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# 1 Peptoid-phthalocyanine architectures with different grafting 2 positions: synthetic strategy and photoproperties

3  
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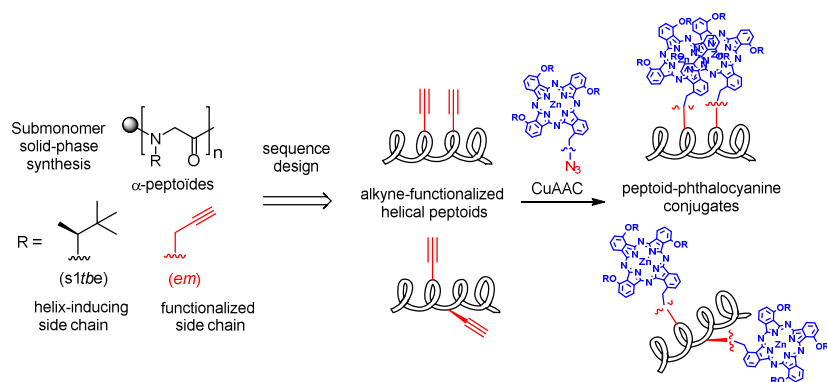
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15

## 16 Graphical abstract



17

18

## 19 Abstract

20 A monoazido Zn phthalocyanine has been grafted onto three different alkynyl-functionalized  
21 peptoid helices using copper-catalyzed azide–alkyne cycloaddition, demonstrating the  
22 feasibility of such conjugates. Their photoproperties have been examined relatively to the  
23 spatial geometry induced by the peptoid helix.

24

## 25 Keywords

26 Peptoid, phthalocyanine, helix, conjugate.

27 **Highlights**

28

29 • Peptoid chains with one and two alkynyl moieties on different residues have been  
30 prepared.

31 • A monoazido Zn phthalocyanine has been grafted using copper-catalyzed azide–  
32 alkyne cycloaddition

33 • The absorption and fluorescence of these conjugates have been recorded

34

## 35 1. Introduction

36

37 Porphyrinoids are aromatic macrocycles with an extended electronic delocalization,  
38 conferring sought-after electronic and spectroscopic properties, amongst others.<sup>1</sup> Many  
39 biomolecules such as chlorophylls,<sup>2</sup> hemes<sup>3</sup> and vitamin B12<sup>4</sup> are natural porphyrinoid  
40 derivatives. The most common porphyrinoids are porphyrins and phthalocyanines, used in  
41 several applications fitting the preoccupations of our modern world, such as artificial  
42 photosynthesis,<sup>5</sup> catalysis<sup>6</sup> and photocatalysis,<sup>7</sup> photosensitisers for photodynamic therapy,<sup>8</sup>  
43 conversion of light into electric current,<sup>9</sup> amongst others. Many of these applications require  
44 multi-porphyrinoids arrays, often to promote photoinduced electron<sup>10</sup> or energy<sup>11</sup> transfers or  
45 also mimic the antennas in the photosynthetic reaction center of photosystems.<sup>12</sup> Several  
46 strategies have been developed to prepare multi-porphyrinoids arrays, either by using covalent  
47 (possibly conjugated) bonds,<sup>13,14</sup> self-assemblies<sup>15,16,17</sup> or supramolecular edifices<sup>18</sup>. Peptoid  
48 backbones have been used recently to prepare cofacial arrays of two porphyrins.<sup>19</sup>

49

50 Peptoids (*N*-substituted glycine oligomers)<sup>20</sup> are a special class of peptide mimics  
51 with desirable pharmacokinetic characteristics,<sup>21</sup> including resistance against enzymatic  
52 degradation.<sup>22</sup> They can be synthesized in a sequence-specific manner with hundreds of  
53 different side chains, thanks to the submonomer method<sup>23</sup> which uses primary amines as a  
54 source of side chains diversity.<sup>24</sup> For these reasons, they have been investigated for numerous  
55 biological applications.<sup>25</sup> The supramolecular assembly properties of peptoids also lead  
56 researchers to envisage their application as biomaterials.<sup>26</sup> The side chains of peptoids are  
57 appended to the amide nitrogen atoms rather than to the  $\alpha$ -carbons, resulting in tertiary amide  
58 bonds. Unlike the peptide secondary amide bonds that are *trans*, with the exception of the  
59 prolyl-amide bonds, the peptoid tertiary amide bonds can adopt both the *cis* and *trans*  
60 conformations, one of the major cause of peptoid chain conformational flexibility.<sup>27</sup> Great  
61 efforts have been devoted to controlling the *cis/trans* isomerization of peptoid amides.<sup>28</sup> This  
62 allowed the identification of predictable discrete secondary structures, *i.e.* peptoid  
63 foldamers.<sup>29</sup> Some foldamers are based on a defined combination of backbone *cis* and *trans*  
64 amides,<sup>30</sup> whereas others comprise only *trans*<sup>31</sup> or *cis*-amides.<sup>32</sup> It is now well established that  
65 peptoids with bulky  $\alpha$ -chiral side chains adopt a helical conformation resembling the all-*cis*  
66 polyproline I (PPI) helix, with a 3-fold periodicity and a pitch of approximately 6 Å. The  
67 handedness of a peptoid PPI-type helix is governed by the side chains stereochemistries. (*S*-  
68  $\alpha$ -chiral side chains induce right-handed helices while the *R* configuration induces the left-

69 handed one. The most widely-used  $\alpha$ -chiral side chains are aromatic, especially the  
70 phenylethyl (*pe*)<sup>33</sup> and 1-naphthylethyl (*Inpe*) side chains.<sup>32a</sup> Beside their bulkiness, it has  
71 been reported that local  $n \rightarrow \pi^*$  interactions between backbone carbonyls and the aromatic  
72 groups might be operative for stabilizing the *cis*-amides required for peptoid helix  
73 formation.<sup>28a-c</sup> The presence of one or several aromatic helical faces may also contribute to the  
74 conformational stability of peptoids.<sup>33a</sup> By contrast, very few studies have so far been  
75 undertaken to achieve conformationally stable peptoid PPI-type helices from non-aromatic  
76 monomers.<sup>32b,34</sup> Recently, we demonstrated that the very bulky *tert*-butyl (*tBu*) and  $\alpha$ -*gem*-  
77 dimethyl groups do provide exclusively the *cis*-amide geometry.<sup>35</sup> Another side chain  
78 proposed by us with structure-inducing properties is the  $\alpha$ -chiral 1-*tert*-butylethyl group  
79 (1*tbe*). An octamer sequence consisting of *NtBu* and *Ns1tbe* monomers in equal proportion  
80 provided a very stable PPI-type helix, the longest linear peptoid ever solved by X-ray  
81 crystallography.<sup>32b</sup>

82

83 So far, the insertion of one or several porphyrins on peptoid scaffolds has been  
84 performed to optimize the cellular uptake efficiency of photosensitising porphyrins<sup>36,37</sup> or  
85 mimic the protein–chlorophyll ensemble found in photosynthetic antenna.<sup>38</sup> Jiwon Seo and his  
86 team beautifully used peptoid scaffolds to control porphyrin interactions, in the case of pure  
87 <sup>19,39</sup> or mixed<sup>40,41,42</sup> porphyrinic arrays.

88

89 In this work, the construction of conformationally stable PPI-type helices is sought  
90 after as topological templates to display phthalocyanine arrays. As such architectures are  
91 entirely new, this exploratory work aims at demonstrating the synthetic feasibility of such  
92 hybrids, at investigating the effect of the peptoid on the photoproperties of the phthalocyanine  
93 core, and at assessing the efficiency of the peptoid backbone in controlling the topology of  
94 dimeric phthalocyanine arrays.

95

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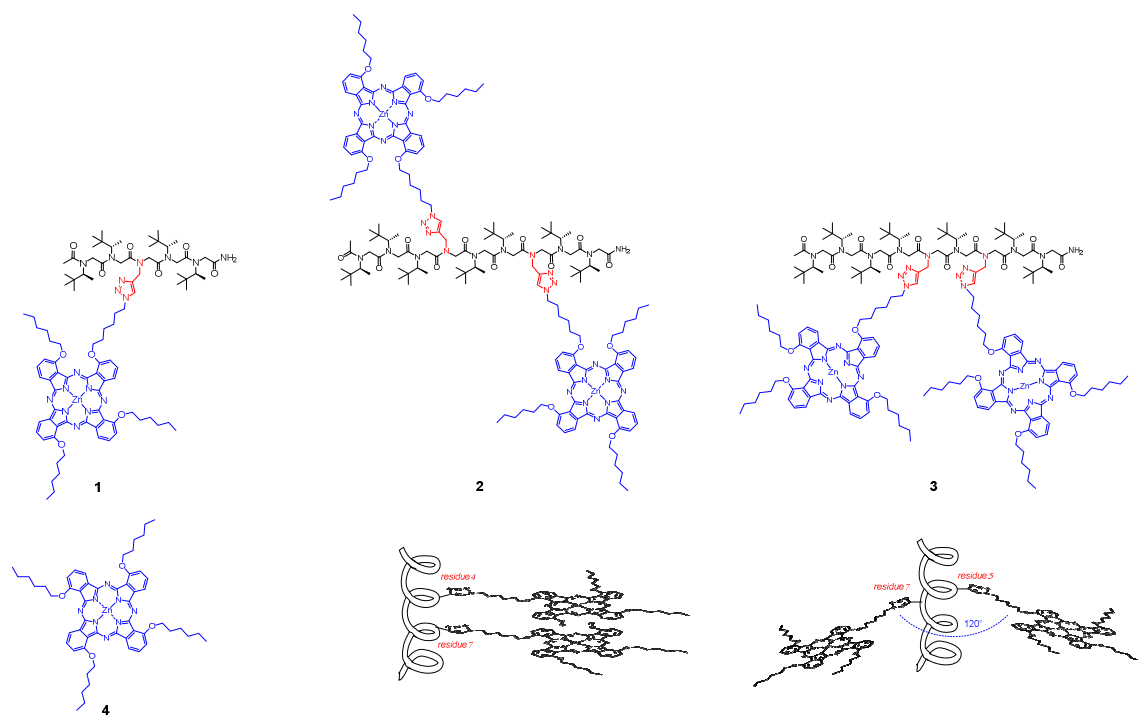
## 97 **2. Results and discussion**

98

### 99 **2.1. Molecular design.**

100 All the hybrid structures designed in the framework of these investigations are  
101 represented in Fig. 1. The first conjugate **1** made of a single phthalocyanine on a pentapeptoid  
102 scaffold was prepared to evaluate the effect of the presence of a peptoid scaffold on the

103 photoproperties of the phthalocyanine core. Hybrids **2** and **3** have each 2 phthalocyanines  
104 grafted on a nonameric peptoid scaffold, with two residues between the two phthalocyanines  
105 in hybrid **2**, and only residue between the two phthalocyanines in hybrid **3**. Phthalocyanine **4**  
106 will be used as the pristine reference phthalocyanine macrocycle.  
107



108  
109 **Fig. 1.** Structures of the peptoid-phthalocyanine architectures **1-3** (with their expected  
110 topology) and of reference phthalocyanine **4**. For each phthalocyanine, only one of the  
111 existing isomers is shown.

