

Late Smithian microbial deposits and their lateral marine fossiliferous limestones (Early Triassic, Hurricane Cliffs, Utah, USA)

Nicolas Olivier, Emmanuel Fara, Emmanuelle Vennin, Kevin Bylund, James Jenks, Gilles Escarguel, Daniel A. Stephen, Nicolas Goudemand, Dawn Snyder, Christophe Thomazo, et al.

▶ To cite this version:

Nicolas Olivier, Emmanuel Fara, Emmanuelle Vennin, Kevin Bylund, James Jenks, et al.. Late Smithian microbial deposits and their lateral marine fossiliferous limestones (Early Triassic, Hurricane Cliffs, Utah, USA). Facies, 2018, 64 (2), pp.13. 10.1007/s10347-018-0526-3. hal-01748441

HAL Id: hal-01748441 https://uca.hal.science/hal-01748441

Submitted on 10 Dec 2019

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1	Late Smithian microbial deposits and their lateral marine fossiliferous limestones (Early
2	Triassic, Hurricane Cliffs, Utah, USA)
3	
4	Nicolas Olivier ¹ \cdot Emmanuel Fara ² \cdot Emmanuelle Vennin ² \cdot Kevin G. Bylund ³ \cdot James F.
5	Jenks ⁴ · Gilles Escarguel ⁵ · Daniel A. Stephen ⁶ · Nicolas Goudemand ⁷ · Dawn Snyder ⁸ · C.
6	Thomazo ² · Arnaud Brayard ²
7	
8	
9 10 11 12 13 14 15 16 17 18 19 20 21 22	 ¹Université Clermont Auvergne, CNRS, Laboratoire Magmas et Volcans, F-63000 Clermont-Ferrand, France ²Biogéosciences, UMR 6282, CNRS, Université Bourgogne Franche-Comté, 6 Boulevard Gabriel, 21000 Dijon, France ³140 South 700 East, Spanish Fork, Utah 84660, USA ⁴1134 Johnson Ridge Lane, West Jordan, Utah 84084, USA ⁵Université de Lyon, Laboratoire d'Ecologie des Hydrosystèmes Naturels et Anthropisés, UMR CNRS 5023, Université Claude Bernard Lyon 1, ENTPE, 6 rue Raphaël Dubois, F-69622 Villeurbanne, France ⁶Utah Valley University, Department of Earth Science, Orem, Utah 84058, USA ⁷Université de Lyon, ENS de Lyon, CNRS, Université Claude Bernard Lyon 1, Institut de Génomique Fonctionnelle de Lyon, UMR 5242, 46 Allée d'Italie, F-69364 Lyon cedex 07, France ⁸7202 Yardley Drive, Katy Texas 77494, USA Corresponding author: nicolas.olivier@uca.fr
23	
24	
25	Abstract
26	Recurrent microbialite proliferations during the Early Triassic are usually explained
27	by ecological relaxation and abnormal oceanic conditions. Most Early Triassic microbialites
28	are described as single or multiple lithological units without detailed ecological information

about lateral and coeval fossiliferous deposits. Exposed rocks along Workman Wash in the
Hurricane Cliffs (south-western Utah, USA) provide an opportunity to reconstruct the spatial

31 relationships of late Smithian microbialites with adjacent and contemporaneous fossiliferous

sediments. Microbialites deposited in an intertidal to subtidal interior platform, are
 intercalated between inner tidal flat dolosiltstones and subtidal bioturbated fossiliferous

limestones. Facies variations along these fossiliferous deposits and microbialites can be traced
laterally over a few hundred of meters. Preserved organisms reflect a moderately diversified

assemblage, contemporaneous to the microbialite formation. The presence of such a fauna,
including some stenohaline organisms (echinoderms) indicates that the development of these
late Smithian microbial deposits occurred in normal marine waters as a simple facies belt

39 subject to relative sea-level changes. Based on this case study, the proliferation of

40 microbialites cannot be considered as direct evidence for presumed harsh environmental41 conditions.

42

43 Keywords Early Triassic · Late Smithian · Microbial deposits · Metazoan fauna · Lingulids ·
44 Biotic recovery · Depositional environments

- 45
- 46

47 **1. Introduction**

48 The biotic recovery after the Permian-Triassic mass extinction is usually considered to 49 have lasted as long as the entire Early Triassic (Erwin 2001; Flügel 2002). The patterns and 50 causes of this delayed recovery are strongly debated, and possibly involve environmental, 51 ecological and preservational factors (Erwin 1996; Payne and Clapham 2012; Pietsch et al 52 2014; Tang et al. 2017; Bagherpour et al., 2017). Even if diagenetic processes can alter 53 primary geochemical signals (Thomazo et al. 2016), Early Triassic sediments record some of 54 the largest known Phanerozoic carbon isotope excursions (Atudorei 1999; Payne et al. 2004; 55 Galfetti et al., 2007; Grasby et al., 2013). Such global and large carbon isotope fluctuations 56 may reflect severe environmental disturbances - e.g. anoxia, euxinia, hypercapnia, high 57 temperature, ocean acidification, nutrient-rich waters – that postponed or interfered with the 58 biotic recovery (Wignall and Twitchett 1996; Payne et al. 2010; Sun et al. 2012; Grice et al. 59 2005; Knoll et al. 2007; Grasby et al. 2013). Moreover, the amount of skeletal organisms in 60 the aftermath of the Permian-Triassic extinction was apparently unusually reduced (Baud et 61 al. 1997; Payne et al. 2006). This managed to an increase in the calcium carbonate saturation 62 state that could explain the flourishing of microbialites in the Early Triassic (Pruss and Bottjer 63 2004; Pruss et al. 2006; Bottjer et al. 2008; Ezaki et al. 2008, 2012; Woods 2014; 64 Abdolmaleki and Tavakoli 2016). However, several authors have argued that some Early 65 Triassic microbialites developed in normal marine settings rather than in harsh environmental 66 conditions (Schubert and Bottjer 1992; Olivier et al. 2014, 2016; Collin et al. 2015; Vennin et 67 al. 2015; Fang et al. 2017). The presence of Early Triassic microbial deposits should also be 68 analysed by taking into account that a significant number of lineages survived in potential 69 refugia, and that skeletal organisms can be observed either as intercalated with or included 70 into microbialites, for instance just after the Permian-Triassic boundary (Brühwiler et al. 2008; Kaim et al. 2010; Hautmann et al. 2011; Forel et al. 2013; Bagherpour et al. 2017; Tang 71 72 et al. 2017). Some authors now regard these Early Triassic microbial deposits as part of 73 diverse, ecologically normal marine ecosystems, qualifying them as "microbialite metazoans"

(Yang et al. 2015). Consequently, the debate hinges on whether Early Triassic microbialites occurred when environmental conditions were hostile to metazoans – notably for CaCO₃precipiting invertebrates (Pörtner et al. 2005), or because the surviving metazoans were too decimated to rapidly recolonize shallow-marine habitats (Algeo et al. 2011). Therefore, detailed sedimentological studies are of primary importance for documenting the palaeoenvironments in which Early Triassic microbial deposits and skeletal metazoans cooccurred (Kershaw 2017).

81 Along the western margin of Pangea, the Sonoma Foreland Basin of the western USA 82 was the site of Early Triassic marine sedimentation (Collinson et al. 1976; Paull and Paull 83 1994; Dickinson 2013; Caravaca et al. 2017). In southwestern Utah, the Lower Triassic 84 sediments exhibit transitional deposits between the marine Thaynes Group and the continental 85 Moenkopi Group (sensu Lucas et al. 2007a). The Hurricane Cliffs expose various 86 sedimentary deposits such as conglomerates, sandstones, and microbial and fossiliferous 87 carbonates (Gregory 1950; Blakey 1979; Lucas et al. 2007a). Several areas give access to 88 stratal architectures that suggest rapid lateral facies variations over several tens of metres 89 (Nielson 1991; Olivier et al. 2014). About four kilometres south of the Hurricane airport, near 90 the head of Workman Wash, exceptional Early Triassic outcrops document the nature and the 91 spatial relationship between late Smithian microbialites and coeval fossiliferous deposits. The 92 aim of this paper is (i) to describe the main sedimentary succession at Workman Wash and to 93 interpret the evolution of the depositional settings, (ii) to demonstrate that microbial and 94 fossiliferous limestones coexisted as lateral facies belts influenced by relative sea-level 95 changes, and (iii) to discuss the environmental and ecological significance of such a 96 microbial-metazoan association in the aftermath of the Permian-Triassic biotic crisis.

- 97
- 98

99 2. Geological framework

During the Early Triassic, the western USA was located at a near-equatorial position, on the western margin of Pangea (Fig. 1b). During the Sonoma Orogeny, the onset of the Golconda Allochthon led to the formation of a north-south foreland sedimentary basin, which covered a large area including eastern Nevada, Utah, Idaho and parts of Wyoming (Collinson et al. 1976; Dickinson, 2006, 2013; Caravaca et al. 2017). From the north towards the southsouthwest, the Sonoma Foreland Basin recorded a major transgressive trend during the Smithian (Paull and Paull 1993; Goodspeed and Lucas 2007; Brayard et al. 2013). The presence of heterogeneities of the basin basement generated differential flexural subsidence,which impacted the sedimentary record and thicknesses (Caravaca et al. 2017).

109 The study area is located in Washington County (southwestern Utah, USA), about 8 110 km south of Hurricane city, near the head of Workman Wash between the Three Brothers and 111 White Face heights (37°5'56.62", 113°17'44.60", Fig. 1a). The studied Permian-Triassic 112 succession is capped by the Three Brothers volcanic rocks, which are remnants of early 113 Pleistocene lava (http://geology.utah.gov/apps/intgeomap/index.html; Hayden, 2004). The 114 Permian-Triassic transition can be observed on the western side of the Hurricane Cliffs, 115 whereas only Triassic rocks outcrop on its eastern side along Workman Wash (Fig. 2). Blakey 116 (1979) was the first to illustrate this area that Brayard et al. (2015) and Jattiot et al. (2017) 117 later referred to as "Black Rock Canyon". Lucas et al. (2007a) recently revised the 118 stratigraphic nomenclature of Early Triassic rocks in southwestern Utah with a lectostratotype 119 section from Timpoweap Canyon, located a couple of kilometres northeast of Hurricane. For 120 these authors, Early Triassic rocks of the Hurricane Cliffs begin with a chert breccia unit of 121 the Rock Canyon Conglomerate Formation (Moenkopi Group), which is overlain by a second 122 unit comprised of the various types of limestones and calcareous shales of the Sinbad 123 Formation (Thaynes Group). Finally, limestones of the Sinbad Formation are capped by 124 clastic deposits of the Lower Red Formation (a new digitation of continental deposits of the 125 Moenkopi Group; see Figure 2 of Lucas et al. 2007a for subdivisions and lateral correlations 126 between stratigraphic units of Thaynes and Moenkopi groups). These Triassic deposits rest 127 unconformably on silicified and recrystallized mudstones of the Permian Kaibab Formation 128 (Reeside and Bassler 1922; Gregory 1950; Blakey 1979; Stewart et al. 1972; Nielson and 129 Johnson 1979; Nielson 1991; Olivier et al. 2014). The presence of the ammonoid genera 130 Owenites, Anasibirites and Wasatchites in the Timpoweap Formation (= junior synonym of 131 the Sinbad Formation) at Virgin Dam and near Cedar City indicates a middle to late Smithian 132 age for this stratigraphic unit in this area (Lucas et al. 2007a; Brayard et al. 2013).

