been proved to be a key factor in the progress to demonstrate the MELISSA loop concept. Results from the first experimental period have been relevant to define the integration conditions for the following step. The system has succeeded to maintain its main functions to guarantee the survival of the animal crew for a long operational period, proving as well its robustness and reliability against perturbations, which is very relevant in the MELISSA loop context. It can be concluded that this experimental work represents a step forward in the demonstration of the MELISSA loop concept and its contribution to bioregenerative life support.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article material, further inquiries can be directed to the corresponding author.

ETHICS STATEMENT

The animal study was reviewed and approved by Committee on Ethics in Animal and Human Research (4017 CEEAH—UAB).

AUTHOR CONTRIBUTIONS

All authors listed have made a substantial, direct, and intellectual contribution to the work and approved it for publication.

FUNDING

The MELISSA Pilot Plant is funded from ESA contributions from Spain (main contributor), Belgium, France, Italy and Norway, under Frame Contract C4000109802/13/NL/CP. Co-funding from Ministerio de Ciencia e Innovación (RD 788/2020), Generalitat de Catalunya and Universitat Autònoma de Barcelona is also acknowledged.

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ACKNOWLEDGMENTS

MELISSA is an international consortium of 15 partners led by the European Space Agency (for information: <https://www.melissafoundation.org>). Its activities are governed by a Memorandum of Understanding (ESA 4000100293/10/NL/PA).

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NOMENCLATURE

C_{LOUT} outlet liquid CO₂ concentration (mmol·L⁻¹)

D dilution rate (h⁻¹)

F_{CO2} inlet CO₂ gas flow (L·h⁻¹)

F_{in} bioreactor inlet gas flow (L·h⁻¹)

F_{out} bioreactor total outlet gas flow (L·h⁻¹)

HRT Hydraulic Residence Time (days)

O_{2out} outlet O₂ fraction (%)

PQ Photosynthetic quotient (mol_{O₂}·mol_{CO₂}⁻¹)

q_O light irradiance in the PAR region (W·m⁻²)

q_{O2} specific O₂ production (mmol·g⁻¹·h⁻¹)

q_{CO2} specific CO₂ consumption (mmol·g⁻¹·h⁻¹)

r_{CO2} CO₂ volumetric consumption rate (g·L⁻¹·h⁻¹)

RQ respiratory quotient (mol_{CO₂}·mol_{O₂}⁻¹)

r_{O2} O₂ volumetric production rate (g·L⁻¹·h⁻¹)

r_X biomass volumetric production rate (g·L⁻¹·h⁻¹)

V_R bioreactor volume (L)

X biomass concentration (g·L⁻¹)

X_{CO2OUT} molar fraction of CO₂ in the outlet gas (mol·mol⁻¹)

X_{CI} molar fraction of CO₂ in the inlet gas (mol·mol⁻¹)