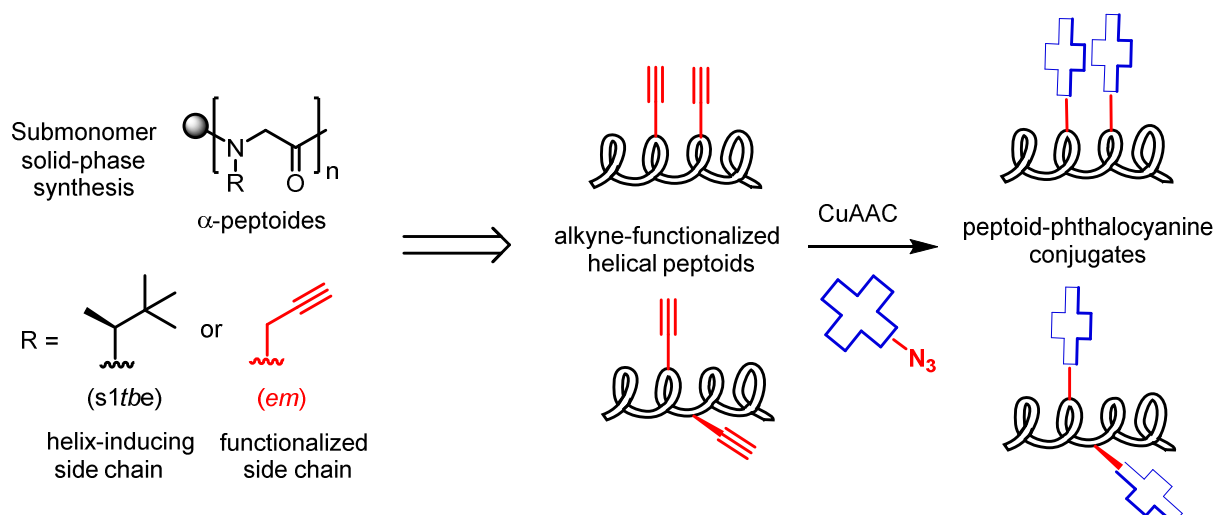
112  
113 The phthalocyanine core was chosen taking into account several considerations: Zn  
114 phthalocyanines have interesting photophysical and photochemical properties which can be  
115 tailored by their substitution pattern,<sup>43</sup> and *O*-hexyl non-peripheral substitution is known to  
116 ensure of a satisfying solubility in common organic solvents.<sup>44</sup>

117  
118 The sequence design of the oligomers lies on the 3-fold periodicity of the PPI-like  
119 helix. The aliphatic *Ns1tbe* monomers have been chosen as helical structure inducers (Fig. 2).  
120 Rather than aromatic side chains likely to interact with the phthalocyanine moieties, aliphatic  
121 side chains have been chosen, also because among all the aliphatic side chains tested so far  
122 the chiral bulky *s1tbe* side chain have achieved the best performance as helical structure  
123 inducer. Three peptoid oligomers have been selected: one pentamer (**5**) and two nonamers (**6**

124 and **7**), each with an acetamide and a carboxamide group at the *N*- and *C*-termini (Table 1).  
 125 The pentamer **5** comprises a (propargyl side chain as the central residue, flanked on each side  
 126 by two *Ns1tbe* residues. Both nonamers are composed of seven *Ns1tbe* and two *Nem* (*N*-  
 127 ethynylmethyl glycine) residues. In the sequence of **6**, the propargyl side chains are appended  
 128 on residues 4 and 7 (*i* and *i*+3), thus the two functional side chains are expected to be on the  
 129 same face of the helix. For the nonamer **7**, the two *Nem* residues correspond to the residues 5  
 130 and 7 of the sequence. Consequently, based on the three-fold periodicity, the propargyl side  
 131 chains are expected to form a 120° degree angle.

132 The synthetic strategy was conceived as detailed on Fig. 2. To anchor the  
 133 phthalocyanines onto the peptoid scaffold, the copper(I)-catalyzed alkyne-azide cycloaddition  
 134 (CuAAC) was chosen for its great versatility<sup>45,46,47</sup> and the easy access to azide-functionalized  
 135 phthalocyanines. This implied the introduction of peptoid side chains containing an alkyne  
 136 function, which we have demonstrated to be achievable.<sup>48</sup> Another expected advantage is that  
 137 the formed 1,2,3-triazole linker is also, at least to some extent, a *cis*-amide inducer,<sup>28c</sup> which  
 138 may help folding the peptoid chain of the conjugate compounds.

139  
 140



141  
 142 **Fig. 2.** Retrosynthetic strategy for the preparation of the targeted peptoid-phthalocyanine  
 143 conjugates

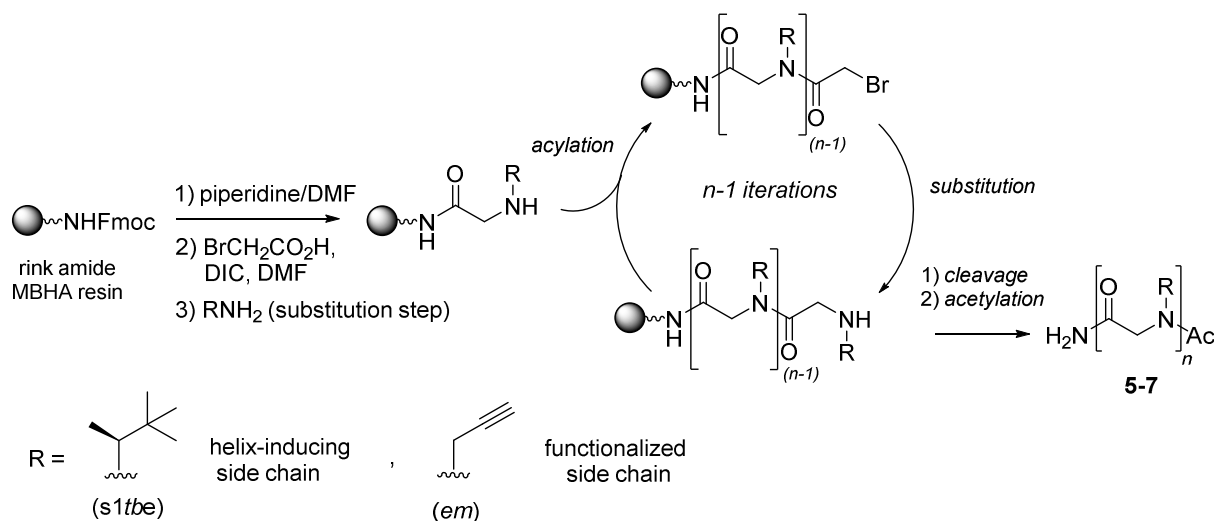
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 145  
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 148

## 2.2. Synthesis and characterization.

149

150 **2.2.1. Propargyl-functionalized peptoid scaffold.** Peptoids are commonly synthesized on  
151 solid support by the submonomer method pioneered by Zuckermann.<sup>23</sup> In this method, each  
152 new monomer is constructed in two iterative steps: (1) bromoacetylation of the *N*-terminus of  
153 the growing chain, (2) introduction of the side chain by bromine atom displacement of the  
154 formed bromoacetamide by a primary amine. This process is most often very efficient,  
155 yielding to pure peptoids in high yields. The major limitation may originate from the steric  
156 hindrance of the formed *N*-terminus secondary amine which may impair chain elongation  
157 efficiency. In this work, the solid-phase synthesis of peptoid oligomers comprising *Ns1tbe*  
158 monomers was evaluated for the first time. The optimized conditions for the synthesis of  
159 peptoids **5-7** are depicted in Scheme 1.

160



169 **Scheme 1.** Solid-phase submonomer synthesis of peptoids **5-7** (optimized conditions).  
170 Acylation step: BrCH<sub>2</sub>CO<sub>2</sub>H (6 equiv., 0.4 M/DMF), diisopropylcarbodiimide (DIC, 8 equiv.,  
171 2 M/DMF), 2 x 5 min at 40 °C. Substitution step: 25 equiv. of HC≡CCH<sub>2</sub>NH<sub>2</sub>, 2M/DMF, 1h  
172 at 40 °C or 15 equiv. of *t*BuCH(CH<sub>3</sub>)NH<sub>2</sub>, 2 M/DMF, 2 x 1h at 40 °C. Cleavage: TFA/DCM  
173 (95/5), RT, 10 min. Acetylation: Ac<sub>2</sub>O (4 equiv.), Et<sub>3</sub>N (2 equiv.), EtOAc (0.2 M), 18h, RT.