133

134

3. Studied sections and lithological units

Five sections have been logged between the Three Brothers and the White Face volcanic peaks (Fig. 1). Section #1 is located on the western side of Hurricane Cliffs (Fig. 2a). About 300 m away, four additional sections (#2-5) have been logged along the northern flank of Workman Wash (Fig. 2b). Among them, section #1 provides the most complete sedimentary succession of Early Triassic deposits that cap Permian limestones of the Kaibab

Formation (Figs. 3 and 4). These latter deposits correspond to a succession of dolostones, 141 142 dolosiltstones and bioclastic limestones that are more or less dolomitized, silicified and 143 bioturbated. This Permian succession is interrupted by a pluri-metre thick channelized breccia 144 interval. Just below this conglomeratic interval, Permian limestones reveal an extensive 145 karstification extending up to 2 metres thick in this area. A second karstified interval is 146 observed approximately 3 metres below a surface that probably represents the top of the 147 Kaibab Formation and the Permian-Triassic transition (Fig. 4). Above this surface, the Early 148 Triassic sedimentary succession is approximately 70 metres thick. Unfortunately, scree slopes 149 hide the outcrop 30 metres before the Early Pleistocene basalts that cap the section (Fig. 2b). 150 This Early Triassic succession displays in ascending order deposits of the Rock Canyon 151 Conglomerate, Sinbad, and Lower Red formations. Along section #1, deposits of the Rock 152 Canyon Conglomerate Formation are about 17 metres thick (Fig. 4). They can be subdivided 153 into 4 successive lithological units. The first one corresponds to a first yellow clastic unit that 154 is at least 3.5 metres thick. It is overlain by a 3 metres thick red bed unit that is itself capped 155 by a second yellow clastic unit. A 5 metre thick conglomeratic breccia unit marks the end of 156 the Rock Canyon Conglomerate Formation. Above it, the Sinbad Formation consists of a 157 single 18 metre thick fenestral-microbial limestone unit. A third level of yellow clastic 158 deposits characterizes the uppermost part of the section, which is considered here to belong to 159 the Lower Red Formation. Section #2 is only 5 metres thick and can be subdivided into two 160 lithological units of the Sinbad Formation (Fig. 5). Most of this section corresponds to a first 161 fenestral-microbial unit. The second lithological unit made of bioclastic limestones is only 162 visible in the first decimetres of the section. Section #3 is around 19 metres thick and can be 163 subdivided into 4 lithological units (Fig. 6). A basal bioclastic unit, capped by a 12 metre 164 thick fenestral-microbial unit and a single bed of a pel-fenestral unit represent the deposits of 165 the Sinbad Formation. The uppermost part of the section corresponds to the yellow limestone unit of the Lower Red Formation. Section #4 is 16 metres thick and consists of four 166 167 lithological units (Fig. 7). A basal breccia unit outcrops over a thickness of one metre; it is 168 capped by a 4 metre thick fenestral-microbial unit. Above this rests a third bioclastic 169 lithological unit of more than 7 metres thick. These first three units belong to the Sinbad 170 Formation. The uppermost part of the section corresponds to the yellow clastic unit of the 171 Lower Red Formation. Section #5 corresponds to one single, 15 metre thick fenestral-172 microbial unit of the Sinbad Formation (Fig. 8).

- 173
- 174

175 **4. Facies descriptions**

A bed-by-bed macro- and microscopic sedimentological analysis of the five sections, which included the observation of more than 150 thin sections, allowed us to identify 13 main facies (Table 1). These facies can be grouped into several categories, including terrigenous (F1, F2), dolomitized (F3-F6), fenestral-microbial limestones (F7) and fossiliferous (F8-F13) deposits. The fenestral-microbial limestones (F7) appear as the most complex facies in composition and the most common at the scale of the outcrop.

- 182
- 183

33 4.1. Fenestral-microbial limestones (Facies F7)

184 Limestones with microbial laminations and fenestrae occur in the five studied sections 185 (Fig. 9A). Macroscopically, the fenestral-microbial beds sometimes appear truncated, and 186 more commonly display lateral thickness variations, forming dm-scale lens-like to flat 187 morphologies (Fig. 10). These fenestral-microbial beds can also locally interfinger with 188 breccia (F1) or bioclastic (F10) deposits (e.g. section #5 in Fig. 8). Another feature of this 189 fenestral-microbial facies is to record common lenses of macroscopic skeletal organisms such 190 as gastropods and ammonoids (Figs. 9C-D and 10). Brayard et al. (2015) identified three main 191 taxa of gastropods in these levels, corresponding to "Coelostylina sp. A", Angularia sp., and 192 Worthenia windowblindensis. All these taxa are large-sized gastropods, with shell sizes 193 reaching ~ 10 cm in length for the largest sampled specimens of "Coelostylina sp. A". 194 Sampled ammonoids correspond to a rich and well-preserved prionitid fauna, typical of the 195 late Smithian Anasibirites multiformis beds in the western USA Basin (Brayard et al. 2013; 196 Jattiot et al. 2016, 2017), including Anasibirites kingianus, A. multiformis, Wasatchites 197 perrini, Hemiprionites walcotti, H. typus and Arctoprionites resseri.

198 Microscopically, F7 is relatively diverse in composition and texture, making it 199 possible to distinguish 5 subfacies (F7a-e; Table 1). In some cases, fenestral-microbial 200 deposits are made of abundant pisoids (subfacies F7a; Fig. 11A) or intraclasts (subfacies F7e; 201 Fig. 11B). Subfacies F7b is characterized by large cm- to dm-scale fenestrae (Fig. 9B). 202 Diversified endostromatolites, commonly made of irregular to domal micritic laminae or 203 sparitic laminae (Fig. 11C-E), developed in the fenestrae. Acicular to botryoidal cements are 204 also observed associated with these endostromatolites (Fig. 11F). Subfacies F7c is 205 characterized by lamellar siliceous sponges co-occurring with micritic stromatolites and 206 common ostracods (Fig. 11G and H). Locally, this subfacies records some root-like structures 207 and circum-granular cracks (Fig. 12A). Subfacies F7d corresponds to microbial-fenestral 208 deposits that display a conglomeratic nature, with abundant polygenic and subangular to subrounded lithoclasts (Fig. 12B). A common feature shared by all these subfacies includes the presence of frequent biofilms that are a few tens of microns thick and that have a micritic nature. These biofilms are observed coating different types of substrates, such as pel-oncoidfenestral deposits, pel-ooidal grainstones, or gastropods and ammonoids (Fig. 12C-E). In other cases, the biofilms developed micritic stromatolites with microscopic planar, irregular to domal morphologies (Fig. 12F and G). Locally, some cement crusts show enigmatic structures (Fig. 12H).

- 216
- 217

7 4.2. Fossiliferous limestones (Facies F8-F13)

218 Six facies (F8-F13) with a moderatly diversified fauna (including echinoderms, 219 gastropods, siliceous sponges, ostracods, bivalves, lingulid brachiopods, and phosphatic 220 remains) and a more or less intense degree of bioturbation are identified (Table 1). These 221 facies are distinguished according to their textures, sedimentary structures, as well as their 222 grain abundances. Facies F8 is a mud-supported limestone with frequent echinoderms. Facies 223 F9 is an intensively bioturbated packstone (locally wackestone) with abundant echinoderms 224 and frequent siliceous sponge spicules (Fig. 13A). Facies F10 is a bioturbated grain-supported 225 bioclastic (mainly gastropods and bivalves) limestone that also includes abundant ooids (Fig. 226 13B).

227 Facies F11 corresponds to an intensively bioturbated and bioclastic packstone 228 composed of abundant bioclasts of gastropods, echinoderms, bivalves and vertebrate 229 phosphate remains. Beds attributed to this facies displays a spectacular abundance of well-230 preserved lingulid brachiopods frequently preserved in their burrows (Figs. 9E and 14). Such 231 a remarkable macroscopic assemblage deserves a short description here as it has important 232 implications in terms of depositional setting and palaeoecology. A majority of lingulid 233 specimens are complete articulated shells preserved perpendicular to the bedding, with the 234 ventral side upward, showing that they are preserved *in situ* and in life position. The shells are 235 elongated, twice as long as wide. Most measurable specimens are between 15 and 20 mm in 236 length; a few complete specimens reach 21 mm. The dorso-ventral thickness is about 5 mm. 237 The posterior part of the shell is rounded, the anterior part is spade-shaped and the lateral 238 borders are parallel. Although no internal structure could be observed, these shells are 239 tentatively attributed to Lingularia sp., the most frequent Mesozoic lingulid genus (Biernat 240 and Emig 1993; Zonneveld et al. 2007; Posenato et al. 2014; Holmer et al. 2016). 241 Interestingly, the shell length of the specimens found at Workman Wash tends to exceed the 242 values reported for other Early Triassic lingulids (e.g. Zonneveld et al. 2007, Posenato et al.

243 2014, Peng et al. 2007). Several shells described here are concordantly emplaced in their 244 dwelling burrows that tend to be deep (about 150 mm), densely distributed locally (about 500 245 burrows/m²) and filled by a yellow-to-orange dolomitized micrite (Fig. 9E). Burrows are 246 cylindrical, vertical though not totally straight, and rounded to slightly elliptical in cross 247 section. Many of them correspond to pedicle traces whose diameters are particularly 248 homogeneous (5 mm in average). Several burrows are spreitenate and they preserve concave-249 up laminae whose width is comprised between 8 and 12 mm (Fig. 14B). Some of these 250 laminae are found immediately below the posterior part of *Lingularia* shells preserved *in situ* 251 and that have the same width. All these features warrant the identification of these trace 252 fossils as *Lingulichnus verticalis* that are typical lingulid equilibration traces (Hakes 1976; 253 Szmuc et al. 1977; Zonneveld and Pemberton 2003; Zonneveld et al. 2007). They are 254 associated with some U-shaped and J-shaped burrows that we identify as Lingulichnus 255 hamatus, an ichnofossils interpreted as a lingulid reburrowing structure (Zonneveld and 256 Pemberton 2003; Zonneveld et al. 2007). Some horizontal, more or less sinuous traces also 257 occur on the top of the beds representing facies F11. The dominance of vertical, cylindrical 258 and U-shaped burrows of suspension feeders, the frequent occurrence of equilibrichnia-259 related laminae, the relative rarity of horizontal structures and the low ichnological diversity 260 suggest an attribution of these beds to the Skolithos ichnofacies (Seilacher 1963; Frey and 261 Seilacher 1980; Frey and Pemberton 1984, 1985; MacEachern et al. 2007; Buatois and 262 Mángano 2011). Finally, it should be noted that the beds attributed to Facies F11 also yields 263 large shells of indeterminate bivalves.