170 The syntheses were performed manually using a Fmoc-protected Rink amide MBHA resin as  
171 the support, leading to *C*-terminus carboxamides upon cleavage. After several round of  
172 optimizations, we found that conducting the two steps of the submonomer protocol under  
173 gentle warming (40 °C) had a beneficial effect on the purity of the peptoids. Other important  
improvements concern the acylation step and the substitution reactions with the bulky (*2S*-

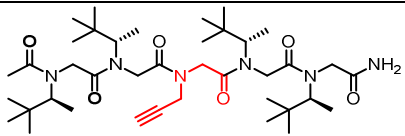
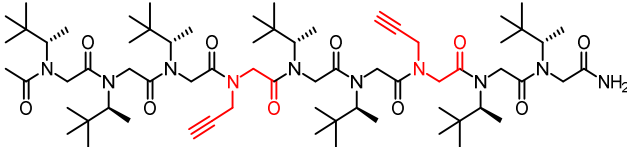
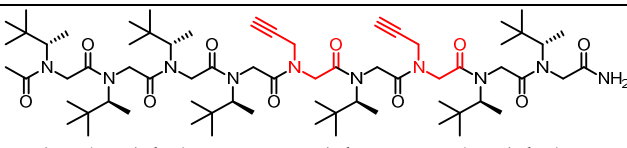


174 3,3-dimethylbutan-2-amine (*sItbe* amine) which were systematically repeated with new  
 175 amount of reagents. The final acetylation of the peptoids was carried out in solution (Ac<sub>2</sub>O,  
 176 Et<sub>3</sub>N), after cleavage from the resin, and the compounds were purified by flash  
 177 chromatography on silica gel. We have indeed observed that the cleavage of the acetylated  
 178 resin-bound peptoids under acidic conditions led to the loss of one residue at the *N*-extremity  
 179 of the oligomers (Table 1).

180

181

182 **Table 1.** Sequence, purity, and mass spectra data of the peptoids **5-7**

peptoid	monomer sequence <sup>a</sup>	overall yield <sup>b</sup>	% purity <sup>c</sup>	expected mass	observed mass
<b>5</b>	 Ac-Ns1tbe-Ns1tbe-Nem-Ns1tbe-Ns1tbe-NH <sub>2</sub>	57	96	718.54	719.54 [M+H] <sup>+</sup>
<b>6</b>	 Ac-(Ns1tbe) <sub>3</sub> -Nem-(Ns1tbe) <sub>2</sub> -Nem-(Ns1tbe) <sub>2</sub> -NH <sub>2</sub>	46	94	1236.92	1237.92 [M+H] <sup>+</sup>
<b>7</b>	 Ac-(Ns1tbe) <sub>4</sub> -Nem-Ns1tbe-Nem-(Ns1tbe) <sub>2</sub> -NH <sub>2</sub>	56	94	1236.92	1237.92 [M+H] <sup>+</sup>

183 <sup>a</sup>The peptoids were drawn with amides in the *trans* conformation by ease of drawing.

184 <sup>b</sup>Based on the resin loading (0.52 mmol.g<sup>-1</sup>), include all the solid-phase steps and the final  
 185 acetylation in solution.

186 <sup>c</sup>Determined by integration of the high-performance liquid chromatography (HPLC) UV trace  
 187 at 214 nm.

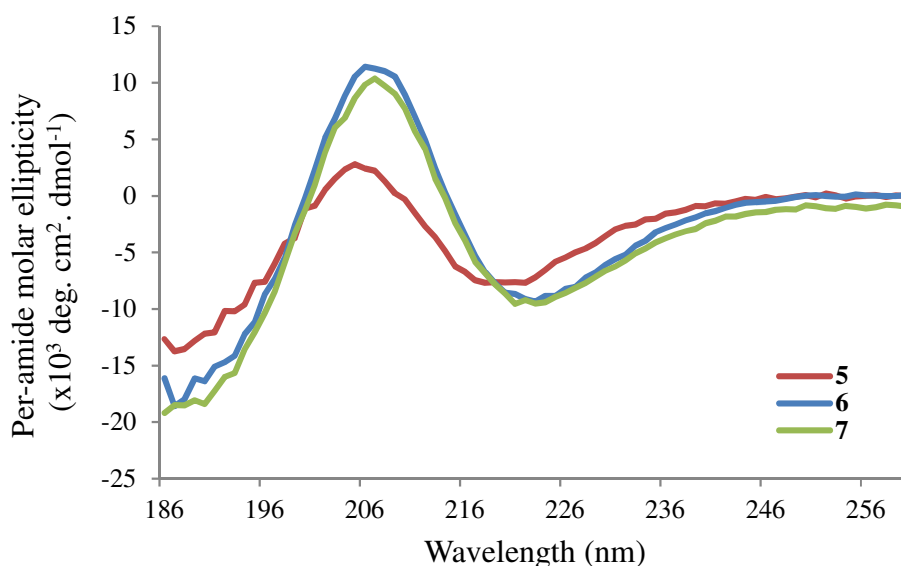
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190 **2.2.2. Conformational studies.** The helical character of the three peptoids **5-7** was assessed by  
 191 Circular Dichroism (CD) in methanol (Fig. 3). The three peptoids display the typical signature  
 192 associated with the PPI-type helical conformation of peptoid oligomers bearing aliphatic side

193 chains. It is worth noting that peptoids with  $\alpha$ -chiral aromatic side chains exhibit a completely  
194 different profile resembling that of the peptide  $\alpha$ -helix. In our case, the *S* configuration of the  
195 *sItbe* side chains produces CD curves with the same right-handed helicity as those of poly-(L-  
196 prolines) in alcohol solvents. The curves are characterized by a positive maximum at around  
197 210 nm and two minima at 190 and 225 nm. The strong intensities of ellipticity on a per-  
198 residue molar basis, at 210 nm for the two nonamers, as compared to literature data, are  
199 indicative of well-structured compounds and likely also reflect a cooperative folding process.  
200 Unsurprisingly, the short length pentamer is less structurally stable; protohelical structures are  
201 generally observed from the tetramer length. Measurements at the Q band wavelength were  
202 not expected to be meaningful due to the high concentrations used to record CD and the  
203 subsequent aggregation affecting their absorption at this wavelength, unlike porphyrins that  
204 are much less prone to aggregation<sup>19</sup>.

205



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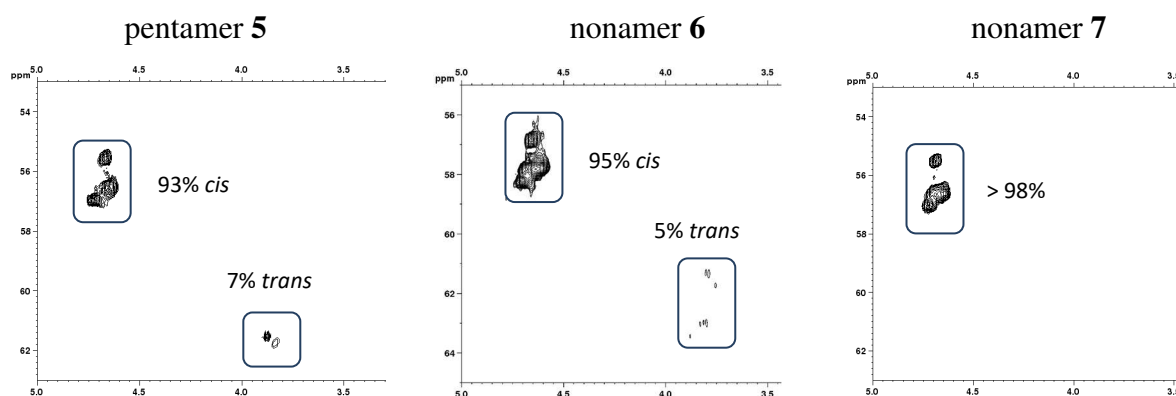
207 **Fig. 3.** CD spectra of peptoid oligomers **5-7** in MeOH at 500  $\mu$ M

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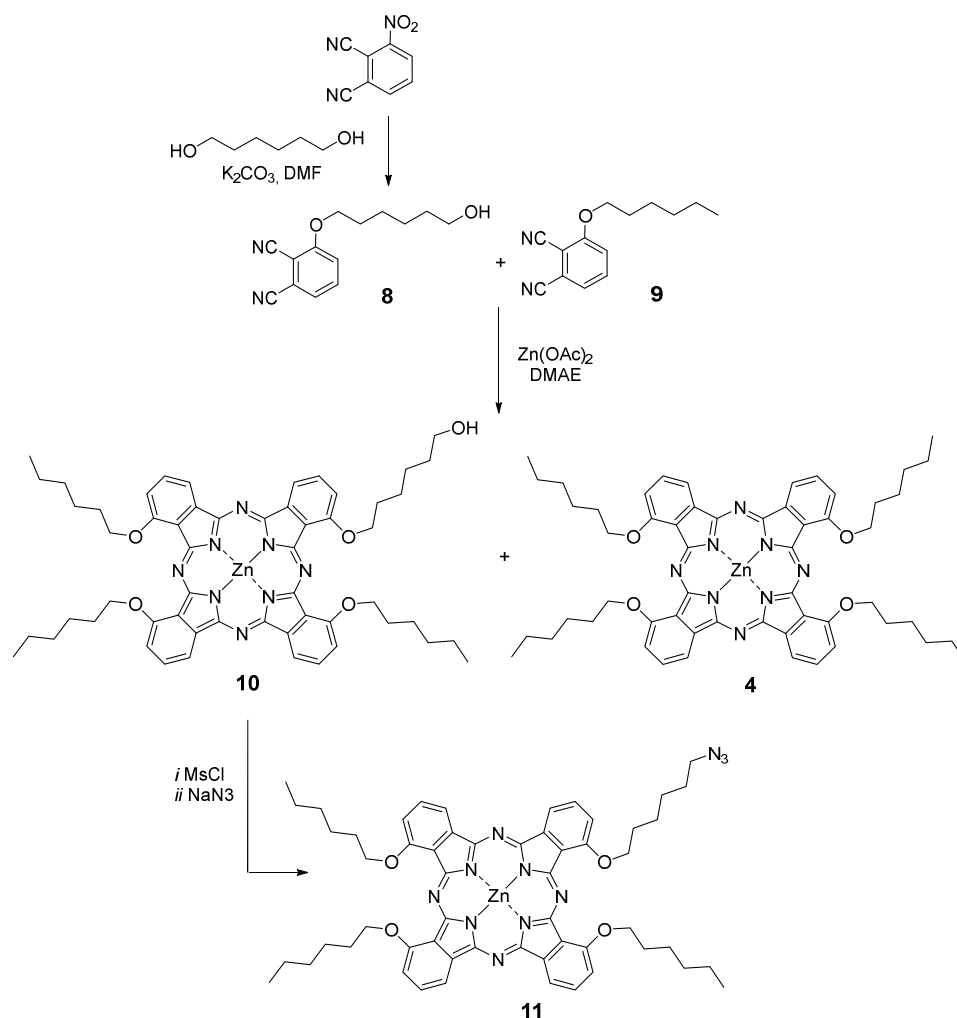
210 As the degree of PPI helical conformational order depends on the population of *cis*-amide  
211 bonds, we also estimated overall backbone amide  $K_{cis/trans}$  values. As recently described by us,  
212 the calculations were made on the basis of the NMR integration (2D-HSQCAD experiments,  
213 Fig. 4) of the methyne protons of the *sItbe* side chains, which show distinct resonances  
214 depending on the *cis* or *trans* conformation of the amides (Supporting information). This  
215 calculation does not take into account the amide bonds connecting the *Nem* residues with their