Facies F12 is a bioclastic packstone-grainstone rich in peloids (Fig. 13C). Facies F13 is a grainstone (locally packstone) notably characterized by frequent intraclasts (Fig. 13D). Apart from Facies F8 and F9, these bioclastic limestones display dm- to m-scale trough cross bedding with low angle fore sets or small asymmetrical ripples, and common mud drapes for F10.

269

270 4.3. Terrigenous (F1-F2) and other carbonate facies (F3-F6)

In addition to the fenestral-microbial (F7) and bioclastic limestones (F8-F13), 6 other facies can be identified (Table 1). Facies F1 corresponds to a channelized conglomerate breccia with angular to subrounded polygenic clasts (Fig. 9F). Facies F2 is a thin-bedded dolosiltstone with common bioturbation, trough cross stratifications and asymmetrical ripples that locally display some mud drapes. The remaining facies are carbonate-dominated, but terrigenous components can be sometimes observed. F3 corresponds to a dolomudstone characterized by some bioturbation and no preserved fauna. F4 is a dolostone devoid of fauna and sedimentary structures. F5 is a silty peloidal packstone with some bivalves, rare gastropods and some micritized ooids. This facies is characterized by some mud clasts, mud drapes and trough cross bedding with low angle foresets. F6 is a wackestone/packstone limestone with keystone vugs. It is characterized by frequent reworked clasts of microbial crusts and rare bioturbation.

- 283
- 284
- 285

5. Multi-scale lateral facies variations

286 Figure 15 shows the correlation of the 5 studied sections, along which no evidence of 287 faults was found. Such a correlative scheme is possible because some beds can be laterally 288 traced across the sections. First, a 20-30 cm thick bed, particularly rich in lingulid 289 brachiopods in section 4, can be traced laterally through sections #3 and #2. This "lingulid" 290 marker bed was not found in section #1 and could not be followed laterally into section #5 291 because of ground vegetation (Fig. 2B). Second, the correlation of section #1 is attempted 292 through its conglomerate unit that has a lateral equivalent observed at the base of section #4. 293 Last, the position of section #5 was estimated with a tolerance of few centimetres, in laterally 294 following as far as possible successive beds of sections #4 and #5.

295 At a first order, the stratigraphic succession of the Black Rock Conglomerate, Sinbad 296 and Lower Red formations can be easily observed laterally although some lithological units 297 display important thickness variations. For example, the yellow clastic unit 3 of section #4 298 stratigraphically starts much lower compared to other sections (Fig. 15). This could be 299 explained in two ways. First, the yellow clastic unit 3 could lay unconformably over the 300 fenestral microbial unit, thus explaining changes in thickness between the different sections. 301 Second, there could be a lateral facies variation between the fenestral microbial unit and this 302 yellow clastic unit. No evidence of a significant erosional surface has been identified in the 303 field. Thus, the studied locality likely records lateral facies variations at the scale of few 304 hundreds of metres. Such lateral facies variations are well expressed in the stratigraphic 305 distribution of the bioclastic unit. Between sections #3 and #4, the bioclastic facies F12 is 306 observed laterally to different subfacies of the fenestral-microbial limestone (F7) over a few 307 tens of metres. Located more than 100 m away, section #5 also records a thin bed of bioclastic 308 limestone (F10), highlighting again the lateral proximity between fossiliferous and microbial 309 deposits. In section #5, the intercalation of a dm-scale thick bed of breccia also emphasizes 310 that such terrigenous sediments are recurrent, even above the Rock Canyon Conglomerate Formation. These observations demonstrate important lateral facies changes over a few tens of metres in the Hurricane Cliffs area, whereas previous studies only presumed their existence after larger-scale lithostratigraphic correlations (Blakey 1979; Nielson and Johnson 1979).

314 Some facies changes can also be laterally traced at the scale of a single bed. For 315 example, across sections #2, #3 and #4, the "lingulid" marker bed exhibits at least two facies. 316 Over a few tens of metres (Fig. 15), this marker bed is made of a packstone rich in siliceous 317 sponge spicules (F9), which evolves into an intensively bioturbated limestone with some 318 lingulids, echinoderms, bivalves and gastropods (F11). As indicated by the number of 319 identified subfacies, the fenestral-microbial limestone (F7) is probably the facies displaying 320 the most important changes in terms of composition. Even within a subfacies, the distribution 321 of some structures such as biofilms or stromatolites has a restricted lateral extension. 322 Likewise, the presence and abundance of some coated grains such as ooids, oncoids, or 323 pisoids can shift markedly at the scale of a thin section, not to mention some macroscopic 324 organisms (ammonoids, gastropods, or ostracods) whose distribution is highly heterogeneous 325 in this fenestral-microbial limestone. Indeed, some accumulations of gastropods are observed 326 only in some local dm-scale lenses (Fig. 9C). Similarly, ammonoids also accumulated in 327 some (pluri)dm-scale lenses (Fig. 9D). Additionally, ammonoids are preferentially 328 concentrated in some specific beds and over a few tens of decimetres (Fig. 10). On a smaller 329 scale, some microscopic skeletal organisms such as ostracods also display a similar 330 heterogeneous distribution. In these fenestral-microbial limestones, ostracods are usually 331 observed associated with siliceous sponges and stromatolites within sediments displaying a 332 micritic fabric (Fig. 11G).

- 333
- 334

6. Depositional environments and relative sea-level fluctuations

336 Observed sedimentary features point towards a mixed siliciclastic-carbonate marine 337 depositional system (Fig. 16), whose minimum depth is probably localized within an area 338 corresponding to a supratidal-subtidal interior platform showing the deposition of fenestral-339 microbial limestones (F7). This facies locally records multiple phases of emersions, as 340 emphasized by frequent pisoids, mud cracks, root structures and truncation surfaces (Figs. 10, 341 11A, 12A). This fenestral-microbial facies belt thus likely separated an inner tidal flat from a 342 subtidal interior platform. In the inner tidal flat, terrigenous deposits (F2) dominated the 343 sedimentation, whereas carbonates only occurred as isolated muddy pools (F3). Additionally, 344 the correlation scheme suggests that the fenestral-microbial facies belt was not laterally 345 continuous (Fig. 15), but was intersected by broad channels into which fossiliferous 346 limestones were deposited (F13). Facies F13 includes sedimentary grains (intraclasts) derived 347 from the fenestral-microbial limestones and bioclasts (echinoderms) characteristic of more 348 open marine settings. These latter settings correspond to the subtidal interior platform where 349 various fossiliferous limestones (F8-F12) were deposited. Both in the inner tidal flat and in 350 the subtidal interior platform, the presence of megaripples with mud drapes and mud clasts 351 emphasizes a tide-influenced regime. Some fossiliferous limestones (F8 and F9) appear 352 devoid of tide- or wave-generated sedimentary structures, suggesting the presence of some 353 protected areas. The presence of breccia (F1), notably intercalated within the fenestral-354 microbial limestones, indicates the proximity of the paleoshoreline, echoing previous 355 observations by Olivier et al. (2014) in the vicinity of Workman Wash. This facies (F1) 356 displays poorly sorted and angular clasts of cherts and limestones that suggest debris flows 357 derived from the Permian Kaibab Formation. Such facies models also fit well with previous 358 paleoenvironmental reconstructions, which highlighted a sedimentary system characterized by 359 a clear polarity with an open sea that stood east of the Hurricane Cliffs (Blakey, 1979). The 360 relatively small size of the Workman Wash depositional system, also characterized by a flat 361 slope and a tidal autocyclic character, generated the rapid facies displacements and the 362 important lateral facies variations observed at the scale of a few tens to hundreds of metres (Fig. 15). 363

364 The studied Early Triassic stratigraphic column, which involves three successive 365 formations (Rock Canyon Conglomerate, Sinbad, Lower Red), displays a clear evolution of 366 the type of sediments and thus of depositional settings (Figs. 15 and 16). The transition from 367 proximal and shallow deposits (F2-F4) of the siliciclastic units (Rock Canyon Conglomerate 368 Formation) to the carbonates (F7) of the fenestral-microbial unit (Sinbad Formation) is 369 consistent with an increase in accommodation coupled with the general retrogradation of 370 these facies belts. The high accumulation of fenestral-microbial limestones (F7) explains the 371 frequent traces of emersion recorded by this facies, despite a more distal and deeper position 372 in the interior platform. The installation of bioclastic and bioturbated facies in the Anasibirites 373 multiformis beds (Brayard et al. 2013; Jattiot et al. 2017) likely reflects a maximum of 374 accommodation recorded in the Workman Wash area. The return of inner tidal flat siliciclastic 375 deposits (F5) in the yellow clastic unit 3 (Lower Red Formation) probably marked an 376 accommodation decrease and a rapid facies progradation. Such facies evolution fits well with 377 the Smithian third order transgressive-regressive sequence recorded in the western USA

Lower Triassic succession (Paull and Paull 1993, 1997), which is characterized by an earlymiddle Smithian transgression (Goodspeed and Lucas 2007; Brayard et al. 2013).