216 preceding residues, which corresponds to one in five amide bonds in the case of the pentamer  
 217 **5**, and two in nine amides in the case of the nonamers **6** and **7**. The overall proportion of *cis*-  
 218 amide determined for the amide bonds connecting the *Ns1*be residues with their preceding  
 219 residu was found to be 93% for the pentamer **5** ( $K_{cis/trans} = 13.2$ ), and 95% for the nonamer **6**  
 220 ( $K_{cis/trans} = 21.5$ ). The *trans*-amide rotamers were no longer detectable in the case of nonamer  
 221 **7**, it can be confidently considered that the proportion of *cis*-amides >98% ( $K_{cis/trans} > 49.0$ ) in  
 222 this case. These results confirm the high helical character of the nonamer peptoids, the lower  
 223 degree of structuration of the pentamer, as seen from CD, appears to be correlated to a  
 224 substantial conformational heterogeneity arising from *cis/trans* amide bond isomerization.  
 225  
 226



227  
 228 **Fig. 4.**  $^1\text{H}$ - $^{13}\text{C}$  HSQCAD NMR spectra of peptoids **5-7**. Panels display side chains *Ns1*be  
 229 methyne protons. Peptoid concentration was ~ 10 mM in  $\text{CD}_3\text{OD}$ .

230  
 231  
 232  
 233 **2.2.3. Synthesis of azidophthalocyanine.** Azidophthalocyanines, especially when an  
 234 asymmetric substitution pattern, are often prepared from corresponding hydroxylated  
 235 phthalocyanines, via successive mesylation then azidation. Hence monohydroxylated  
 236 phthalocyanine **10** was first prepared from phthalonitriles **8** and **9**, reference phthalocyanine **4**  
 237 being produced concomitantly. The significant difference in polarity of both phthalocyanines  
 238 allowed the quite easy separation by silica gel chromatography. Monohydroxylated  
 239 phthalocyanine **10** was first mesylated, and the crude product was immediately reengaged  
 240 without further purification in the nucleophilic substitution by sodium azide, yielding  
 241 phthalocyanine **11** (Scheme 2).



242

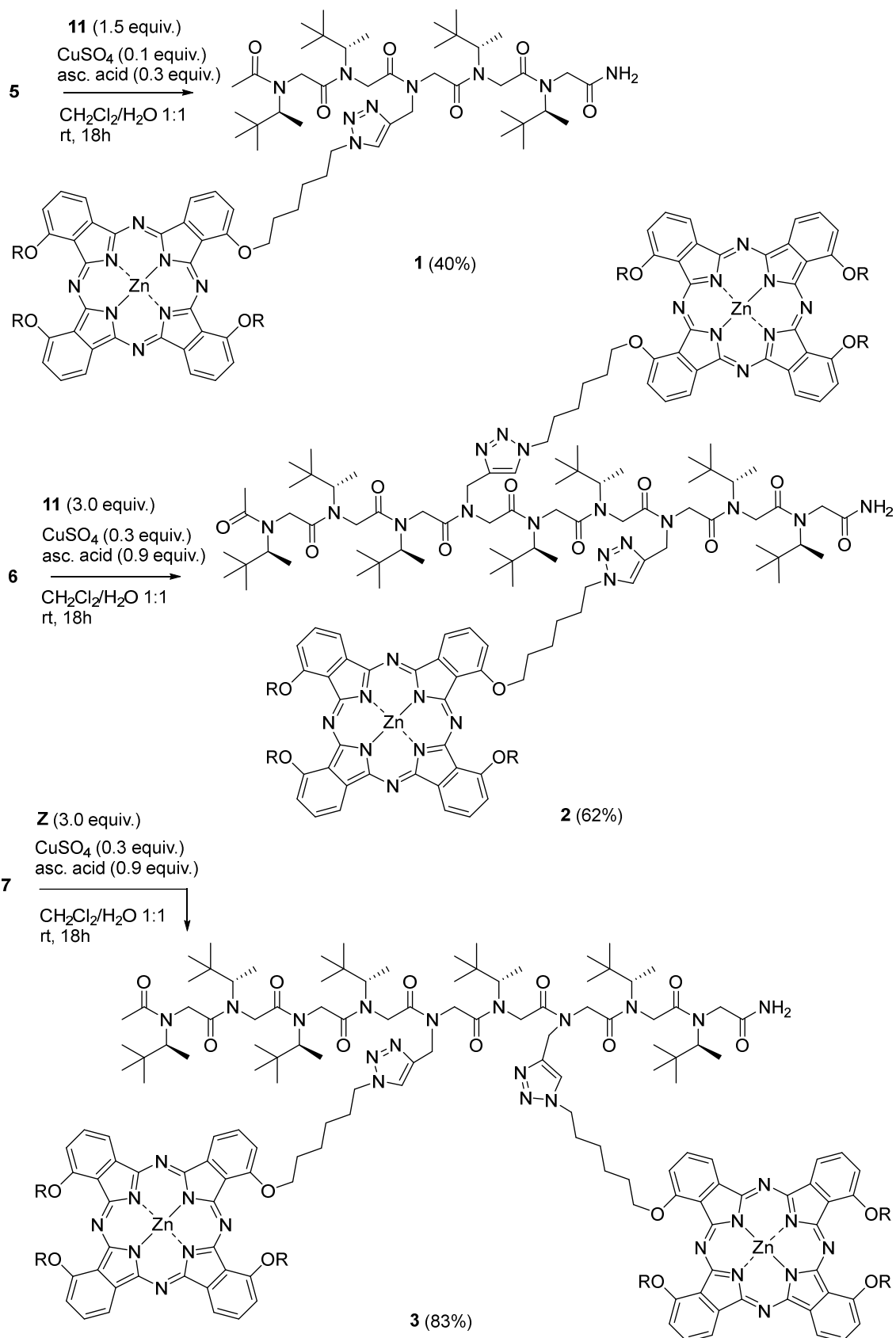
243 **Scheme 2.** Synthesis of azidophthalocyanine **11** and of the reference phthalocyanine **4**. For  
 244 each phthalocyanine, only one of the existing isomers is shown.

245

246

247 **2.2.4. Peptoid-phthalocyanine conjugates 1-3.** Access to the three conjugates **1-3** by copper-  
 248 catalysed azide-alkyne cycloadditions (CuAAC) was successfully achieved using ligand-free  
 249 conditions. The azidophthalocyanine **11** and alkyne-functionalized peptoids were simply  
 250 combined in presence of  $CuSO_4$  and ascorbic acid in a biphasic  $CH_2Cl_2/H_2O$  solvent system  
 251 known to increase CuAAC reaction rates (Scheme 3).<sup>49</sup> The azidophthalocyanine **11** was used  
 252 in a 1.5-fold excess per alkyne functional group to facilitate peptoid conversion. The three  
 253 conjugates were first purified on silica gel to remove excess azidophthalocyanine **11**, and then  
 254 by size-exclusion chromatography, using THF or dichloromethane as eluent, to eliminate  
 255 residual amount of starting peptoids.

256



257

258 **Scheme 3.** Synthesis of the peptoid-phthalocyanine conjugates **1-3**. For each phthalocyanine,

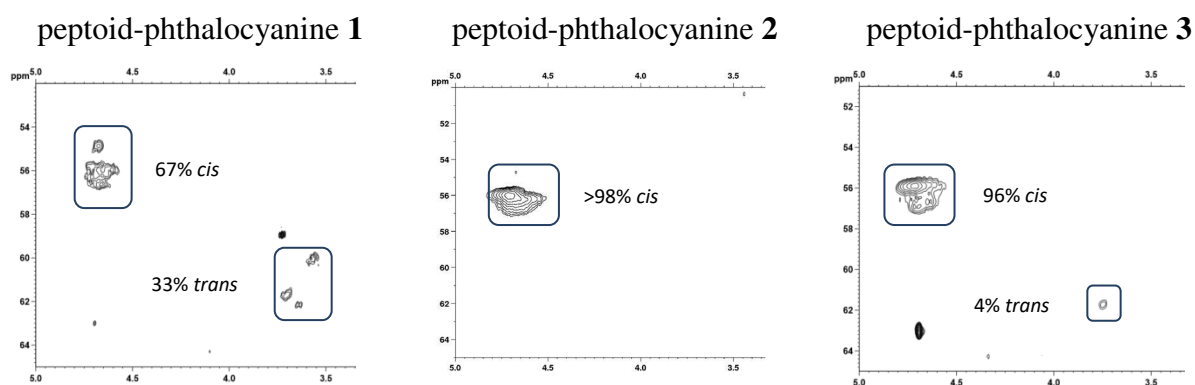
259 only one of the existing isomers is shown.

260

261 CD analysis of peptoid backbone is based on the amide chromophore in the far UV region  
262 (240-180 nm), which necessitates the use of UV-transparent solvents, typically, acetonitrile,  
263 methanol and water. Unfortunately, the poor solubility of peptoid-phthalocyanine conjugates  
264 **1-3** in these solvents prevented their CD analysis.