- 380
- 381

382 7. Ecological and environmental implications

383 It is usually assumed that a return to pre-extinction levels of taxonomic and functional 384 diversity after the end-Permian extinction crisis did not occur until the end of the Early 385 Triassic, or at least late in the Spathian (Schubert and Bottjer, 1995; Flügel 2002; Payne et al. 386 2006; Lehrmann et al. 2006; Chen and Benton 2012; Pietsch and Bottjer 2014; but see e.g. Brayard et al. 2009, 2017; Foster et al. 2015). In the presumed absence of developed benthic 387 388 faunas, Early Triassic microbialites have been initially considered as disaster forms (i.e. 389 opportunistic generalist taxa sensu Schubert and Bottjer 1992). These disaster microbialites 390 were first considered to have formed in normal marine settings (Schubert and Bottjer 1992, 391 1995), like several subsequent studies (Kershaw et al. 1999; Lehrmann 1999; Lehrmann et al. 392 2001; Ezaki et al. 2008; Yang et al. 2011). Nevertheless, as noted by Tang et al. (2017), Early 393 Triassic microbialites were progressively reinterpreted to have flourished as a consequence of 394 harsh environmental conditions, such as hot temperatures, high carbonate saturation, or 395 elevated CO₂ (Pruss and Bottjer 2004; Pruss et al. 2006; Mary and Woods 2008; Mata and 396 Bottjer 2011; Pietsch et al. 2014; Song et al. 2014; Abdolmaleki and Tarakoli 2016). 397 Lehrmann et al. (2015) suggested that ecological relaxation and abnormal ocean chemistry 398 were not mutually exclusive to explain the proliferation of Early Triassic microbial deposits. 399 However, diversified assemblages have been documented from various Early Triassic 400 intervals and latitudes (Krystyn et al. 2003; Twitchett et al. 2004; Beatty et al. 2008; Kaim et 401 al. 2010; Hautmann et al. 2011, 2013; Hofmann et al. 2011, 2013a, b, 2014; Baud et al. 2015; 402 Fu et al. 2016; Foster et al. 2017; Brosse et al. in press). Moreover, the onset of complex 403 ecosystems is now documented to have occurred rapidly after the end-Permian mass 404 extinction (Haig et al. 2015; Brayard et al. 2017). Hence, the scenario of an ecological 405 vacation that led to a global post-extinction delayed recovery has become increasingly 406 unlikely. Moreover, new evidences that Early Triassic microbialites could have formed in 407 normal marine environments are growing (Olivier et al. 2014, 2016; Vennin et al. 2015; Tang 408 et al. 2017; Bagherpour et al. 2017). Therefore, the causes of the Early Triassic microbialite 409 proliferation and their relationships with a delayed biotic recovery scenario remain highly 410 controversial. This notably results from the poor knowledge of the physico-chemical 411 parameters of Early Triassic seas into which microbialite developed. Indeed, such 412 palaeoenvironments are reconstructed from analyses of the microbialites themselves without 413 information from lateral coeval sediments. Sedimentological and palaeontological data from 414 other types of coeval sediments can indeed indirectly provide insights on the chemical 415 composition of waters in which microbial deposits developed (Bagherpour et al. 2017).

416

417 Among the four episodes of microbialite expansion initially identified for the Early 418 Triassic (Pruss et al. 2006; Baud et al. 2007), only the Permian-Triassic boundary 419 microbialites (PTBMs) have a geographical extent large enough to have a global significance 420 (Kershaw et al. 2012). Deep-water forms of PTBMs with metazoans are known and recently 421 described by Baud et al. (2017) and by Friesenbichler et al. (2018). Within interior platform 422 PTBMs, skeletal remains of some brachiopods, bivalves, microconchids, gastropods, 423 echinoderms and sparse ammonoids are mainly observed as interstratified beds or lenses 424 (Lehrmann et al. 2001; Baud 2007; Hautmann et al. 2015; Tang et al. 2017; Bagherpour et al. 425 2017). Thus, reliable ecological and environmental information from contemporaneous 426 deposits of PTBMs remains to be documented in more detail.

427 Post-PTBMs microbial episodes in the Early Triassic have much more restricted 428 geographic distributions (Mata and Bottjer 2012). Those observed in the southwestern USA 429 basin are crucial since local and regional studies suggest the existence of lateral correlations 430 between Smithian microbialites and different types of sediments containing an abundant and 431 diversified fauna (i.e. brachiopods, bivalves, gastropods, ostracods, echinoderms, sponges and 432 ammonoids; Stewart et al. 1972; Blakey 1974, 1977; Nielson and Johnson 1979; Dean 1981; 433 Nielson 1991; Goodspeed and Lucas 2007; Ritter et al. 2013; Olivier et al. 2016). However, a 434 major challenge on the field lies in the capacity to observe and to demonstrate the lateral 435 transition and the contact between microbial deposits and these open marine skeletal 436 limestones. Such lateral facies variation has been observed along an outcrop a few tens of 437 meters long in Timpoweap Canyon, north of the Hurricane Cliffs (Olivier et al. 2014). At this 438 site, a fenestral-microbial limestone deposited in contact with a dolomudstone characterized 439 by sparse bioturbations and rare bioclasts (serpulids, bivalves and echinoderms). A few 440 meters away, this mudstone evolves into a bioturbated limestone with a wackestone (locally 441 packstone) texture and some gastropods, serpulids, bivalves and echinoderm plates. However, 442 the limited extent of this outcrop does not provide a good spatial comprehension of its 443 depositional setting.

444 At Workman Wash, the lateral continuity of deposits over a few hundred meters 445 highlights the lateral transition between fenestral-microbial deposits and limestones 446 characterized by a relatively abundant metazoan fauna (Figs. 15 and 16). With some ostracods 447 and gastropods, the fauna observed in the fenestral-microbial facies is neither diversified nor 448 abundant, reflecting a microbe-dominated ecosystem. As microbialites flourished on this 449 shallow sea floor of the internal platform, they were potentially a profuse resource for grazing 450 gastropods (Batten and Stokes 1986; Schubert and Bottjer 1995; Pietsch et al. 2014), although 451 this hypothesis remains to be corroborated (see discussion in Brayard et al. 2015). 452 Furthermore, sponges are also observed associated with microbialites (Fig. 11G, H). This 453 confirms the common occurrence of these metazoan reef-builders in Early Triassic reefs 454 (Brayard et al. 2011; Marenco et al 2012; Baud 2013, Vennin et al. 2015; Olivier et al. 2016; 455 Baud et al. 2017; Friesenbichler et al. 2018). It implies that the local trophic network not only 456 consisted of primary but also meso-consumers such as sponges. The common lenses of 457 ammonoids preserved in the Workman Wash microbialites indicate the presence of higher-458 level consumers. The abundant lingulid brachiopods observed in a 30 cm thick interval could 459 also be interpreted as opportunist meso-consumers (Rodland and Bottjer 2001). Thus, 460 considered separately, both the microbial or lingulid-rich facies actually do not illustrate an 461 ecosystem with a trophic level higher than microbe-dominated and possibly opportunist-462 dominated ones (Chen and Benton 2012). However, even if lingulid brachiopods are 463 effectively common in some beds, the facies F11 in which they occur also includes frequent 464 to abundant fragments of crinoids, gastropods and bivalves (Fig. 16). These observations 465 better fit recent studies that point toward lingulids as ecological opportunists that dominated 466 locally in some Early Triassic shallow marine environments rather than a disaster taxon 467 (Zonneveld et al. 2007; McGowan et al. 2009). Other adjacent deposits of microbialites such 468 as the facies F9 also comprises abundant elements of crinoids and frequent sponge spicules, 469 coupled with an abundant infaunal activity. Overall, the depositional system reconstructed at 470 Workman Wash shows that microbial and lingulid-dominant deposits are coeval with a 471 moderately diversified benthic metazoan ecosystem. Such a biocoenosis including siliceous 472 sponges (either present in reefs or in lateral deposits), crinoids, lingulid brachiopods, large-473 size gastropods and higher-level consumers, such as ammonoids and vertebrates, indicates 474 that some Early Triassic marine ecosystems were not ecologically depauperate (Krystyn et al. 475 2003; Foster and Twitchett 2014; Baud et al. 2015; Brosse et al. in press). Additionally, the 476 actual diversity of the Workman Wash palaeocommunity is probably underestimated, as 477 suggested by ammonoids that are only preserved in the microbial-fenestral facies. Indeed, 478 microbial deposits are known to favour rapid sediment stabilization and early lithification 479 (Gall 1990; Tomescu et al. 2016). This may explain a differential preservation of some 480 organisms such as ammonoids. All these mechanisms highlight how differential taphonomic481 pathways can bias the fossil record in some Early Triassic marine settings.

482

483 From an environmental point of view, the presence of stenohaline organisms such as 484 echinoderms in deposits lateral to the microbial-fenestral facies indicates normal marine 485 conditions. The abundant lingulids observed in facies F11 also reflect an intense infaunal 486 activity, implying the presence of nutrient-rich waters. The proximity of emerged lands may 487 have provided an adequate nutrient supply to the depositional system, which in turn could 488 have promoted the development of microbialites (Algeo et al. 2011; Olivier et al. 2016). In 489 subtidal settings of the inner tidal flat (Fig. 16), the presence of an important terrigenous flux 490 and intense sediment mobility (caused by tidal currents) could have inhibited microbialite 491 development (Olivier et al., 2016). In such siliciclastic peritidal environments, microbial 492 deposits are nevertheless known in other Early Triassic successions of the Sonoma Foreland 493 Basin as microbially induced sedimentary structures (MISS; Grosjean et al., 2018). In the 494 more open subtidal settings of the interior platform, the abundance of lingulid equilibration 495 structures (in response to burial) and reburrowing structures (in response to exhumation) are 496 consistent with a tidal regime. This relatively intense infaunal activity, coupled with the 497 migration of tidal bars, may have prevented the development of microbialites in the more 498 distal parts of the Workman Wash depositional setting.

- 499
- 500

501 **8. Conclusions**

502 For the first time, the late Smithian rocks of Workman Wash in the Northern 503 Hurricane Cliffs (southwestern Utah, USA) provide a direct and continuous observation of the 504 lateral facies transition between microbialites and contemporaneous fossiliferous limestones 505 in the Early Triassic. Facies analysis reveals a shallow and proximal tide-dominated and 506 mixed siliciclastic-carbonate sedimentary system. It was located along the southwestern 507 margin of an epicontinental sea tongue corresponding to the Sonoma Foreland Basin. Above 508 Permian limestones, the studied interval includes breccia and dolosiltstones of the Rock 509 Canyon Conglomerate Formation, fenestral-microbial and bioclastic limestones of the Sinbad 510 Formation, and yellow dolosiltstones of the Lower Red Formation. This lithological 511 succession is in accordance with the Smithian third-order transgressive-regressive sequence 512 recorded in the western USA.

513 Located in an interior platform and over only a few hundreds of meters, intertidal to 514 subtidal microbialites pass laterally to bioturbated and fossiliferous limestones containing a 515 moderately abundant and diversified fauna including echinoderms, lingulid brachiopods, 516 gastropods, bivalves, ostracods, siliceous sponges and vertebrate remains. Such a faunal 517 assemblage coeval with microbialites reveals a moderately diversified biocoenosis, which is 518 in stark contrast to traditional views of depauperate ecosystems in the Early Triassic. The 519 fossiliferous limestones comprising stenohaline organisms also suggest normal marine 520 conditions contrary to the harsh environmental conditions classically invoked to explain the 521 presence of microbialites. At Workman Wash, microbialites rather reflect a shallow and 522 proximal facies belt controlled by terrigenous flux, nutrient inputs and relative sea-level 523 fluctuations.