265 Interestingly, we were able to determine the peptoid amide  $K_{cis/trans}$  for the three conjugates,  
266 from  $^1\text{H}$ - $^{13}\text{C}$ -HSQCAD experiments in deuterated dichloromethane (Fig. 5). As previously,  
267 the methyne *cis* and *trans* peaks of the sltbe side chains were integrated for  $K_{cis/trans}$   
268 determination. We observed a dramatic decrease (84%) in average  $^{N_{sltbe}}K_{cis/trans}$  value for the  
269 shorter peptoid oligomer conjugate **1** (*cis/trans* ratio ~2:1), relative to its parent peptoid **5**  
270 (>13:1). By contrast, peptoid amide isomerism was not significantly affected in the conjugates  
271 **2** and **3** relative to peptoid nonamers **6** and **7**, respectively. Interestingly, the conformational  
272 homogeneity is even increased for the conjugate **2**, for which the *trans* rotamers are no longer  
273 detectable. The opposite is observed for the conjugate **3** relative to **7**. A small proportion of  
274 *trans*-amide rotamers (4%) are observed for **3**, while the parent nonamer **7** was completely  
275 homogeneous in term of amide rotamerism. Overall, the conformational homogeneity of the  
276 two peptoid-phthalocyanine conjugates **2** and **3** is remarkable, especially considering the size  
277 of the phthalocyanine moieties relative to the peptoid templates, and also, the potential intra-  
278 and inter-molecular interactions they can establish. The unique conformation known for  
279 peptoids displaying all-*cis* amides is the PPI helix, the most likely outcome for the two  
280 nonameric conjugates.

281  
282



283 **Fig. 5.**  $^1\text{H}$ - $^{13}\text{C}$  HSQCAD NMR spectra of peptoids **1-3**. Panels display side chains  $N_{sltbe}$   
284 methyne protons. Peptoid concentration was ~ 5 mM in deuterated dichloromethane.

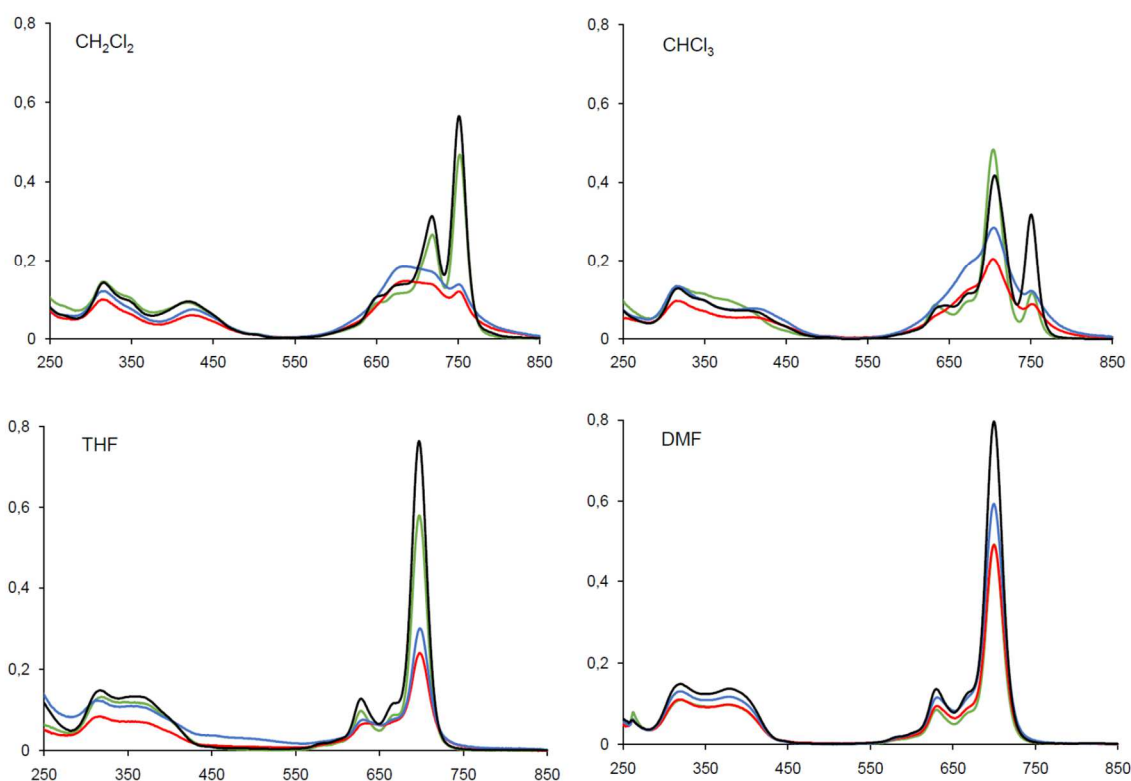
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### 286 **2.3. UV-vis spectroscopy.**

287 The photoproperties of phthalocyanines are depending on structural factors (metal,  
288 nature, number and position of substituents) as well as on external parameters such as the  
289 solvent, concentration and temperature. As all compounds **1-4** have the same phthalocyanine  
290 core, and for the sake of comparisons, UV-vis spectra of **1-4** have been first recorded in  
291 different solvents. As derivatives **2** and **3** have twice the amount of phthalocyanines, 2  $\mu\text{M}$   
292 concentrations were used for **2** and **3** whereas 4  $\mu\text{M}$  concentrations were used for **1** and **4**  
293 (Fig. 6). The maximum absorption for the Q band in each solvent is summarized in Table 2,  
294 together with the extinction coefficient values. Spectra used to determine these values are  
295 displayed in the supplementary material (Figs S1-16).

296

297



298

299 **Figure 6.** Superimposed UV-vis spectra of **1** (green, 4  $\mu\text{M}$ ), **2** (red, 2  $\mu\text{M}$ ), **3** (blue, 2  $\mu\text{M}$ ) and  
300 **4** (black, 4  $\mu\text{M}$ ) in dichloromethane (top left), chloroforme (top right), tetrahydrofurane  
301 (bottom left) and dimethylformamide (bottom right)

302

303 The first observation is that the presence of a peptoid helix and of the triazole ring does not  
304 affect by itself the photoproperties of the phthalocyanine macrocycle, as can be deduced when  
305 comparing spectra of **1** and **4**, whose spectra have roughly the same shape and intensity in all  
306 solvents, with the presence of H-aggregates in chlorinated solvents characterised by the

307 apparition of a red-shifted second Q band in THF and DMF. Conjugates **2** and **3** are very  
 308 aggregated in dichloromethane and in chloroform, as evidenced by their large and potato-  
 309 shaped Q band. In THF and DMF, their Q band is sharper but the absorption is much less  
 310 intense than for **1** and **4**. As the overall concentration of phthalocyanines is the same in all the  
 311 samples, one can infer that it is due to their spatial proximity induced by the grafting of two  
 312 phthalocyanine units on a same peptoid backbone, who therefore interact with each other. The  
 313 position of the phthalocyanine on the peptoid backbone does not affect significantly this  
 314 phenomenon. Even though the helix retains its conformation after the grafting, the flexibility  
 315 of the C6 spacer between the peptoid backbone and the phthalocyanine core provide enough  
 316 freedom of movement to the two macrocycles whose  $\pi$ - $\pi$  stacking occurs similarly for both  
 317 conjugates **2** and **3** in dichloromethane and chloroform. Tetrahydrofuran and *N,N*-  
 318 dimethylformamide both appear to inhibit this intramolecular aggregation, in line with  
 319 previous general observations on phthalocyanines.

320

321 **Table 2**

Compound	Solvent	$\lambda_{\max}$ Q band (nm)	log $\epsilon$
<b>1</b>	CH <sub>2</sub> Cl <sub>2</sub>	752	5.1
	CHCl <sub>3</sub>	704	5.1
	THF	698	5.2
	DMF	700	5.1
<b>2</b>	CH <sub>2</sub> Cl <sub>2</sub>	683	4.9
	CHCl <sub>3</sub>	704	5.0
	THF	698	5.0
	DMF	700	5.4
<b>3</b>	CH <sub>2</sub> Cl <sub>2</sub>	682	5.0
	CHCl <sub>3</sub>	704	5.1
	THF	698	5.2
	DMF	698	5.5
<b>4</b>	CH <sub>2</sub> Cl <sub>2</sub>	751	5.2
	CHCl <sub>3</sub>	705	5.1
	THF	697	5.3
	DMF	700	5.3

322

323 **2.3. Fluorescence**



324 Fluorescence has been recorded in THF to explore more in detail the behaviour of the set of  
325 compounds, as it known to be a more sensitive technique. Spectra are displayed on Fig. 7 and  
326 data are gathered in Table 3.

327

328

329 **Table 3.**

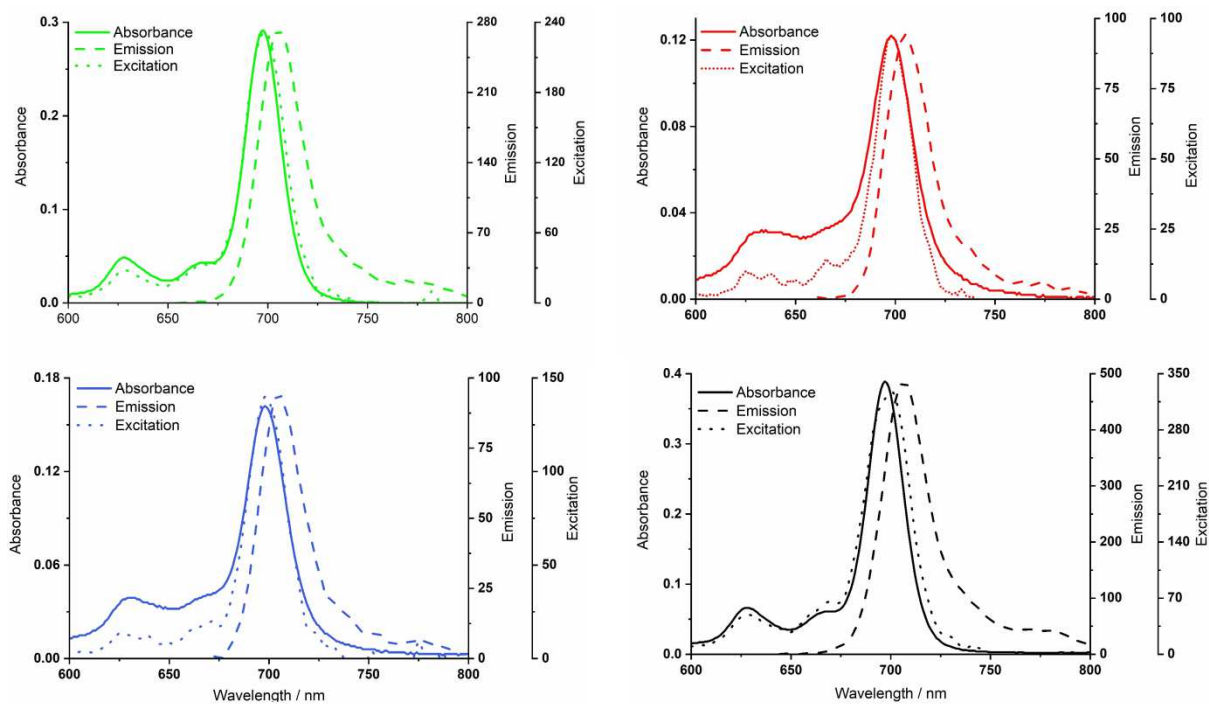
	Excitation $\lambda_{\text{Ex}}$ (nm)	Emission $\lambda_{\text{Em}}$ (nm)	Stokes shift $\Delta_{\text{Stokes}}$ (nm)	$\Phi_{\text{F}}$
<b>1</b>	698	705	7	0.168
<b>2</b>	698	705	7	0.098
<b>3</b>	698	705	7	0.074
<b>4</b>	700	707	10	0.270