- 524
- 525
- 526
- 527

528 Acknowledgements

529 This work is a contribution to the ANR project AFTER (ANR-13-JS06-0001-01). The CNRS 530 INSU Interrvie, and the French ANR @RAction grant (project EvoDevOdonto) also 531 supported this study. D.A. Stephen is grateful for the ongoing financial support of the College 532 of Science & Health at Utah Valley University. Michael Hautmann is thanked for his 533 assistance in bivalve taxonomy and ecology. Our thanks to Marilyne Imbault for her 534 contribution to the ammonoid determination. The Workman Wash area is located on US 535 public land under the stewardship of the Bureau of Land Management (BLM) of the US 536 Department of the Interior; access to this land is gratefully acknowledged. We would like to 537 thank Wolfgang Kießling and two anonymous reviewers for their helpful comments.

- 538
- 539
- 540
- 541

542 **References**

Abdolmaleki J, Tavakoli V (2016) Anachronistic facies in the Early Triassic successions of
the Persian Gulf and its palaeoenvironmental reconstruction. Palaeogeogr Palaeoclimatol
Palaeoecol 446:213–224

- Algeo TJ, Chen ZQ, Fraiser ML, Twitchett RJ (2011) Terrestrial-marine teleconnections in
 the collapse and rebuilding of Early Triassic marine ecosystems. Palaeogeogr
 Palaeoclimatol Palaeoecol 308:1–11
- 549 Atudorei NV (1999) Constraints on the Upper Permian to Upper Triassic marine carbon
 550 isotope curve. Case studies from the Tethys. PhD thesis, University of Lausanne:1–155
- 551 Bagherpour B, Bucher H, Baud A, Brosse M, Vennemann, T, Martini R, Guodun K (2017)
- 552 Onset, development, and cessation of basal Early Triassic microbialites (BETM) in the 553 Nanpanjiang pull-apart Basin, South China Block. Gondwana Res 44:178–204
- Batten RL, Stokes WL (1986) Early Triassic gastropods from the Sinbad Member of the
 Moenkopi Formation, San Rafael Swell, Utah. American Mus Novit 2864:1–33
- Baud A (2007) Lower Triassic microbialites versus skeletal carbonates, a competition on the
 Gondwana Margin. N M Mus Nat Hist Sci Bull 41:23
- Baud A (2013) The Smithian (Early Triassic) red ammonoid limestone of Oman, refuge for
 sponge-microbial build-ups during recovery phase. GSA Annu Meet in Denver
- Baud A, Cirilli S, Marcoux J (1997) Biotic response to mass extinction: the lowermost
 Triassic microbialites. Facies 36:238–242
- Baud A, Richoz S, Pruss S (2007) The lower Triassic anachronistic carbonate facies in space
 and time. Global Planet Change 55:81–89
- Baud A, Friesenbichler E, Richoz S, Krystyn L, Sahakyan L, (2017) Induan (Early Triassic)
 giant sponge-microbial build-ups in Armenia. 5th IGCP 630 International conference and
 field workshop, Erevan 8-14 October 2017 Program and Abstract: 13.
- Beatty TW, Zonneveld JP, Henderson CM (2008) Anomalously diverse Early Triassic
 ichnofossil assemblages in northwest Pangea: A case for a shallow-marine habitable zone.
 Geology 36(10):771–774
- 570 Biernat G, Emig CC (1993) Anatomical distinctions of the Mesozoic lingulide brachiopods.
 571 Acta Palaeontol Pol38:1–20
- 572 Blakey RC (1974) Stratigraphic and depositional analysis of the Moenkopi Formation,
 573 Southeastern Utah. Utah Geol Miner Surv Bull 104:1–81
- 574 Blakey RC (1977) Petroliferous lithosomes in the Moenkopi Formation, Southern Utah. Utah
 575 Geol 4:67–84
- 576 Blakey RC (1979) Oil impregnated carbonate rocks of the Timpoweap Member Moenkopi
- 577 Formation, Hurricane Cliffs area, Utah and Arizona: Utah Geol 6:45–54

- Bottjer DJ, Clapham ME, Fraiser ML, Powers CM (2008) Understanding mechanisms for the
 end-Permian mass extinction and the protracted Early Triassic aftermath and recovery.
 GSA Today 18:4–10
- 581 Brayard A, Escarguel G, Bucher H, Monnet C, Brühwiler T, Goudemand N, Galfetti T, Guex
- J (2009) Good genes and good luck: Ammonoid diversity and the end-Permian mass
 extinction. Science 325: 1118-1121
- 584 Brayard A, Vennin E, Olivier N, Bylund KG, Jenks J, Stephen DA, Bucher H, Hofmann R,
- Goudemand N, Escarguel G (2011) Transient metazoan reefs in the aftermath of the endPermian mass extinction. Nature Geosci 4:693–697
- Brayard A, Bylund KG, Jenks JF, Stephen DA, Olivier N, Escarguel G, Fara E, Vennin E
 (2013) Smithian ammonoid faunas from Utah: implications for Early Triassic
- biostratigraphy, correlations and basinal paleogeography. Swiss J Palaeontol 132:141–219
- 590 Brayard A, Meier M, Escarguel G, Fara E, Nützel A, Olivier N, Bylund KG, Jenks JF,
 591 Stephen DA, Hautmann M, Vennin E, Bucher H (2015) Early Triassic Gulliver gastropods:
- spatio-temporal distribution and significance for the biotic recovery after the end-Permian
 mass extinction. Earth Sci Rev 146:31–64
- Brayard A, Krumenacker LJ, Botting JP, Jenks JF, Bylund KG, Fara E, Vennin E, Olivier N,
 Goudemand N, Saucède T, Charbonnier S, Romano C, Doguzhaeva L, Thuy B, Hautmann
 M, Stephen DA, Thomazo C, Escarguel G (2017) Unexpected Early Triassic marine
 ecosystem and the rise of the Modern evolutionary fauna. Science Adv 3:e1602159
- Brosse M, Bucher H, Baud A, Hagdorn H, Hautmann M, Nützel A, Ware D, Frisk Å,
 Goudemand N (in press) New data from Oman indicate benthic high biomass productivity
 coupled with low taxonomic diversity in the aftermath of the Permian-Triassic Boundary
 mass extinction. Lethaia.
- Brühwiler T, Brayard A, Bucher H, Guodun K (2008) Griesbachian and Dienerian (Early
 Triassic) ammonoid faunas from northwestern Guangxi and southern Guizhou (south
 China). Palaeontol 51:1151–1180
- Buatois LA, Mángano, MG (2011) Ichnology: Organism-substrate interactions in space and
 time. Cambridge University Press
- 607 Caravaca G, Brayard A, Vennin E, Guiraud M, Grosjean AS, Olivier N, Thomazo C, Fara E,
 608 Escarguel G, Bylund K, Jenks J (2017) Controlling factors for differential subsidence in
 609 the Sonoma Foreland Basin (Early Triassic, western USA). Geol Mag In press.
 610 10.1017/S0016756817000164

- 611 Chen ZQ, Benton, MJ (2012) The timing and pattern of biotic recovery following the end612 Permian mass extinction. Nature Geosci 5:375–383
- 613 Collin PY, Kershaw S, Tribovillard N, Forel MB, Crasquin S (2015) Geochemistry of post-
- extinction microbialites as a powerful tool to assess the oxygenation of shallow marine
 water in the immediate aftermath of the end-Permian mass extinction. Int J Earth Sci
 104:1025–1037
- 617 Collinson JW, Kendall CGSC, Marcantel JB (1976) Permian-Triassic boundary in eastern
- 618 Nevada and west-central Utah. Bulletin of the Geological Society of America 87:821–824
- 619 Dean JS (1981) Carbonate petrology and depositional environments of the Sinbad Limestone
- Member of the Moenkopi Formation in the Teasdale Dome Area, Wayne and GarfieldCounties, Utah. Brigham Young Univ Geol Stud 28:19–51
- 622 Dickinson WR (2006) Geotectonic evolution of the Great Basin. Geosphere 2:353–368
- Dickinson WR (2013) Phanerozoic palinspastic reconstructions of Great Basin geotectonics
 (Nevada-Utah, USA). Geosph 9:1384–1396
- Erwin DH (1996) Understanding biotic recoveries: extinction, survival, and preservation
 during the end-Permian mass extinction. Evolutionary paleobiology. University of Chicago
 Press, Chicago: 398–418
- Erwin DH (2001) Lessons from the past: biotic recoveries from mass extinctions. Proc Natl
 Acad Sci USA 98:5399–5403
- Ezaki Y, Liu J, Nagano T, Adachi N (2008) Geobiological aspects of the earliest Triassic
 microbialites along the southern periphery of the tropical Yangtze Platform: initiation and
 cessation of a microbial regime. Palaios 23:356–369
- Ezaki Y, Liu JB, Adachi N (2012) Lower Triassic stromatolites in Luodian County, Guizhou
 Province, South China: evidence for the protracted devastation of the marine
 environments. Geobiol 10:48–59
- Fang Y, Chen ZQ, Kershaw S, Li Y, Luo M (2017) An Early Triassic (Smithian) stromatolite
 associated with giant ooid banks from Lichuan (Hubei Province), South China:
 Environment and controls on its formation. Palaeogeogr Palaeoclimatol Palaeoecol in press
- 639 Flügel E (2002) Triassic reef patterns. Phanerozoic Reef Patterns. SEPM: 391–463
- 640 Forel MB, Crasquin S, Kershaw S, Collin PY (2013) In the aftermath of the end-Permian
- 641 extinction: the microbialite refuge? Terra Nova 25:137–143
- 642 Foster WJ, Twitchett RJ (2014) Functional diversity of marine ecosystems after the Late
- 643 Permian mass extinction event. Nature Geosci 7:233–238