330

331

332 At the working concentrations (normalized at absorption 0.2), a significant effect of the sole  
333 presence of the peptoid helix is observed when comparing **1** and **2**. The fluorescence quantum  
334 yield ( $\Phi_{\text{F}}$ ) of **1** is indeed only 2/3 of those of the reference phthalocyanine **4**. When comparing  
335 the data for **3** and **4**,  $\Phi_{\text{F}}$  value is significantly lower than for the monomeric derivatives, and is  
336 interestingly slightly higher for **2** than for **3**, even though the helix of conjugate **2** is more  
337 likely to promote intramolecular interactions as it would promote co-facial geometry. It  
338 confirms the fact the phthalocyanines have enough freedom of movement and hence that their  
339 relative configuration is not much affected by the helix configuration but rather by being  
340 maintained close to each other by the grafting on the same peptoid backbone.

341



**Fig. 7** Normalized fluorescence spectra of compounds **1** (green), **2** (red), **3** (blue) and **4** (black) in tetrahydrofuran

### 3. Experimental

#### 3.1. Materials and methods.

*Synthesis and analysis of compounds 1-3 and 5-7:* THF, CH<sub>2</sub>Cl<sub>2</sub> were dried over aluminum oxide via a solvent purification system. EtOAc, CH<sub>2</sub>Cl<sub>2</sub>, cyclohexane, and MeOH for column chromatography were obtained from commercial sources and were used as received. All other solvents and chemicals obtained from commercial sources were used as received. NMR spectra were recorded on a 400 MHz Bruker Avance III HD spectrometer. Chemical shifts are referenced to the residual solvent peak and *J* values are given in Hertz. The following multiplicity abbreviations are used: (s) singlet, (bs) broad singlet, (d) doublet, (t) triplet, (q) quartet, (m) massif, and (br) broad. TLC was performed on Merck TLC aluminum sheets, silica gel 60, F254. Visualizing of spots was effected with UV-light and vanillin in EtOH/H<sub>2</sub>SO<sub>4</sub>, or ninhidrin or phosphomolybdic acid in EtOH. Flash chromatography was performed with Merck silica gel 60, 40–63 μm. HRMS was recorded on a Micromass Q-ToF Micro (3000 V) apparatus or a Q Exactive Quadrupole-Orbitrap Mass Spectrometer. LC–MS was recorded a Q Exactive Quadrupole-Orbitrap mass spectrometer coupled to a UPLC

363 Ultimate 3000 (Kinetex EVO C18; 1.7  $\mu\text{m}$ ; 100 mm  $\times$  2.1 mm column with a flow rate of  
364 0.45 mL min<sup>-1</sup> with the following gradient: a linear gradient of solvent B from 5% to 95%  
365 over 7.5 min (solvent A = H<sub>2</sub>O + 0.1% formic acid, solvent B = acetonitrile + 0.1% formic  
366 acid) equipped with a DAD UV/vis 3000 RS detector.

367 *Synthesis of compounds 4 and 8-11:* All solvents and reagents were of reagent grade quality  
368 and obtained commercially from Aldrich, Fluka or Merck. Phthalonitrile **9**<sup>44</sup> and 3-  
369 nitrophthalonitrile<sup>50</sup> were prepared as previously reported. Previously reported procedure for  
370 phthalonitrile **8** was slightly modified<sup>51</sup>. IR spectra were recorded between 4000 and 650 cm<sup>-1</sup>  
371 using a PerkinElmer Spectrum 100 FT-IR spectrometer. Mass spectra were recorded on a  
372 MALDI (matrix assisted laser desorption ionization) BRUKER Microflex LT using dithranol  
373 (DIT) or 2,5-dihydroxybenzoic acid (DHB) as the matrix. High resolution mass spectra were  
374 measured on an Agilent 6530 Accurate-Mass Q-TOF LC/MS spectrometer equipped with  
375 electrospray ionization (ESI) source. NMR spectra of **4**, **8**, **10** and **11** were recorded in  
376 deuterated solvents (CDCl<sub>3</sub>) on a Varian 500 MHz spectrometer at 298 K. Electronic  
377 absorption spectra were measured with a Shimadzu UV-2600 UV-vis spectrophotometer, and  
378 a Varian Cary Eclipse spectrofluorometer is used to record steady-state fluorescence emission  
379 spectra were recorded by using at room temperature, using 10 mm path length cuvettes.  
380 Details about the determination of the fluorescence quantum yields are provided in the  
381 supplementary material.

382

### 383 **3.2. Synthesis of peptoid oligomers.**

384 Peptoid oligomers **5-7** were prepared by solid-phase submonomer synthesis. All steps were  
385 carried out manually in 5 mL plastic fritted syringes fitted with a stopcock. The syringes were  
386 placed on a benchtop orbital shaker for reaction incubations. Peptoids were synthesized on  
387 polystyrene resin with an Fmoc-protected Rink amide MBHA linker (Novabiochem 100-200  
388 mesh; 0.52 mmol g<sup>-1</sup>). The Fmoc-protected resin (0.150 g, 0.078 mmol) was swollen in DMF  
389 (2 mL) for 10 min. The swelling was repeated (2x). The Fmoc group was removed by soaking  
390 the resin in a 20% piperidine/DMF solution for 15 min at rt after which time the resin was  
391 washed with DMF (5 $\times$ 2 ml) and drained. The deprotection reaction was repeated (1x). A  
392 solution of bromoacetic acid (65 mg, 0.46 mmol, 6 equiv) in DMF (1.2 mL) was added to the  
393 deprotected resin, followed by a solution of diisopropylcarbodiimide (DIC) (0.1 mL, 0.63  
394 mmol, 8 equiv) in DMF (0.3 mL). The reaction was heated at 40 °C under stirring for 10 min.  
395 This acylation step was repeated (1x) when the growing chain was ended by a bulky *N*-  
396 *tert*butylethyl group. The resin was filtered, washed with DMF (5 $\times$ 2 ml) and drained. A

397 solution of amine (for s1tbe 0.16 mL / 0.6 mL DMF 1.17 mmol, 15 equiv; for em 0.124 mL/  
398 0.6 mL DMF, 1.95 mmol, 25 equiv) added to the resin, and the reaction mixture was agitated  
399 at 40 °C for 1 h.. This step was repeated (1x) in the case of a substitution by the (2S)-3,3-  
400 dimethylbutan-2-amine. The resin was filtered, washed with DMF (5x2 mL) and drained. This  
401 2-steps sequence of acylation and substitution was repeated until the target oligomer was  
402 obtained. After the elongation steps, the resin was washed with CH<sub>2</sub>Cl<sub>2</sub> (5 x 2 mL) and  
403 drained. The peptoid was cleaved from the resin by stirring in 95% trifluoroacetic  
404 acid/CH<sub>2</sub>Cl<sub>2</sub> (2 mL) for 10 min at rt. This cleavage procedure was repeated (1x). The resin  
405 was washed with CH<sub>2</sub>Cl<sub>2</sub> (2 x 2 mL) and the filtrate evaporated under reduced pressure. The  
406 residual TFA was eliminated by adding CH<sub>2</sub>Cl<sub>2</sub> (5 mL) and evaporation. Identity of the  
407 peptoids was controlled by LC-MS (UV detection at 220 nm). The crude oligomers were  
408 acetylated in solution. The peptoid was dissolved in EtOAc (0.2 M) and cooled to 0 °C under  
409 Ar. Et<sub>3</sub>N (2.0 equiv) was added to the stirring solution of amine, followed by addition of  
410 acetic anhydride (4 equiv) under Ar. After 2 h at 0 °C, the mixture was allowed to stand  
411 overnight at room temperature. The mixture was filtered, and the solids washed with EtOAc.  
412 The filtrate was then concentrated in vacuo, yielding the crude *N*-acetylated compound which  
413 was purified by flash column chromatography on silica gel. Purity was determined by HPLC  
414 (UV detection at 220 nm), and the product identity was confirmed by HRMS before CD and  
415 NMR analyses.

416 **Peptoid 5** (*Ac-NsItbe-NsItbe-Nem-NsItbe-NsItbe-NH<sub>2</sub>*). Pentamer **5** was isolated as a white  
417 solid after purification by flash column chromatography on silica gel. 32.8 mg (0.045 mmol)  
418 were obtained. *R<sub>f</sub>* = 0.63 (9:1 CH<sub>2</sub>Cl<sub>2</sub>/MeOH). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ (ppm): 0.76-  
419 1.41 (m, 48 H, CH(CH<sub>3</sub>)C(CH<sub>3</sub>)<sub>3</sub>), 1.92-2.26 (m, 4 H, COCH<sub>3</sub> and C≡CH), 3.33-5.10 (m, 16  
420 H, 5xNCH<sub>2</sub>CO, NCH<sub>2</sub>C≡C and 4xCH(CH<sub>3</sub>)C(CH<sub>3</sub>)<sub>3</sub>), 5.52-7.95 (m, 2 H, CONH<sub>2</sub>). HRMS  
421 (TOF MS ES+) *m/z* calcd for C<sub>39</sub>H<sub>71</sub>N<sub>6</sub>O<sub>6</sub><sup>+</sup> [M+H]<sup>+</sup> 719.5429; found 719.5417.