- Foster WJ, Danise S, Sedlacek A, Price GD, Hips K, Twitchett RJ (2015) Environmental
 controls on the post-Permian recovery of benthic, tropical marine ecosystems in western
 Palaeotethys (Aggtelek Karst, Hungary). Palaeogeogr Palaeoclimatol Palaeoecol 440:374–
 394
- Foster WJ, Danise S, Price GD, Twitchett RJ (2017) Subsequent biotic crises delayed marine
 recovery following the late Permian mass extinction event in northern Italy. PloS One 12:
 e0172321
- Frey RW, Seilacher A (1980) Uniformity in marine invertebrate ichnology. Lethaia 13:183–
 207
- Frey RW, Pemberton SG (1984) Trace Fossil Facies Models. In: Walker RG (ed) Facies
 Models, 2nd edn. Geosci Canada, Reprint Series 1(41), pp 223–237
- Frey RW, & Pemberton, S. G. (1985). Biogenic structures in outcrops and cores. I.
 Approaches to ichnology. Bull Can Pet Geol 33:72–115
- Friesenbichler E, Richoz S, Baud A, Krystyn L, Sahakyan L, Vardanyan S, Peckmann J,
 Reitner J, Heindel K (2018) Sponge-microbial build-ups from the lowermost Triassic
 Chanakhchi section in southern Armenia: Microfacies and stable carbon isotopes.
 Palaeogeogr Palaeoclimatol Palaeoecol 90:653–672
- Fu W, Jiang DY, Montañez IP, Meyers SR, Motani R, Tintori A (2016) Eccentricity and
 obliquity paced carbon cycling in the Early Triassic and implications for post-extinction
 ecosystem recovery. Sci rep 6:27793
- Galfetti T, Bucher H, Ovtcharova M, Schaltegger U, Brayard A, Brühwiler T, Goudemand N,
 Weissert H, Hochuli PA, Cordey F, Guodun K (2007) Timing of the Early Triassic carbon
 cycle perturbations inferred from new U–Pb ages and ammonoid biochronozones. Earth
 Planet Sci Lett 258:593–604
- 668 Gall JC (1990) Les voiles microbiens. Leur contribution a la fossilisation des organismes au
 669 corps mou. Lethaia 23:21–28
- Goodspeed TH, Lucas SG (2007) Stratigraphy, sedimentology, and sequence stratigraphy of
 the Lower Triassic Sinbad Formation, San Rafael Swell, Utah. N M Mus Nat Hist Sci Bull
- 672 40:91–101
- 673 Grasby SE, Beauchamp B, Embry A, Sanei H (2013) Recurrent Early Triassic ocean anoxia.
 674 Geology 4:175–178
- 675 Gregory HE (1950) Geology and geography of the Zion [National] Park region, Utah and 676 Arizona. U S Geol Surv Prof Pap 220:1–200

- Grice K, Cao C, Love GD, Böttcher ME, Twitchett RJ, Grosjean E, Summons RE, Turgeon
 SC, Dunning W, Jin Y (2005) Photic zone euxinia during the Permian-Triassic
 superanoxic event. Science 307:706–709
- Grosjean AS, Vennin E, Olivier N, Caravaca G, Thomazo C, Fara E, Escarguel G, Bylund
 KG, Jenks JF, Stephen DA, Brayard A (2018) Early Triassic environmental dynamics and
 microbial development during the Smithian–Spathian transition (Lower Weber Canyon,
 Utah, USA). Sediment Geol 363:136–151
- Haig DW, Martin SK, Mory AJ, McLoughlin S, Backhouse J, Berrell RW, Kear BP, Hall R,
- 685 Foster CB, Shi GR, Bevan JC (2015) Early Triassic (early Olenekian) life in the interior of
- East Gondwana: mixed marine-terrestrial biota from the Kockatea Shale, Western
 Australia. Palaeogeogr Palaeoclimatol Palaeoecol 417:511–533
- Hakes WG (1976) Trace fossils and depositional environment of four clastic units, Upper
 Pennsylvanian megacyclothems, northeast Kansas. Univ Kans Paleontol Contrib Artic
 63:46 p
- Hautmann M, Bucher H, Brühwiler T, Goudemand N, Kaim A, Nützel A (2011) An
 unusually diverse mollusc fauna from the earliest Triassic of South China and its
 implications for benthic recovery after the end-Permian biotic crisis. Geobios 44:71–85
- Hautmann M, Smith AB, McGowan AJ, Bucher H (2013) Bivalves from the Olenekian (Early
 Triassic) of south-western Utah: systematics and evolutionary significance. J Syst
 Palaeontol 11:263–293
- Hautmann M, Bagherpour B, Brosse M, Frisk Å, Hofmann R, Baud A, Nützel A, Goudemand
 N, Bucher H (2015) Competition in slow motion: the unusual case of benthic marine
 communities in the wake of the end-Permian mass extinction. Palaeontol 58:871–901
- Hayden JM (2004) Geologic map of The Divide quadrangle, Washington County, Utah, Utah
 Geological Survey Map 197, scale 1:24,000: 32 p
- Hofmann R, Goudemand N, Wasmer M, Bucher H, Hautmann M (2011) New trace fossil
 evidence for an early recovery signal in the aftermath of the end-Permian mass extinction.
 Palaeogeogr Palaeoclimatol Palaeoecol 310:216–226
- Hofmann R, Hautmann M, Bucher, H (2013a) A new paleoecological look at the Dinwoody
 Formation (Lower Triassic, western USA): intrinsic versus extrinsic controls on ecosystem
 recovery after the end-Permian mass extinction. J Paleontol 87:854–880
- Hofmann R, Hautmann M, Wasmer M, Bucher H (2013b) Palaeoecology of the Spathian
 Virgin Formation (Utah, USA) and its implications for the Early Triassic recovery. Acta
 Palaeontol Pol 58:149–173

- Hofmann R, Hautmann M, Brayard A, Nützel A, Bylund KG, Jenks JF, Vennin E, Olivier N,
 Bucher H (2014) Recovery of benthic marine communities from the end–Permian mass
 extinction at the low latitudes of eastern Panthalassa. Palaeontol 57:547–589
- Holmer LE, Popov LE, Klishevich I, Ghobadi Pour M (2016) Reassessment of the early
 Triassic lingulid brachiopod '*Lingula' borealis* Bittner, 1899 and related problems of
 lingulid taxonomy. GFF, doi:10.1080/11035897.2016.1149216
- 717 Jattiot R, Bucher H, Brayard A, Monnet C, Jenks JF, Hautmann M (2016) Revision of the
- genus *Anasibirites* Mojsisovics (Ammonoidea): an iconic and cosmopolitan taxon of the
 late Smithian (Early Triassic) extinction. Pap Palaeontol 2:155–188
- Jattiot R, Bucher H, Brayard A, Brosse M, Jenks J, Bylund KG (2017) Smithian ammonoid
 faunas from northeastern Nevada: implications for Early Triassic biostratigraphy and
 correlation within the western USA basin. Palaeontogr Abt A 309: 1–89
- Kaim A, Nützel A, Bucher H, Brühwiler T, Goudemand N (2010) Early Triassic (Late
 Griesbachian) gastropods from South China (Shanggan, Guangxi). Swiss J Geosci
 103:121–128
- Kershaw S (2017) Palaeogeographic variation in the Permian–Triassic boundary
 microbialites: A discussion of microbial and ocean processes after the end-Permian mass
 extinction. J Palaeogeogr 6:97–107
- Kershaw S, Zhang T, Lan G (1999) A ?microbialite carbonate crust at the Permian–Triassic
 boundary in South China, and its palaeoenvironmental significance. Palaeogeogr
 Palaeoclimatol Palaeoecol 146:1–18
- Kershaw S, Crasquin S, Li Y, Collin PY, Forel MB, Mu X, Baud A, Wang Y, Xie S, Maurer
 F, Guo L (2012) Microbialites and global environmental change across the Permian–
 Triassic boundary: a synthesis. Geobiol 10:25–47
- Knoll AH, Bambach RK, Payne JL, Pruss S, Fischer WW (2007) Paleophysiology and endPermian mass extinction. Earth Planet Sci Lett 256:295–313
- 737 Lehrmann DJ (1999) Early Triassic calcimicrobial mounds and biostromes of the
 738 Nanpanjiang basin, south China. Geology 27:359--362
- 739 Lehrmann DJ, Wan Y, Wei J, Yu Y, Xiao J (2001) Lower Triassic peritidal cyclic limestone:
- an example of anachronistic carbonate facies from the Great Bank of Guizhou,
 Nanpanjiang Basin, Guizhou province, South China. Palaeogeogr Palaeoclimatol
 Palaeoecol 173:103–123
 - 22

- Lehrmann DJ, Ramezani J, Bowring SA, Martin MW, Montgomery P, Enos P, Payne JL,
 Orchard MJ, Wang H, Wei J (2006) Timing of recovery from the end-Permian extinction:
 Geochronologic and biostratigraphic constraints from south China. Geology 34:1053–1056
- 715 Geochionologie and biostratigraphic constraints from south china. Geology 54.1055 1050
- 746 Lehrmann DJ, Bentz JM, Wood T, Goers A, Dhillon R, Akin S, Li X, Payne JL, Kelley BM,
- 747 Meyer KM, Schaal EK, Suarez MB, Yu M, Qin Y, Li R, Minzoni M, Henderson CM
- 748 (2015) Environmental controls on the genesis of marine microbialites and dissolution
- surface associated with the end-Permian mass extinction: new sections and observations
 from the Nanpanjiang Basin, South China. Palaios 30:529–552
- Lucas SG, Krainer K, Milner AR (2007a) The type section and age of the Timpoweap
 Member and stratigraphic nomenclature of the Triassic Moenkopi Group in Southwestern
 Utah. N M Mus Nat Hist Sci Bull 40:109–117
- Lucas SG, Goodspeed TH, Estep JW (2007b) Ammonoid biostratigraphy of the Lower
 Triassic Sinbad Formation, East-Central Utah. N M Mus Nat Hist Sci Bull 40:103–108
- MacEachern JA, Pemberton SG, Gingras MK, Bann KL (2007) The Ichnofacies ParadigmChapter 4: A Fifty-Year Retrospective
- Marenco PJ, Griffin JM, Fraiser ML, Clapham M E (2012) Paleoecology and geochemistry of
 Early Triassic (Spathian) microbial mounds and implications for anoxia following the end Permian mass extinction. Geology 40:715–718
- Mary M, Woods AD (2008) Stromatolites of the Lower Triassic Union Wash Formation, CA:
 evidence for continued post-extinction environmental stress in western North America
 through the Spathian. Palaeogeogr Palaeoclimatol Palaeoecol 261:78–86
- Mata SA, Bottjer DJ (2011) Origin of Lower Triassic microbialites in mixed carbonate siliciclastic successions: ichnology, applied stratigraphy, and the end-Permian mass
 extinction. Palaeogeogr Palaeoclimatol Palaeoecol 300:158–178
- Mata SA, Bottjer DJ (2012) Microbes and mass extinctions: paleoenvironmental distribution
 of microbialites during times of biotic crisis. Geobiol 10:3–24
- McGowan AJ, Smith AB, Taylor PD (2009) Faunal diversity, heterogeneity and body size in
 the Early Triassic: testing post-extinction paradigms in the Virgin Limestone of Utah,
 USA. Aust J Earth Sci 56:859–872
- Nielson RL (1991) Petrology, sedimentology and stratigraphic implications of the Rock
 Canyon Conglomerate, southwestern Utah. Utah Geol Surv Misc Publ:91–7
- Nielson RL, Johnson JL (1979) The Timpoweap Member of the Moenkopi Formation.
 Timpoweap Canyon, Utah: Utah Geol 6:17–27

- Olivier N, Brayard A, Fara E, Bylund KG, Jenks JF, Vennin E, Stephen DA, Escarguel G
 (2014) Smithian shoreline migrations and depositional settings in Timpoweap Canyon
 (Early Triassic, Utah, USA). Geol Mag 151:938–955
- Olivier N, Brayard A, Vennin E, Escarguel G, Fara E, Bylund KG, Jenks JF, Caravaca G,
 Stephen DA (2016) Evolution of depositional settings in the Torrey area during the
 Smithian (Early Triassic, Utah, USA) and their significance for the biotic recovery. Geol J
 51: 600–626
- Paull RA, Paull RK (1993) Interpretation of Early Triassic nonmarine–marine relations, Utah,
 U.S.A. N M Mus Nat Hist Sci Bull 3:403–409
- Paull RK, Paull RA (1994) Shallow marine sedimentary facies in the earliest Triassic
 (Griesbachian) Cordilleran miogeocline, USA. Sediment Geol 93:181–191
- Paull RK, Paull RA (1997) Transgressive conodont faunas of the early Triassic: an
 opportunity for correlation in the Tethys and the circum-Pacific. In: Late Palaeozoic and
- Early Mesozoic Circum-Pacific Events and their Global Correlation, Dickins JM, Zunyi Y,
- Hongfu Y, Lucas SG, Acharyya SK (eds). Cambridge University Press, World and
 Regional Geology 10:158–167
- Payne JL, Clapham ME (2012) End-Permian mass extinction in the oceans: an ancient analog
 for the twenty-first century? Annu Rev Earth Planet Sci 40:89–111
- Payne JL, Lehrmann DJ, Wei J, Orchard MJ, Schrag DP, Knoll AH (2004) Large
 perturbations of the carbon cycle during recovery from the end-Permian extinction.
 Science 305:506–509
- Payne JL, Lehrmann DJ, Wei J, Knoll AH (2006) The pattern and timing of biotic recovery
 from the end-Permian extinction on the Great Bank of Guizhou, Guizhou Province, China.
 Palaios 21:63–85
- Payne, J. L., Turchyn, A. V., Paytan A, DePaolo DJ, Lehrmann DJ, Yu M, Wei J (2010)
 Calcium isotope constraints on the end-Permian mass extinction. Proc Natl Acad Sci 107:8543–8548
- Peng Y, Shi GR, Gao Y, He W, Shen S (2007) How and why did the Lingulidae
 (Brachiopoda) not only survive the end-Permian mass extinction but also thrive in its
 aftermath? Palaeogeogr Palaeoclimatol Palaeoecol 252:118–131
- 806 Pietsch C, Bottjer DJ (2014) The importance of oxygen for the disparate recovery patterns of
- the benthic macrofauna in the Early Triassic. Earth Sci Rev 137:65–84

- Pietsch C, Mata SA, Bottjer DJ (2014) High temperature and low oxygen perturbations drive
 contrasting benthic recovery dynamics following the end-Permian mass extinction.
 Palaeogeogr Palaeoclimatol Palaeoecol 399:98–113
- 811 Pörtner HO, Langenbuch M, Michaelidis B (2005) Synergistic effects of temperature
 812 extremes, hypoxia, and increases in CO2 on marine animals: From Earth history to global
 813 change. J Geophys Res: Oceans, 110(C9)
- 814 Posenato R, Holmer LE, Prinoth H (2014) Adaptive strategies and environmental significance
- of lingulid brachiopods across the late Permian extinction. Palaeogeogr Palaeoclimatol
 Palaeoecol 399:373–384
- Pruss SB, Bottjer DJ (2004) Late Early Triassic microbial reefs on the western United States:
 a description and model for their deposition in the aftermath of the end-Permian mass
 extinction. Palaeogeogr Palaeoclimatol Palaeoecol 211:127–137
- 820 Pruss SB, Bottjer DJ, Corsetti FA, Baud A (2006) A global marine sedimentary response to
- the end-Permian mass extinction: Examples from southern Turkey and the western United
 States. Earth Sci Rev 78:193–206
- Reeside Jr JB, Bassler H (1922) Stratigraphic sections in southwestern Utah and northwestern
 Arizona. US Geol Surv Prof Papr 129(D): 53–77
- Ritter S, Osborn C, Goodrich C (2013) Sedimentology and reservoir characteristics of the
 Lower Triassic (Smithian) Sinbad Limestone Member of the Moenkopi Formation, San
 Rafael Swell, Utah. In: Morris TH, Ressetar R (eds) The San Rafael Swell and Henry
- 828 Mountains Basin—Geologic Centerpiece of Utah. Utah Geol Assoc Publ 42:199–222
- Rodland DL, Bottjer DJ (2001) Biotic recovery from the end-Permian mass extinction:
 behavior of the inarticulate brachiopod Lingula as a disaster taxon. Palaios 16:95–101
- 831 Schubert JK, Bottjer DJ (1992) Early Triassic stromatolites as post-mass extinction disaster
 832 forms. Geology 20:883–886
- Schubert JK, Bottjer DJ (1995) Aftermath of the Permian-Triassic mass extinction event:
 paleoecology of Lower Triassic carbonates in the western USA. Palaeogeogr
 Palaeoclimatol Palaeoecol 116:1–39
- 836 Seilacher A (1963) Lebensspuren und Salinitätsfazies. Fortschr Geol Rheinl Westfal 10:81–
 837 94
- 838 Song H, Wignall PB, Chu D, Tong J, Sun Y, Song H, He W, Tian L (2014) Anoxia/high
- temperature double whammy during the Permian-Triassic marine crisis and its aftermath.
- 840 Sci rep4:4132

- Stewart JH, Poole FG, Wilson RF (1972) Stratigraphy and origin of the Triassic Moenkopi
 Formation and related strata in the Colorado Plateau region: U. S. Geol Surv Prof Pap
 691:195 p
- Sun Y, Joachimski MM, Wignall PB, Yan C, Chen Y, Jiang H, Wang L, Lai X (2012)
 Lethally hot temperatures during the Early Triassic greenhouse. Science 338:366–370
- 846 Szmuc EJ, Osgood RG, Meinke DW (1977) Synonymy of the ichnogenus Lingulichnites
- 847 Szmuc, E.J., Osgood, R.G., and Meinke, D.W. 1976, with Lingulichnus Hakes, 1976.
 848 Lethaia 10:106
- Tang H, Kershaw S, Liu H, Tan X, Li F, Hu G, Huang C, Wang L, Lian C, Li L, Yang X
 (2017) Permian–Triassic boundary microbialites (PTBMs) in southwest China:
 implications for paleoenvironment reconstruction. Facies 63:2
- 852 Thomazo C, Vennin E, Brayard A, Bour I, Mathieu O, Elmeknassi S, Olivier N, Escarguel G,
 853 Bylund KG, Jenks J, Stephen DA, Fara E (2016) A diagenetic control on the Early Triassic
 854 Smithian–Spathian carbon isotopic excursions recorded in the marine settings of the
- Smithian–Spathian carbon isotopic excursions recorded in the marine settings of the
 Thaynes Group (Utah, USA). Geobiol 14:220–236 DOI: 10.1111/gbi.12174
- Tomescu AM, Klymiuk AA, Matsunaga KK, Bippus AC, Shelton GW (2016) Microbes and
 the Fossil Record: Selected Topics in Paleomicrobiology. In: Their World: A Diversity of
 Microbial Environments (pp. 69-169). Springer International Publishing
- Twitchett RJ, Krystyn L, Baud A, Wheeley JR, Richoz S (2004) Rapid marine recovery after
 the end-Permian mass-extinction event in the absence of marine anoxia. Geology 32:805–
 861 808
- Vennin E, Olivier N, Brayard A, Bour I, Thomazo C, Escarguel G, Fara E, Bylund KG, Jenks
 JF, Stephen DA, Hofmann R (2015) Microbial deposits in the aftermath of the endPermian mass extinction: a diverging case from the Mineral Mountains (Utah, USA).
 Sedimentol 62:753–792
- Wignall PB, Twitchett RJ (1996) Oceanic anoxia and the end Permian mass extinction.
 Science 272:1155
- Woods AD (2014) Assessing Early Triassic paleoceanographic conditions via unusual
 sedimentary fabrics and features. Earth Sci Rev 137:6–18
- 870 Yang H, Chen ZQ, Wang Y, Tong J, Song H, Chen J (2011) Composition and structure of
- 871 microbialite ecosystems following the end-Permian mass extinction in South China.
- 872 Palaeogeogr Palaeoclimatol Palaeoecol 308:111–128

873	Yang H, Chen ZQ, Wang Y, Ou W, Liao W, Mei X. (2015) Palaeoecology of microconchids
874	from microbialites near the Permian-Triassic boundary in South China. Lethaia 48:497-
875	508
876	Zonneveld JP, Pemberton SG (2003) Ichnotaxonomy and behavioral implications of
877	lingulide-derived trace fossils from the Lower and Middle Triassic of Western Canada.
878	Ichnos 10:25–39
879	Zonneveld JP, Beatty TW, Pemberton SG (2007) Lingulide brachiopods and the trace fossil
880	Lingulichnus from the Triassic of western Canada: implications for faunal recovery after
881	the end-Permian mass extinction. Palaios 22:74–97
882	
883	
884	
885	
886	
887	
888	Figure captions
889	
890	
891	Fig. 1 a Present-day location of the study area in southwestern Utah, along the Hurricane
892	Cliffs, about 8 km south of Hurricane city. b Early Triassic paleogeographic map showing the
893	location of southwestern USA along the Pangea margin (after Brayard et al. 2013)
894	
895	Fig. 2 a Field view of section #1 on the western side of the Hurricane Cliffs (see geographic
896	location on Fig. 1a) showing the Permian-Triassic transition. The studied outcrop is about 70
897	m thick. b Field view of the eastern side of the Hurricane Cliffs, just south of the Three
898	Brothers Tertiary volcanic peaks. Sections #2-5 were logged along the head end of Workman
899	Wash. Geologists near section #5 for scale
900	
901	Fig. 3 Legend for Figures 4-8 and 15
902	
903	Fig. 4 Log of the studied interval on the western side of the Hurricane Cliffs. See location on
904	Figures 1a and 2, Table 1 for facies descriptions, and Figure 3 for legend
905	

Fig. 5 Log of section #2. See location on Figures 1a and 2b, Table 1 for facies descriptions,and Figure 3 for legend

908

Fig. 6 Log of section #3. See location on Figures 1a and 2b, Table 1 for facies descriptions,and Figure 3 for legend

911

Fig. 7 Log of section #4. See location on Figures 1a and 2b, Table 1 for facies descriptions,and Figure 3 for legend