422 **Peptoid 6** (*Ac-NsItbe-NsItbe-NsItbe-Nem-NsItbe-NsItbe-Nem-NsItbe-NsItbe-NH<sub>2</sub>*).  
423 Peptoid **6** was isolated as a white solid after purification by flash column chromatography on  
424 silica gel. 25.17 mg (0.020 mmol) were obtained. *R<sub>f</sub>* = 0.65 (9:1 CH<sub>2</sub>Cl<sub>2</sub>/MeOH). <sup>1</sup>H NMR  
425 (400 MHz, CDCl<sub>3</sub>) δ (ppm): <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ (ppm): 0.71-1.36 (m, 84 H,  
426 CH(CH<sub>3</sub>)C(CH<sub>3</sub>)<sub>3</sub>), 1.92-2.17 (m, 3 H, COCH<sub>3</sub>), 2.26-2.49 (m, 2 H, C≡CH), 3.47-4.88 (m, 29  
427 H, 9xNCH<sub>2</sub>CO, 2xNCH<sub>2</sub>C≡C and 7xCH(CH<sub>3</sub>)C(CH<sub>3</sub>)<sub>3</sub>), 5.97-7.92 (m, 2 H, CONH<sub>2</sub>). HRMS  
428 (TOF MS ES+) *m/z* calcd for C<sub>68</sub>H<sub>121</sub>N<sub>10</sub>O<sub>10</sub><sup>+</sup> [M+H]<sup>+</sup> 1237.9262; found 1237.9267.

429 **Peptoid 7** (*Ac-NsItbe-NsItbe-NsItbe-NsItbe-Nem-NsItbe-Nem-NsItbe-NsItbe-NH<sub>2</sub>*).  
430 Peptoid **7** was isolated as a white solid after purification by flash column chromatography on

431 silica gel. 54.2 mg (0.043 mmol) were obtained.  $R_f = 0.65$  (9:1  $\text{CH}_2\text{Cl}_2/\text{MeOH}$ ).  $^1\text{H NMR}$   
432 (400 MHz,  $\text{CDCl}_3$ )  $\delta$  (ppm): 0.62-1.37 (m, 84 H,  $\text{CH}(\text{CH}_3)\text{C}(\text{CH}_3)_3$ ), 2.00-2.07 (m, 3 H,  
433  $\text{COCH}_3$ ), 2.26-2.39 (m, 2 H,  $\text{C}\equiv\text{CH}$ ), 3.39-4.88 (m, 29 H,  $9\times\text{NCH}_2\text{CO}$ ,  $2\times\text{NCH}_2\text{C}\equiv\text{C}$  and  
434  $7\times\text{CH}(\text{CH}_3)\text{C}(\text{CH}_3)_3$ ), 5.97-7.92 (m, 2 H,  $\text{CONH}_2$ ). HRMS (TOF MS ES+)  $m/z$  calcd for  
435  $\text{C}_{68}\text{H}_{121}\text{N}_{10}\text{O}_{10}^+$   $[\text{M}+\text{H}]^+$  1237.9262; found 1237.9247.

436

### 437 **3.3. Synthesis of azidophthalocyanine 11**

438 **Phthalonitrile 8.** 3-nitrophthalonitrile (3.46 g, 20 mmol), 1,6 hexanediol (100 mmol, 12.0 g)  
439 and  $\text{K}_2\text{CO}_3$  (500 mmol, 69 g) were stirred overnight at 60 °C in DMF (150 mL), then the  
440 reaction mixture was poured into water. The resulting white precipitate was filtrated and  
441 purified by column chromatography (silica gel) using  $\text{CH}_2\text{Cl}_2/\text{EtOH}$  (50/1) as the eluent.  
442 Yield: 53% (2.6 g).  $\text{C}_{14}\text{H}_{16}\text{N}_2\text{O}_2$ , MW: 244.29 g/mol. FT-IR ( $\nu$ ,  $\text{cm}^{-1}$ ): 3315, 3088, 2935,  
443 2861, 2238, 2226, 1978, 1580, 1473, 1453, 1399, 1291, 1177, 1033, 923, 844, 795, 729, 684.  
444  $^1\text{H NMR}$  (500 MHz,  $\text{CDCl}_3$ )  $\delta$  (ppm): 1.47 (2 H), 1.56 (m, 3 H), 1.90 (2 H), 3.67 (t, 2 H),  
445 4.14 (t, 2 H), 7.21 (d, 1 H), 7.33 (d, 1 H), 7.62 (t, 1 H).  $^{13}\text{C NMR}$  (125 MHz,  $\text{CDCl}_3$ ),  $\delta$   
446 (ppm): 25.58, 25.84, 28.81, 32.68, 62.91, 70.03, 110.01, 116.69, 117.37, 125.02, 134.57,  
447 161.63.

448 **Hydroxyphthalocyanine 10 and phthalocyanine 4.** Phthalonitrile **8** (300 mg, 1.23 mmol),  
449 phthalonitrile **9** (1.4 g, 6.15 mmol), and zinc acetate (0.13 g, 0.61 mmol) were stirred at 140  
450 °C for 18 h under argon atmosphere in dry dimethylaminoethanol (6 mL). After evaporation  
451 of the solvent, the residue was dissolved in dichloromethane and purified by column  
452 chromatography (silica gel) using  $\text{CH}_2\text{Cl}_2/\text{EtOH}$  (20/1) as the eluent. Symmetrically  
453 substituted phthalocyanine **4** was first eluted, followed by the monohydroxylated  $\text{AB}_3$   
454 derivative **10**.

455 **Symmetric phthalocyanine 4.** Dark-blue waxy solid. 50 mg.  $\text{C}_{56}\text{H}_{64}\text{N}_8\text{O}_4\text{Zn}$ , MW 978.56.  
456 ATR-IR:  $\nu_{\text{max}}$  ( $\text{cm}^{-1}$ ) 2924, 2853 (aliph. C-H), 1594 (C=N).  $^1\text{H NMR}$  (500 MHz,  $\text{CDCl}_3$ )  $\delta$   
457 ppm: 0.89-1.06 (m, 12 H,  $\text{CH}_3$ ), 1.25-2.58 (m, 32 H,  $\text{CH}_2$ ), 3.95-4.89 (m, 8 H,  $\text{CH}_2$ ), 6.71-  
458 9.04 (m, 12 H, ArCH).  $^{13}\text{C NMR}$  (125 MHz,  $\text{CDCl}_3$ )  $\delta$  ppm: 14.36, 22.98-32.44, 68.73-69.56,  
459 68.55, 70.59, 110.31-157.47. Anal. calc. for  $\text{C}_{56}\text{H}_{64}\text{N}_8\text{O}_4\text{Zn}$ : 68.74 C, 6.59; H, 11.45; N %,  
460 Found: C, 68.50; H, 6.40; N, 11.20. MALDI-TOF:  $m/z$  979.12  $[\text{MH}]^+$ . HR-ESI-MS  $m/z$ :  
461 Calcd for  $\text{C}_{56}\text{H}_{65}\text{N}_8\text{O}_5\text{Zn}$ : 977.4420, Found 977.4331.

462 **Monohydroxyphthalocyanine 10.** Dark-blue waxy solid. 20 mg.  $\text{C}_{56}\text{H}_{64}\text{N}_8\text{O}_5\text{Zn}$ , MW 994.56.  
463 ATR-IR:  $\nu_{\text{max}}$  ( $\text{cm}^{-1}$ ) 3410 (OH), 2924, 2853 (aliph. C-H), 1594 (C=N).  $^1\text{H NMR}$  (500 MHz,  
464  $\text{DMSO}-d_6$ )  $\delta$  ppm: 0.81-1.01 (m, 9 H,  $\text{CH}_3$ ), 1.26-1.78 (m, 24 H,  $\text{CH}_2$ ), 1.85-2.31 (m, 10 H,

465 CH<sub>2</sub>), 3.50-3.58 (m, 1 H, OH), 4.07-4.94 (m, 8H, CH<sub>2</sub>O), 7.27-8.25 (m, 12 H, ArCH). <sup>13</sup>C  
466 NMR (125 MHz, DMSO-*d*<sub>6</sub>) δ ppm: 14.65, 22.73-33.40, 61.12, 61.44, 68.55, 70.59, 113.16-  
467 156.50. Anal. calc. for C<sub>56</sub>H<sub>64</sub>N<sub>8</sub>O<sub>5</sub>Zn: 67.63 C, 6.49; H, 11.27; N %, Found: C, 67.50; H,  
468 6.55; N, 11.35. MALDI-TOF: *m/z* 992.52 [M]<sup>+</sup>. HR-ESI-MS *m/z*: Calcd for C<sub>56</sub>H<sub>65</sub>N<sub>8</sub>O<sub>5</sub>Zn:  
469 993.4369, Found 993.4282.