914

Fig. 8 Log of section #5. See location on Figures 1a and 2b, Table 1 for facies descriptions,and Figure 3 for legend

917

918 Fig. 9 Field illustrations of some facies and macroorganisms. a Fenestral-microbial facies 919 (F7). The lower part of the picture shows a macroscopic laminated microbial structure 920 whereas the upper part displays a fenestral fabric (subfacies F7b; section #1, about 52 m). **b** 921 Fenestral-microbial limestones (F7) with large fenestrae and endostromatolites (subfacies 922 F7b; section #2, about 4 m). c Accumulation of large-sized gastropods (subfacies F7b; section 923 #4, about 3 m). d Accumulation of ammonoids (subfacies F7c; section #5, about 6 m). e 924 Intensively bioturbated interval corresponding to the marker bed with lingulid brachiopods 925 (Facies F11; section #4, about 4.5 m). f Conglomerate breccia (Facies F1; base of section #4). 926 g Yellow clastic unit (Facies F5; section #4, about 15 m)

927

Fig. 10 a Field view and b cartoon of the fenestral-microbial facies (F7) locally showing
reefal morphology and truncation surfaces above the lingulid marker bed

930

931 Fig. 11 Natural-light thin-section photographs of the fenestral-microbial facies (F7). a Oo-932 pisoid subfacies F7a with pisoids (sometimes truncated), ooids and peloids in a sparitic 933 cement. **b** Intraclastic subfacies F7e with abundant intraclasts in a sparitic cement. **c-f** 934 Fenestral-limestone subfacies F7b with various types of endostromatolites. c Thin crusts of 935 dense micrite with downward or upward growth directions at the roof and the floor of a 936 fenestrae. d Coalescent stromatolitic columns with a marked lamination, which locally displays some putative residual calcified sheaths of filamentous bacteria (arrows). e Complex 937 938 crust made of an alternation of sparitic and micritic laminae. f Acicular to botryoidal cements 939 associated with a lamination of an endostromatolites in a large fenestrae. g Subfacies F7c with

- an interval made of domal to columnar stromatolites, topped by a lamellar siliceous sponge. h
 Lamellar siliceous sponge (subfacies F7c)
- 942

Fig. 12 Natural-light thin-section photographs of the fenestral-microbial facies (F7). a Root
structures and circum-granular cracks, subfacies F7c. b Subfacies F7d with small fenestrae
and abundant extraclasts. c Biofilm (white arrow) coating a pel-fenestral structure, subfacies
F7c. d Biofilms coating a peloidal grainstone above an oncoidal-fenestral structure, subfacies
F7c. e Biofilms coating a peloidal grainstone above a micritic stromatolite, subfacies F7c. f A
stromatolitic crust displaying a columnar microfabric, subfacies F7c. g Planar to wavy
structure of a stromatolite, subfacies F7c. h Enigmatic cemented crust, subfacies F7c

- Fig. 13 Natural-light thin-section photographs of the bioclastic facies. a Spicule-echinoderm
 limestone (F9). Note the common echinoderm fragments (white arrows) and the darker (more
 micritic) areas reflecting a probable infaunal activity. b Oo-pel-bioclastic limestone (F10). c
 Pel-bioclastic limestone (F12). d Intra-bioclastic limestone (F13)
- 955

Fig. 14 Lingulid brachiopods in Facies F11, section #4, about 8 m (see Figure 2b and 7). a Insitu large complete shell of *Lingularia* in life position characterizing the marker bed attributed to facies F11 at Workman Wash. b Typical example of *Lingulichnus verticalis* associated to lingulid shells in facies F11. Note the pedicle trace and the set of downward-deflected laminae produced by upward adjustment of the lingulid in response to sediment aggradation (equilibration structure). Scale bar (valid for both photographs) is 5 mm

- 962
- 963 Fig. 15 Correlation of sections. See figures 1 and 2 for their location
- 964
- Fig. 16 Schematic 3D reconstruction of the depositional setting for the late Smithiansedimentary rocks observed at Workman Wash
- 967
- 968 **Table 1** Facies description and depositional environments





Workman Wash

Lithologies Limestones Coarse dolomitic limestones a Clays s Silts Cg Conglomerates	Sedimentary grains•Peloids•Ooids•Cortoids•Oncoids•Pisoids•Aggregates•Intraclasts	Sedimentary structures v Bioturbation v Fenestrae v Keystone vugs v Mud cracks v Cross-bedding v Asymmetric ripples	 Biota Ammonoids Bivalves (undifferentied) ↓ Lingulid brachiopods ★ Echinoderms ③ Gastropods ♦ Ostracods
B Boundstone R Rudstone F Floatstone G Grainstone Mic P Packstone W Wackestone M Mudstone	 Extraclasts Mud clasts <i>trobial structures</i> Large fenestrae with endostromatolites Microbial laminae (biofilms) 	 Gypsum crytais Acicular crytais Karstification Cherty nodules Erosional surface 	 Phosphate remains Serpulids Siliceous sponges Sponge spicules





Figure04suite































Facies		Textures	Biotic/non biotic grains	Matrix/ cements	Sedimentary structures	Zones	Environments			
F1	Co	Conglomerate			Polygenic clasts (cherts, bioclastic limestones, red beds), with diameter up to 15 cm (commonly elongated). Poorly sorted and angular to subrounded clasts.	Silty matrix	Stacked and amalgamated channel complexes (with individual channel up to 2 m large and 1,3 m thick). Locally, some m-scale isolated channels.		Chennalized inner tidal flat	
F2	2 Dolosiltstone			Common peloids. Abundant well sorted and subangular to subrounded quartz grains.	Dolomitized micritic matrix	Trough cross bedding and asymmetrical ripples. Mud drapes. Common bioturbation.				
F3	3 Dolomudstone		М	Dolomitized Trough cross bedding with low angle f Asymmetrical ripples. Common biotur		Trough cross bedding with low angle foresets. Asymmetrical ripples. Common bioturbation.	Peritidal	Inner		
F4	-4 Dolostone				Dolomitic cement		(oublidal)	tidal flat		
F5	Si lin	ilty nes	pelo ston	oidal ne	Р	Frequent to rare bivalves. Rare gastropods. Abundant peloids. Frequent to common micritized ooids. Common cortoids and intraclasts. Rare oncoids. Rounded to subrounded mudclasts.	 gastropods. Abundant peloids. J ooids. Common cortoids and nded to subrounded mudclasts. Dolomitized micritic matrix Dolomitized micritic matrix Trough cross bedding with low angle fore sets. Mud drapes and mud clasts. Rare extraclasts. Surimposed asymmetrical ripples. Common to rare bioturbation. 			
F6	Ke lin	eys nes	ston ston	ie vug ie	P/W	Frequent peloids, common micritic and microbial (reworked?) clasts.	Dolomitized micritic matrix	Frequent keystone vugs. Rare bioturbation.	Swash	-
Education	stone	F7	7a	Oo-pisoid imestone	R(G/P) /B	Abundant ooids. Abundant to common pisoids and intraclasts. Frequent to common peloids. Rare (fragmented) gastropods and ostracods.	Micritic matrix/ sparitic cement	Abundant small fenestra. Some large fenestrae with endostromatolites. Locally, some vadose silt. Sparse microbial laminae.		
	bial limes	F7	7b	Fenestral- limestone	P(G,W)/ F/B(M)	Frequent to rare ostracods. Locally, frequent ammonoids. Abundant to rare gastropods. Frequent to rare bivalves. Rare echinoderms. Abundant to frequent peloids. Abundant to rare ooids. Common oncoids and intraclasts. Common to rare cortoids and pisoids.	Sparitic cement/ micritic matrix/ acicular crystals	Abundant small fenestrae. Locally large fenestrae (with upward and downward growth stromatolitic crusts). Mud cracks and vadose silt. Common microbial laminae. Rare bioturbation (?).	Peritidal	
	micro	F7	7c	Sponge-micro- bial limestone	В	Common ostracods and sponges. Rare bivalves.	Micritic laminae. Acicular crystals	Stromatolites (micritic lamination). Locally, some root structures and circum-granular cracks.	(supratidal to subtidal)	1
	estral-	F7	7d	Conglomerate imestone	B/P-M	Frequent to rare gastropods and ostracods. Rare bivalves. Frequent to rare peloids and ooids. Rare intraclasts and oncoids. Abundant extraclasts (polygenic, poorly sorted, elongated up to 7 cm long).	Micritic matrix	Abundant thin, elongated and aligned fenestrae or large fenestrae with endostromatolites. Sparse microbial laminae.		
	Fer	F7	7e	Intraclastic imestone	G/B	Abundant intraclasts (fragmented pisoidal, ooidal, and peloidal clasts). Rare bivalves, ostracods, and gastropods.	Sparitic cement	Frequent fenestrae. Rare microbial laminae (and bioturbation?).		Interior
F8	Ec dc	chir olor	noderm W Frequent echinoderms. Common to rare bivalves. Rare Dolomitized micritic matrix Sparse bioturbation.		Sparse bioturbation.		platform			
F9	Spicule- echinoderm limestone		P(W)	Abundant echinoderms. Frequent sponge spicules, bivalves, and gastropods. Common ostracods. Rare phosphate remains. Common peloids.	Micritic matrix	Abundant bioturbation.				
F10	0 Oo-pel-bioclastic limestone		Р	Abundant gastropods. Frequent bivalves. Common echinoderms. Abundant ooids. Common peloids. Rare intraclasts	Micritic matrix	Trough cross bedding with low angle foresets. Some mud drapes and mud clasts. Frequent bioturbation.	Subtidal			
F11	11 Bioturbated and bioclastic limestone		Р	Abundant to rare echinoderms. Abundant to frequent gastropods and bivalves. Common to rare ostracods. Frequent to rare phosphate remains. Some lingulid brachiopods. Frequent to rare peloids. Rare ooids.	Micritic matrix	Abundant bioturbation (<i>Arenicolites</i> , <i>Thalassinoides</i> , and <i>Paleoptycus</i>). Some small asymmetrical ripples.	(tide dominated)			
F12	12 Pel-bioclastic limestone		G/P	Frequent bivalves. Frequent to common gastropods. Frequent to rare echinoderms. Rare ostracods and phosphatic remains. Abundant peloids. Common to rare ooids. Rare intraclasts.	Micritic matrix/ sparitic cement	Trough cross bedding with low angle foresets. Small erosive channel with lateral accretion. Frequent to common bioturbation.				
F13	3 Intra-bioclastic limestone		G(P)	Frequent gastropods and bivalves. Rare echinoderms, ostracods, and phosphatic remains. Abundant peloids. Frequent intraclasts. Common ooids.	Sparitic cement	Trough cross bedding with low angle foresets.				