470 **Azido-phthalocyanine (II)**. Monohydroxlated **10** (50 mg, 0.05 mmol) and triethylamine (0.50  
471 mL) were stirred in dichloromethane (2 mL) in an ice bath, then mesyl chloride (0.20 mL)  
472 was added drop by drop over 10 min. The stirring continues overnight at room temperature,  
473 then the reaction mixture was washed by a dilute solution of potassium carbonate then by  
474 water. The organic phase was dried on sodium sulfate. The resulting crude mesylated  
475 derivative and NaN<sub>3</sub> (0.5 mmol) are stirred in DMF at 80 °C overnight. CAUTION: NaN<sub>3</sub>  
476 must be kept away from acid medium and be handled with care. Water is then added to the  
477 cooled reaction mixture and the resulting precipitate is filtrated, recovered in dichloromethane  
478 and purified by column chromatography (silica gel) using CH<sub>2</sub>Cl<sub>2</sub>:EtOH (20:1) as the mobile  
479 phase. Yield 90% (46 mg). C<sub>56</sub>H<sub>63</sub>N<sub>11</sub>O<sub>4</sub>Zn, MW 1019.57. ATR-IR: ν<sub>max</sub> (cm<sup>-1</sup>) 2924, 2854  
480 (aliph. C-H), 2092 (N<sub>3</sub>), 1590 (C=N). <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>) δ ppm: 0.77-1.05 (m,  
481 9H, CH<sub>3</sub>), 1.20-2.37 (m, 34H, CH<sub>2</sub>), 4.65, 4.90 (b, 8H, CH<sub>2</sub>-O), 7.62-9.08 (m, 12H, ArCH).  
482 <sup>13</sup>C NMR (125 MHz, DMSO-*d*<sub>6</sub>) δ ppm: 14.21, 22.52-32.21, 51.29, 55.44, 68.83, 70.76,  
483 113.30-156.38. Anal. calc. for C<sub>56</sub>H<sub>63</sub>N<sub>11</sub>O<sub>4</sub>Zn: 65.97 C, 6.23; H, 15.11; N %, Found: C,  
484 65.40; H, 6.35; N, 15.45. MALDI-TOF: *m/z* 1020.50 [MH]<sup>+</sup>. HR-ESI-MS *m/z*: Calcd for  
485 C<sub>56</sub>H<sub>64</sub>N<sub>11</sub>O<sub>4</sub>Zn: 1018.4434, Found 1018.4331.

486

### 487 **3.4. Synthesis of peptoid-phthalocyanine conjugate**

488 **Synthesis of peptoid-phthalocyanine conjugate 1**. To a solution of peptoid **5** (13 mg, 0.0180  
489 mmol) and phthalocyanine **11** (20 mg, 0.0216 mmol, 1.2 equiv) dissolved in a biphasic  
490 CH<sub>2</sub>Cl<sub>2</sub>/H<sub>2</sub>O (1:1) solvent system (0.5 mL) were added freshly prepared 0.1 M aq. ascorbic  
491 acid (0.3 equiv.), and 0.1 M aq. CuSO<sub>4</sub> (0.1 equiv.). After vigorous stirring for 24 h, the  
492 mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub> (6 mL), washed with water (2 x1 mL), dried over Na<sub>2</sub>SO<sub>4</sub>,  
493 and concentrated in vacuo. The crude residue was purified first by flash column  
494 chromatography (SiO<sub>2</sub>, 0.1 % EtOH in CH<sub>2</sub>Cl<sub>2</sub>) and then on Bio-Beads S3 stationary phase  
495 (CH<sub>2</sub>Cl<sub>2</sub>), to yield the peptoid-phthalocyanine conjugates **1** (12.4 mg, 40%) as a green  
496 powder. TLC : R<sub>f</sub> = 0.63 (9:1 CH<sub>2</sub>Cl<sub>2</sub>/MeOH). <sup>1</sup>H NMR (400 MHz, DMF-*d*<sub>7</sub>) δ (ppm): 0.56-  
497 2.32 (m, 91 H, 4×CH(CH<sub>3</sub>)C(CH<sub>3</sub>)<sub>3</sub>, 3×OCH<sub>2</sub>(CH<sub>2</sub>)<sub>4</sub>CH<sub>3</sub>, and OCH<sub>2</sub>(CH<sub>2</sub>)<sub>5</sub>), 2.40 (bs, 3 H,  
498 COCH<sub>3</sub>), 3.63-4.86 (m, 20 H, 4×OCH<sub>2</sub>, 5×NCH<sub>2</sub>CO, and NCH<sub>2</sub>-triazole), 4.88-5.19 (m, 4 H,

499  $4\times\text{CH}(\text{CH}_3)\text{C}(\text{CH}_3)_3$ , 7.09-9.24 (m, 13 H,  $4\times\text{Ar}(\text{CH})_3$ , and  $\text{C}=\text{CHN}$ -triazole). HRMS (TOF  
500 MS ES+)  $m/z$  calcd for  $\text{C}_{95}\text{H}_{134}\text{N}_{17}\text{O}_{10}^{64}\text{Zn}$   $[\text{M}+\text{H}]^+$  1736.9785; found 1736.9771. MALDI-  
501 TOF:  $m/z$  1735.9712 (calcd. for  $\text{C}_{95}\text{H}_{133}\text{N}_{17}\text{O}_{10}^{64}\text{Zn}$ ;  $[\text{M}]^+$ ; found 1735.171).

502 **Synthesis of peptoid-phthalocyanine conjugate 2.** To a solution of peptoid **6** (10.5 mg,  
503 0.0084 mmol) and phthalocyanine **11** (25.1 mg, 0.025 mmol, 3 equiv) dissolved in a biphasic  
504  $\text{CH}_2\text{Cl}_2/\text{H}_2\text{O}$  (1:1) solvent system (0.66 mL) were added freshly prepared 0.1 M aq. ascorbic  
505 acid (0.9 equiv.), and 0.1 M aq.  $\text{CuSO}_4$  (0.3 equiv.). After vigorous stirring for 24 h, the  
506 mixture was diluted with  $\text{CH}_2\text{Cl}_2$  (10 mL), washed with water (2 x2 mL), dried over  $\text{Na}_2\text{SO}_4$ ,  
507 and concentrated in vacuo. The crude residue was purified first by column chromatography  
508 ( $\text{SiO}_2$ , 0.1 % EtOH in  $\text{CH}_2\text{Cl}_2$ ) and then by Bio-Beads S3 (THF), to yield the peptoid-  
509 phthalocyanine conjugates **2** (17.2 mg, 62 %) as a green powder. TLC:  $R_f$  = 0.65 (9:1  
510  $\text{CH}_2\text{Cl}_2/\text{MeOH}$ ).  $^1\text{H}$  NMR (400 MHz,  $\text{DMF-}d_7$ )  $\delta$  (ppm): 0.64-2.62 (m, 173 H,  
511  $7\times\text{CH}(\text{CH}_3)\text{C}(\text{CH}_3)_3$ ,  $6\times\text{OCH}_2(\text{CH}_2)_4\text{CH}_3$ ,  $2\times\text{OCH}_2(\text{CH}_2)_5$  and  $\text{COCH}_3$ ), 3.53-5.33 (m, 45 H,  
512  $8\times\text{OCH}_2$ ,  $9\times\text{NCH}_2\text{CO}$ ,  $2\times\text{NCH}_2$ -triazole and  $7\times\text{CH}(\text{CH}_3)\text{C}(\text{CH}_3)_3$ ), 6.34-9.30 (m, 26 H,  
513  $8\times\text{Ar}(\text{CH})_3$ , and  $2\times\text{C}=\text{CHN}$ -triazole). HRMS (TOF MS ES+)  $m/z$  calcd for  
514  $\text{C}_{180}\text{H}_{248}\text{N}_{32}\text{O}_{18}^{64}\text{Zn}_2$   $[\text{M}+2\text{H}]^{2+}$  1636.9023; found 1636.9044. MALDI-TOF:  $m/z$  3271.790  
515 (calcd. for  $\text{C}_{180}\text{H}_{246}\text{N}_{32}\text{O}_{18}^{64}\text{Zn}_2$ ;  $[\text{M}]^+$  3271.7901).

516 **Synthesis of peptoid-phthalocyanine conjugate 3.** The same protocol as for the synthesis of  
517 compound **2** was applied to the synthesis of conjugate **3**, which was isolated in 83% yield (22  
518 mg), starting from 10 mg of peptoid **7**. TLC:  $R_f$  = 0.65 (9:1  $\text{CH}_2\text{Cl}_2/\text{MeOH}$ ).  $^1\text{H}$  NMR (400  
519 MHz,  $\text{DMF-}d_7$ )  $\delta$  (ppm): 0.56-2.65 (m, 173 H,  $7\times\text{CH}(\text{CH}_3)\text{C}(\text{CH}_3)_3$ ,  $6\times\text{OCH}_2(\text{CH}_2)_4\text{CH}_3$ ,  
520  $2\times\text{OCH}_2(\text{CH}_2)_5$  and  $\text{COCH}_3$ ), 3.54-5.39 (m, 45 H,  $8\times\text{OCH}_2$ ,  $9\times\text{NCH}_2\text{CO}$ ,  $2\times\text{NCH}_2$ -triazole  
521 and  $7\times\text{CH}(\text{CH}_3)\text{C}(\text{CH}_3)_3$ ), 6.82-9.30 (m, 26 H,  $8\times\text{Ar}(\text{CH})_3$ , and  $2\times\text{C}=\text{CHN}$ -triazole). HRMS  
522 (TOF MS ES+)  $m/z$  calcd. for  $\text{C}_{180}\text{H}_{249}\text{N}_{32}\text{O}_{18}^{64}\text{Zn}_2$   $[\text{M}+3\text{H}]^{3+}$  1091.6039; found 1091.6047.  
523 MALDI-TOF:  $m/z$  3271.790 (calcd. for  $\text{C}_{180}\text{H}_{246}\text{N}_{32}\text{O}_{18}^{64}\text{Zn}_2$ ;  $[\text{M}]^+$  3271.7901).

524

#### 525 4. Conclusions

526 In this work, peptoid oligomers have been used as scaffolds to display phthalocyanines dyes.  
527 In this way, the first synthesis and analysis of the photoproperties of peptoid-phthalocyanine  
528 conjugates has been achieved. The peptoid oligomers with aliphatic side chains to prevent  
529 aromatic interactions were designed to adopt the PolyProline type I conformation  
530 characterized by *cis*-amide bonds and the phthalocyanines were grafted by CuAAC reactions.  
531 Very importantly, the helix content of the peptoids is conserved upon phthalocyanine grafting,  
532 allowing for an accurate comparison of the photoproperties of the conjugates. Measurements

533 of the electronic absorption and fluorescence spectra of the conjugates and of a monomeric  
534 reference phthalocyanine showed that the presence of the peptoid backbone has no effect on  
535 the photoproperties of the phthalocyanines. Conjugates in which two phthalocyanine units are  
536 grafted on a same peptoid backbone exhibit intramolecular aggregation behaviour, due to the  
537 fact that the two phthalocyanines are kept close to each other, quite independently of their  
538 relative position on the peptoid backbone, probably due to the important flexibility of the  
539 spacer used. Further works with more rigid spacers between the phthalocyanine and the helix  
540 backbone should allow to better control the interactions between the macrocycles.

541

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